# Handedness in nature 

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#### Abstract

Nature manifests handedness at various levels in the animal as well as vegetable world. It expresses itself in the genetic material, proteins and polypeptides. The predominant conformation in which the DNA molecule exists is the right handed form known as B-DNA. The $A$ form is also right-handed. Since the discovery of left-handed $Z$ form of DNA, several questions have been raised about its role in the evolutionary process and metabolism. It is now well established that whereas the right handed form can accommodate any sequence of nucleotides. the $Z$ form is generally found in sequences having alternating purines and pyrimidines only, e.g. poly $(G-C)$.


In the present communication we report a study of the reversible transitions between $B, Z$ and other condensed states $\left[\psi(+)\right.$. $\left.\psi^{\prime}(-)\right]$ of DNA. This is based on the one dimensional Ising model propounded for ferromagnetic transition and modified suitably by introducing singular interaction between $1: 4$ neighbours (hydrogen bond formation) for the helix $\underset{\sim}{\boldsymbol{*}}$ coil transition observed in several proteins and polypeptides. Results for $B \geqslant Z \geqslant \psi(-) \geqslant$ coil transitions in poly (dG-dC) and poly(dG-me ${ }^{5}$ dC ) have been obtained. Results are consistent with the change in the environmental conditions e.g. change in the cationic concentration of the solvent and methylation of the cytosine residue. The nucleation parameter is consistent with the stabilization/destabilization of the ordered state under these conditions. Possible reasons for the preponderance of right handed DNA over left-handed are discussed.

Key-words-Symmetry, DNA, conformation, transition.

## INTRODUCTION

SYMMETRY is an integral part of nature and activity of man (Text-figs 1-6). Most of the natural macromolecules which constitute the vegetable world (polysaccharides, proteins, nucleic acids, etc.) have symmetry elements present in them. Man made molecules such as polyethylene, polypropylene, polytetrafluoroethylene, polypeptides, polyaminoacids, polynucleic acids etc. are also all symmetric. Whether it is a piece of architecture, painting or other creative activities, symmetry adds beauty, harmony and proportion to it. The crystals of snow flakes are an excellent example of the symmetry in nature (Text-fig. 1). However, symmetry conveys much more than these aesthetic attributes. It implies economy of design and underlines laws of conservation. The translational, rotational and mirror symmetry are manifestations of conservation of linear momentum, angular momentum and parity respectively. These conservation laws, in turn, depend on the nature of interactions. For example, if the frictional forces are present, momentum and energy are not conserved.

The abundance of symmetry in the vegetable world is evident in leaves, flowers, fruits and many a time even
in the expansion or growth of a plant. Handedness which could be right or left handed is only one of the symmetry elements. It is not merely an accident or coincidence that proteins, nucleic acids and enzymes are all right handed. It means that only the L -amino acids can combine to give a conformation which is biologically active. The way many creepers wind themselves around the support and the dangling tandrils all have a helical symmetry.

The symmetry of the organic world is entirely different from that of the inorganic world. In an aging process it is observed that left-handed residues enter in a sequence of right handed residues. Too many of such odd residues can result in death. Handedness is therefore characteristic of life. In fact, in some ways search for extraterrestrial life is based on the search for biomolecules which go into right-handed conformation. No such importance is attached to symmetry in inorganic molecules.

The preponderance of right-handed over left-handed DNA raises many important questions from the view point of natural laws of nature. What are the forces which are responsible for this? Some comments on this would be made towards the end of this paper.

Since the uncoiling of a DNA molecule (melting) is closely related to the translation and transcription of


Text-figure 1. Crystalline forms of snow flakes. (Courtesy of the U.S. Weather Bureau: Photo by W.A. Bently.).
genetic information, it has been the subject of extensive experimental and theoretical studies (Mc Campbell et al., 1989; Ramsing et al., 1989; Ya Azbel, 1980 a,b,c). This form of double stranded DNA is the right handed B-DNA which has a pitch of $34 \AA$ and 10 base pairs/turn. However, with the discovery of Z-DNA by Rich et al.


Text-figure 2. Tiger swallowtail butterfly. (Courtesy of the Field Museum of Natural History. Chicago, Illinois.).


Text-figure 3. Broad-banded copperhead snake. (Courtesy of the Field Museum of Natural History. Chicago. Illinois.).
(1984)such studies have received an additiona push. All spectroscopic (proton NMR, ORD. CD) and physicochemical studies such as X-ray diffraction, I.R., U.V. etc. (Behe \& Felsenfeld 1985; Chaires \& Sturtevant, 1986, 1988; Gulatto et al., 1989; atha \& Brahmchari 1985. 1986: Latha et al., 1983: Narasimhan \& Bryan 1984: Ramesh \& Brahmchari, 1983; Souche et al. 1985; Feigon et al., 1985; Zacharias et al, 1982) indicate that Z-DNA is a left-handed structure. It has a pitch of $44.6 \AA$ and 12 base pairs/turn. This conformation is chiefly observed in Poly ( $\mathrm{dG}-\mathrm{dC}$ ) under proper environmental conditions. Although Poly (dA-dT) does not exist in the $Z$ form, it is found that isolated A-T pairs can be accommodated in the sequence of $\mathrm{C}-\mathrm{G}$ pairs (Latha \& Brahmchari, 1986; Wang et al., 1985: Rich et al., 1984; Jovin et al., 1987). It may however be added here that in a recent report Adam et al. (1986 in Jovin et al., 1987) have reported existence of Z-conformation


Text-figure 4. Taj Mahal, Agra, India. (Coutesy of the Government of India Tourist Office, 201 N. Michigan Avc.. Chicago, Illinois.).


Text-figure 5. Banded butterfly fish (Courtesy of the Field Museum of Natural History. Chicago.).
in Poly (dA-dT) films containing $\mathrm{Ni}^{++}$and low water activity. Z-DNA has also been detected in fibres and films of DNA, poly- and oligo-mers, in supercoiled plasmids, and in chromosomes and cells. Z structure is unique in many ways. It is considered as variant of sugar modification and has been observed in double stranded RNA as well (Cruz et al., 1986 in Jovin et al., 1987). Further, it is observed that $\mathrm{d}(\mathrm{C}-\mathrm{G})_{\mathrm{n}}$ oligomers ( $\mathrm{n}=2,3$ and 4 ) crystallize in the left handed Z conformation but the corresponding $\mathrm{d}(\mathrm{G}-\mathrm{C})_{n}$ oligomers are not found to do so (Jovin et al., 1987). Thus the two sequence isomers must have different energetics.

Methylation of the cytosine residue at the C 5 position is found to bring about changes in the biological activity of the molecule, e.g., it is prevalent in eukaryotic and prokaryotic genome of viral DNA (Souche et al., 1985). Methylation of cytosine residue at C5 position is also


Text-figure 7. Theoretical transition-curves for poly ( $\mathrm{dG}, \mathrm{IC}^{\circ}$ ) :
(A) $B \rightleftharpoons Z$ and $Z \neq{ }^{\prime \prime}$
(B) I $\mathrm{P}(-) \geq$ coil


Text-figure 6. Flower symmetry
responsible for the destabilization of B structure (Latha \& Brahmchari, 1986; Souche et al., 1985: ehe \& Felsenfeld, 1985; Narsimhan \& Bryan, 1984; Chaires \& Sturtevant. 1986, 1988). This is mainly due to unfavourable interactions between the hydrophobic methyl group and water molecule. This destabilization of B helix would in turn, lead t the stabilization of Z conformation.

The study of Z-DNA is of both physicochemical and biological interest. The conformation is considered to be important in regulating gene activity, and it is an important intermediate in genetic recombination in a simple eukaryote, the yeast like fungus called Ustilago (Marx, 1985). It is quite possible that the preponderance of the right-handed over the left-handed DNA is related to the biological evolution process of the universe.

The $\mathrm{B} \gtrless \mathrm{Z}$ transition takes place as a result of rotation


Text-figure 8. Theoretical transition-curves for poly (dG-dC) :
( A$) \mathrm{B}$ き $\boldsymbol{M}(-)$
(B) $\Vdash(-) \neq$ coil
around the glycosidic bond resulting in syn－conformation and C3＇endo sugar pucker of purines．This combined with the rotation of cytidine residue in the anti－conforma－ tion gives rise to the characterisic zig－zag of the sugar phosphate backbone（Jovin et al．，1987；Rich et al．． 1984；Wang et al．，1979）．

The exact mechanism of $B \geqslant Z$ transition is not yet fully understood．However，it is definitely related to the lowering of water activity in presence of reagents like salts，alcohols，increasing temperature and in particular， presence of cations like $\mathrm{Na}^{+}$and $\mathrm{Mg}^{++}$．The cations play an important role in masking the phosphate repulsions． These stabilizing forces are further reinforced by modifica－ tion of water structure around the DNA molecule in the presence of salts．The stability／destability of the $B$ helix is inherent in the sugar phosphate backbone conforma－ tion．

Various theories have been advanced for explaining the $\mathrm{B} \rightleftharpoons \mathrm{Z}$ and $\mathrm{Z} き$ coil transitions．The later transition，in certain cases is preceded by another transition called $Z \geqslant$ $\psi(-)$ transition（Chaires \＆Sturtevant，1986，1988）．The $\psi(-)$ is the condensed state of the DNA with left－handed conformation and the geometry of B form．The $\psi(-)$ form is found in solution containing polyethylene glycol and a critical NaCl concentration（Chaires \＆Sturtevant，1988）． The counter $\psi(+)$ form is also reported to occur in sodium acetate solution or in the presence of Hexamine cobalt． It has a right－handed conformation but different geometry．

In the present communication we have made efforts to explain order $\geqslant$ order and order $\geqslant$ disorder transitions in a variety of conformationally permissible polynucleotide system．We have also tried to explain the specific heat measurements in the neighbourhood of transition．In brief，the theoretical approach is a modifica－ tion and development of the Zimm and Bragg（1959） theory of helix $\geqslant$ coil transition．In fact all theories which have been framed to explain phase transition in biomolecules are based on a two phase cooperative model first enunciated by Ising for a chain of linear spins to explain ferromagnetism（Agnihotri et al．，1981）．

## THEORY

The present theory is a variant of the Zimm and Bragg（1959）formalism enuciated for helix き coil transi－ tion．This has been earlier used to explain order ₹ order transition in case of Poly－L－Proline I き Poly－L－Proline II （right handed to left handed）transition（Agnihotri et al．， 1981）．In Brief if we assume $\sigma_{1}$ and $\sigma_{2}$ to be the nucleation parameters and s1 and s2 are the growth parameters in the states $h_{1}$ and $h_{2}$ which represent the right－handed and left－handed helices in orderきorder tran－ ，sition，e．g．，$B \rightleftharpoons Z$ transition then the transition matrix can
be written down as：

|  | $\mathrm{h}_{1}$ | $\mathrm{h}_{2}$ |
| :---: | :---: | :---: |
| $M=\begin{aligned} & h_{1} \\ & h_{2}\end{aligned}$ | $\mathrm{s}_{1}$ | $\mathrm{\sigma}_{2} \mathrm{~S}_{2}$ |
|  | $\sigma_{1} s_{1}$ | $\mathrm{s}_{2}$ |

Going through the matrix evaluation formalism out－ lined by Zimm and Bragg（1959），we obtain expression for the fraction of segments in the form I（say from B in $B \geqslant Z$ transition）：
$f_{1}=\frac{1}{N} \frac{\ln Z}{\ln s_{1}}=\frac{\frac{s_{1}}{\lambda_{1}} \frac{\partial \lambda_{1}}{\partial s_{1}}+B \frac{s_{1}}{\lambda_{2}} \frac{\partial \lambda_{2}}{\partial s_{1}}+\frac{1}{N}\left(\frac{s_{1}}{A_{1}} \frac{\partial A_{1}}{\partial s_{1}}+B \frac{s_{1}}{A_{2}} \frac{\partial A_{2}}{\partial s_{1}}\right)}{(1+B)}-$（2）
where Z is the partition function of N segment chain． $\lambda_{1}$ and $\lambda_{2}$ are eigen values of $M$ ．
$B=\left(\frac{X_{2}}{X} s u b\right)^{N}\left(\frac{A_{2}}{A_{1}}\right)$
with $X_{1}=s_{1} \lambda_{1}$ and $X_{2}=s_{1} \lambda_{2}$
$A_{1}=\frac{s_{1}+s_{2} x-b \lambda_{2}}{\lambda_{1}-\lambda_{2}}$
$A_{2}=\frac{s_{1}+s_{2} x-b \lambda_{1}}{\lambda_{2}-\lambda_{1}}$
$b=\frac{1+x-\sigma_{2}-\sigma_{1} x}{1-\sigma_{1} \sigma_{2}}$
and $\mathrm{x}=\sigma^{\prime \prime} / \sigma^{\prime}$
Where $\sigma^{\prime}$ and $\sigma^{\prime \prime}$ are the parameters giving the interaction of end segments with the solvent when they are in states $h_{1}$ and $h_{2}$ respectively．
$\frac{s_{1}}{\lambda_{1}} \frac{\delta \lambda_{1}}{\delta s_{1}}=\frac{X_{1}-1}{X_{1}-X_{2}}$
$\frac{s_{1}}{\lambda_{2}} \frac{\delta \lambda_{2}}{\delta s_{1}}=\frac{X_{2}-1}{X_{2}-X_{1}}$
$\frac{s_{1}}{A_{1}} \frac{\delta A_{1}}{\delta s_{1}}=\frac{s^{2}(2 \sigma-1+b-x-b \sigma)+s(1+x-b+b \sigma-2 \sigma x)}{\left[(1-s)^{2}+4 \sigma x\right)\left(s+x-b X_{1}\right)}$
$\frac{s_{1}}{A_{2}} \frac{\delta A_{2}}{\partial s_{1}}=\frac{s^{2}(2 \sigma-1+b-x-b \sigma)+s(1+x-b+b \sigma-2 \sigma x)}{\left[(1-s)^{2}+4 \sigma x\right]\left(s+x-b X_{2}\right)}$
where $\sigma=\sigma_{1} \sigma_{2}$ and $s=s_{1} / \mathrm{s}_{2}$
The extension of this formalism for specific heat is straightforward．The specific heat is related to the molar enthalpy and entropy changes in the transition from state I to state II．From well known thermodynamic relation free energy $F$ is given by
$F=-K T 1 n Z$
and the internal energy $U$ is given by
$U=-T^{2} \frac{\delta}{\delta T}\left(\frac{F}{T}\right)$
Differentiating the above equation with respect to temperature and making use of

$$
\begin{equation*}
s=\exp \left[\frac{\Delta H}{R}\left(\frac{1}{T}-\frac{1}{T_{f}}\right)\right] \tag{10}
\end{equation*}
$$

where $\Delta \mathrm{H}$ is the molar change in enthalpy about the transition point and $T_{f}$ is the transition temperature.

We get specific heat $C_{v}$ as

$$
\begin{equation*}
C_{v}=\left(\frac{\delta U}{\delta T}\right)_{V}=N k\left(\frac{\Delta H}{R T}\right)^{2}\left(s \frac{\delta f_{1}}{\delta s}\right) \tag{11}
\end{equation*}
$$

where

$$
\begin{align*}
& \frac{\delta f_{1}}{\delta s}=-\frac{f_{1}}{1+B} \frac{\delta B}{\delta s}+\frac{1}{(1+B)}\left[\frac{\delta}{\delta s}\left(\frac{s_{1}}{\lambda_{1}} \frac{\partial \lambda_{1}}{\delta s_{1}}\right)+\frac{1}{N} \frac{\partial}{\delta s}\left(\frac{s_{1}}{A_{1}} \frac{\partial A_{1}}{\delta s_{1}}\right)\right. \\
& +\frac{\delta B}{\delta s}\left(\frac{s_{1}}{\lambda_{2}} \frac{d l t a \lambda_{2}}{\delta s_{1}}+\frac{1}{N} \frac{s_{1}}{A_{2}} \frac{\delta A_{2}}{\delta s_{1}}\right)+B\left\{\frac { \delta } { \partial s } \left(\frac{s_{1}}{\lambda_{2}} \frac{\left.\frac{\partial \lambda_{2}}{\partial s_{1}}\right)}{\left.\left.\quad+\frac{1}{N} \frac{\delta}{\delta_{s}}\left(\frac{s_{1}}{A_{2}} \frac{\delta A_{2}}{\delta s_{1}}\right)\right\}\right]}\right.\right. \\
& \quad \text { with } \tag{12}
\end{align*}
$$

$\frac{\delta B}{\delta s}=N B\left[\frac{1}{X_{2}} \frac{\delta X_{2}}{\delta s}-\frac{1}{X_{1}} \frac{\delta X_{1}}{\delta s}\right]$
$+B\left[\frac{1-b \frac{\delta X_{1}}{\delta s}}{\left(s+x-b X_{1}\right)}-\frac{1-b \frac{\delta X_{2}}{\delta s}}{\left(s+x-b X_{2}\right)}\right]$
$\frac{\delta X_{1}}{\delta s}=\frac{1}{2}+\frac{[(s-1)+2 \sigma]}{2\left[(s-1)^{2}+4 \sigma s\right]^{12}}$.
$\frac{\delta X_{2}}{\delta s}=\frac{1}{2}-\frac{[(s-1)+2 \sigma]}{2\left[(s-1)^{2}+4 \sigma s\right]^{12}}$
$\frac{\delta}{\delta s}\left(\frac{s_{1}}{\lambda_{1}} \frac{\delta \lambda_{1}}{\delta s_{1}}\right)=\frac{\sigma(s+1)}{\left[(s-1)^{2}+4 \sigma s\right]^{3 / 2}}$
$\frac{\delta}{\delta s}\left(\frac{s_{1}}{\lambda 2} \frac{\delta \lambda_{2}}{\delta s_{1}}\right)=-\frac{\sigma(s+1)}{\left[(s-1)^{2}+4 \sigma s\right]^{3 / 2}}$
Now in case of helix coil transition, in the present notation the fraction of ordered segments
$f_{h}=\frac{1}{N} \frac{\ln Z}{\ln s}=\frac{\left[1-C_{1}(1-B)\right]-\left(\alpha_{N}\right)\left[1-C_{1}(1+B)\right]}{1+B}$
where
$B=\left(\frac{X_{2}}{X_{1}}\right)^{N}\left(\frac{C_{2}}{C_{1}}\right)$
$C_{1}=\frac{X_{1-s}}{X_{1-X_{2}}} ; \quad C_{2}=\frac{X_{2-s}}{X_{2-X_{1}}}$
with $\alpha=x / y ; x=X_{1}+X_{2}$ and $y=X_{1}-X_{2}$
For $\mathrm{N} \longrightarrow \infty$ if the roots are not degenerate and $\mathrm{X}_{1}>\mathrm{X}_{2}$
$f_{h}=\left[1-C_{1}\left(\frac{1-B}{1+B}\right)\right]$
The specific heat $C_{V}$ is given as
$C_{v}=N k\left(\frac{\Delta H}{R T}\right)^{2} s \frac{\partial f_{h}}{\partial s}$
$s \frac{\partial f_{h}}{\partial s}=\frac{1}{(1+B)}\left[\left(s \frac{\delta B}{\partial s}\right)\left(C_{1}-f_{h}+C_{1} u N\right)+u C_{1}\left(C_{1}-1\right)\right.$
$\left[(1-B)-u^{(1+B h N]}-(1 / N)\left[1-(1+B) C_{1}\right]^{(1-X / 2)}\left(u^{2}-1\right)\right]$
with
$s \frac{\delta B}{\delta s}=B\left[N\left(2 C_{1}-1\right)+\alpha\right]$

## RESULTS AND DISCUSSION

All the input parameters, i.e. enthalpy change around the transition point for a given chain length, transition temperatures and nucleation parameters are given below (Table 1-3). The data in table 1 and table 2 correspond to Poly(dG-dC) ( $\mathrm{N}=2150$ ) and Poly $(\mathrm{dG}-\mathrm{dC})(\mathrm{N}=800)$ respectively ${ }_{5}$ and the data in table 3 corresponds to Poly(dG-me $\left.{ }^{5} \mathrm{dC}\right)(\mathrm{N}=500)$ : These data have been obtained from the work of Chaires and Sturtevant (1986, 1988) and Behe and Felsenfeld(1985).

## ORDER き ORDER TRANSITION

Theoretical transition curves for $\mathrm{B} \rightleftharpoons \mathrm{Z}, \mathrm{Z} \rightleftharpoons \psi(-), \mathrm{Z} \rightleftharpoons$ $B$ and $B \rightleftharpoons \psi(-)$ transitions have been obtained from equation(2). These transition curves are shown in text figures $7(\mathrm{~A}), 8(\mathrm{~A}), 9(\mathrm{~A}), 10(\mathrm{a})$ and $11(\mathrm{a})$. Apparent discontinuity at a point $D$ in figures is due to the jump in the temperature scale. Various transition parameters which provide the best fit to the experimental data are given in


Text－figure 9．Theoretical transition－curves for poly（ dG Gme 5 dC ）： （A）$B \rightleftharpoons Z$ and $Z \geqslant \Psi(-) \quad(B) \Psi(-) \geqslant$ coil
table 1,2 ，and 3 ．The end effect parameter $\times$ has been chosen to be equal to unity because end effects are neglegible for a very long chain．The conversion of Poly （dG－dC）and its methylated analogue from a right－handed to a left－handed helix has been ascribed to the salt related electrostatic interactions of polynucleotide and to the alteration of the water structure around the guanine amino group．From text－figures 7（A）and 10（a）it is clear that the sharpness of $\mathrm{B} \rightleftharpoons \mathrm{Z}$ transition increases with increase in $\mathrm{Mg}^{++}$concentration which results in the smaller value of $\sigma$ at the higher $\mathrm{Mg}^{++}$concentration．This is unlike $\psi(-)$ to coil transition where the transition is sharper at lower $\mathrm{Mg}^{++}$concentration．In general the B form is favoured at low temperatures．However，as the concentration is raised（say at 2.35 M NaCl ）the Z form is favoured．Now if the temperature is raised polynucleotide conformation shifts towards the B form（Text－fig 11 a ）． Higher salt concentration favours the Z form by masking the phosphate repulsions．From text－figure 7（A）and 9 $(\mathrm{A})$ it is obvious that $\mathrm{B} \rightleftharpoons \mathrm{Z}$ transition is sharper in the case


Text－figure 10．（a）Experimental（ $\cdot$ ）and theoretical（ $)$ transition－cur－ ves for $B \geqslant Z$ in poly（ $\mathrm{dG}-\mathrm{dC}$ ）（b）Theoretical Specific heat－curve for $B \rightleftharpoons Z$ in poly（ $d G-d C$ ）


Text－figure 11．（a）Experimental（ $\cdot$ ）and theoretical（ $(-)$ transition－cur－ ves for $\mathrm{Z} \geq \mathrm{B}$ in poly（ $\mathrm{dG}-\mathrm{dC}$ ）（b）Theoretical specific heat－curve for $Z \geqslant B$ in poly（ $d G-d C$ ）
of Poly（ $\mathrm{dG}-\mathrm{dC} \mathrm{)} \mathrm{as} \mathrm{compared} \mathrm{to} \mathrm{its} \mathrm{methylated} \mathrm{analogue}$ but in case of $Z \geqslant \psi(-)$ transition situation is otherwise． Methylation leads to the stabilization of the Z form．The B form is stabilized by the spine of water in the minor groove．The methylation of cytosine at C5 position destabilizes the B form due to unfavourable interaction between the hydrophobic methyl group and the water－ molecule．It stabilizes the Z form due to favourable inter－ actions of the methyl group with the imidazole ring of guanine and the carbon atom of sugar．In addition to the $B \geqslant Z$ transition Poly（dG－dC）also shows a right handed to left handed $(B \geqslant \psi(-))$ transition with a geometry dif－ ferent from that of $Z$（Text－fig． 13 A ）．

## HELIX COIL TRANSITION

The helix coil transition $(\psi(-)=$ coil）profiles obtained from expression（16）in case of Poly $(\mathrm{dG}-\mathrm{dC})$ and Poly $(\mathrm{dG}-$


Text－figure 12．Experimental（ $\cdot$ ）and theoretical $(-)$ specific heat－cur－ ves for poly（dGdC）：
（A）B a Z and Z きリ（－）（B）リ（ ）a coil


Text figure 13. Experimental ( $\cdot$ ) and theoretical $(-)$ specific heat-curves for poly (dG-dC):
(A) $\mathrm{B} \rightleftharpoons \Psi(-) \quad(\mathrm{B}) \boldsymbol{\Psi}(-)=$ coil
$\mathrm{me}{ }^{5} \mathrm{dC}$ ) at different $\mathrm{Mg}^{++}$concentrations are shown in text-figure $7(\mathrm{~B}) .8(\mathrm{~B})$ and $9(\mathrm{~B})$. From the text-figures, it is obvious that the transition is sharper at lower $\mathrm{Mg}^{++}$ concentration. This is also evident from the smaller value of the nucleation parameter and the larger concomitant change in enthalpy. As stated earlier, methylation leads to stabilization of the Z helix. This stabilization will also manifest itself in a smaller value of nucleation parameter while going from the left handed $Z$ to the left handed $\psi(-)$ but not necessarily in going from $\psi_{( }^{(-)}$to coil form because the degree of stabilization is different in the two forms. The experimental data also supports this. The higher value of $\sigma$ is indicative of the fact that methylation leads to a perturbed and destabilized $\mu(-)$ helix. As expected $\Delta \mathrm{H}$ is also small.

## SPECIFIC HEAT

Specific heat is an important thermodynamic quantity which characterizes the conformational and dynamical state of a biopolymeric system. Recently measurements of specific heat have been reported by Chaires and Sturtevant (1986, 1988). Specific heat is the second derivative of Gibbs free energy and theoretical details leading to the expressions (11) and (19) have been given earlier in this communication. The theoretical specific heat curves for various polynucleotides under different environmental conditions and varying chain lengths are given in text-figures $10(\mathrm{~b}), 11(\mathrm{~b}), 12(\mathrm{~A}), 13(\mathrm{~A})$ and $14(\mathrm{~A})$. Experimental results are shown by circles. These results are in the neighbourhood of the transition points for Poly (dG-dC) ( $\mathrm{N}=2150$ and $\mathrm{N}=800$ ) and Poly (dG-me ${ }^{5} \mathrm{dC}$ ) ( $\mathrm{N}=500$ ). The former undergoes $B \geqslant Z \gtrless \psi(-) \gtrless$ coil and $B \geqslant y(-) \geqslant$ coil transitions under appropriate environmental conditions whereas in case of latter the transitions observed are $B \gtrless Z \geqslant(-) \rightleftharpoons$ coil. All these specific heat curves
have a lambda point anomaly and would tend towards Delta function singularity in case of infinitely long system with large enthalpy changes. Sharpness of transition can be characterized by their half-widths $(\Delta \mathrm{T})$ given in table 1 , 2 and 3 . The closeness of the experimental and theoretical values of $\Delta T$ is a measure of the goodness of the agreement with the experimental measurements. In addition to half-width of transition the lambda curves also have a certain degree of asymmetry in the region of half-width measurement and much more so in the tail-end regions. These are most probably due to the presence of more than two phases in the system in the neighbourhood of the transition. A multiphasecomponent theory is therefore necessary.

In case of helix to coil transition, the specific heat in general has a higher magnitude as compared with order $\stackrel{\rightharpoonup}{\tau}$ order transition. This is mainly due to large enthalpy changes involved. This could also be due to large enthalpy changes involved. This could also be due to the existence of a large number of short ordered segments in the disordered state, which play an important role in detailed balance of vibrational energy.

The preponderance of right handed over all other forms of DNA can be understood from the heat of transition given in table 1 . The heat of transition from B to Z and Z to $\psi_{( }^{(-)}$is $(2.02+2.9) \mathrm{KCal} /$ mole bp whereas directly from B to $\psi(-)$ is $2.82 \mathrm{KCal} / \mathrm{mole} \mathrm{bp}$. The former takes place under $1.0 \mathrm{mM} \mathrm{Mg}^{++}$concentration and the later at $5.0 \mathrm{mM} \mathrm{M} \mathrm{M}^{++}$concentration. Since the physiological conditions are nearer the first one, the transitions from $B$ to $Z$ and $Z$ to $\psi(-)$, which are reversible. maintain the equilibrium concentration of right handed DNA whereas the other transitions from $B$ to $\psi(-)$ even though being somewhat easier (lesser heat of transition) results in the loss of right handed form. But there is also a significant probability of $\psi(-)$ reverting back to the right


Text figure 14. Experimental ( $\cdot$ ) and theoretical $(-)$ specific heat-curves for poly ( $\mathrm{dG}-\mathrm{me} 5 \mathrm{dC}$ ):
(A) $\mathrm{B} \geqslant \mathrm{Z}$ and $\mathrm{Z} \geqslant \Psi(-) \quad(\mathrm{B}) \Psi(-) \geqslant$ coil
handed DNA via left handed $Z$ form. Thus the right handed form tries to maintain higher concentration even after some loss in the left handed form. The same is also supported by the relative values of the nucleation parameter.

The more important question arises regarding the very synthesis of the left-handed form from the premodial matter. General opinion is that the preponderance of right-handed over left-handed biomolecules is related to the nonconservation of parity. The nonconservation of parity gives rise to larger emission of electrons with left handedness over those with right handedness. This in turn would imply that electromagnetic radiation related to left handed electrons will give rise to right handed polarisation and vice versa. Thus the electromagnetic radiation with right handed polarization will prevail over the left handed one. Since all chemical reactions result from an interaction of electromagnetic radiation with the reacting species, it would produce a larger number of right handed products over left handed. Further the energy difference between a right handed and left handed molecule has been worked out to be very small, i.e. roughly of the order of $10^{-7}$. This difference is extremely small in itself. But it becomes large over a period of millions of years and as such results in the dominance of right handed species over left handed. Nature is thus basically asymmetric even though it loves order. The universe heading towards the state of maximum entropy is closely related to the asymmetric nature of the universe.

## REFERENCES

Agnihotri, R.C., Mehrotra, C.. Gupta. V.D. \& Srivastava. V. 1981. Order-order transition in a macromolecular system-poly-L-proline I and II. Pramana 17 : 361-368.
Azbel. M.Y.1980a. DNA sequencing and helix-coil transition. I. Theory of DNA melting. Biopolymers $19:$ : 61-80.
Azbel. M.Y. 1980b. DNA sequencing and helix-coil transition. II. Loop entropy and DNA melting. Biopolymers $19: 81-93$.
Azbel, M.Y. 1980c. DNA sequencing and helix-coil transition. III. DNA sequencing. Biopolymers 19: 95-109.
Behe, M.J. \& Felsenfeld. G. 1985. Temperature dependent conformational transitions in poly ( $\mathrm{dG}-\mathrm{dC}$ ) and poly (dG-me ${ }^{5}$ dC).

Biopolymers 24 : 289-300.
Chaires. J.B. \& Sturtevant. J.M. 1986 Thermodynamics of B to Z transitions in poly (dG-me ${ }^{\text {r }}$ dC). Biopolymers 27: 1375-1386.
Feigon. J. Wang. A.H.J., Vander Marel, G.A., Van Boom. J.H. \& Rich. A. 1985. Z-DNA without an alternating purine-pyrimidine sequence in solution. Science 230 : 82-84.
Gulatto. M.. Goss. D.J. Diem. M. 1989. IR vibrational CD in model deoxyoligonucleotides: Observation of the $\mathrm{B}=\mathrm{Z}$ phase transition and extended coupled oscillor intensity calculations. Biopolymers 28 : 2047-2058.
Jovin, T.M.. Soumpasis. D.N. \& Mc Intash. L.P. 1987. The transition between B-DNA and Z-DNA. Ann. Rev. Phys. Chem. 38: 521560 .
Latha. P.K. \& Brahmachari, S.K. 1985. A novel structural transitions in poly (dG-me $\left.{ }^{5} \mathrm{dC}\right): \mathrm{Z}=\mathrm{B}=\mathrm{Z}$. FEBS Letters $182: 315-318$.
Latha. P.K. \& Brahmachari. S.K. 1986. B to $Z$ transitions in DNA and their biological implications. J. Sci. Ind. Res. 45: 521-533.
Latha. P.K. Majumder. K. \& Brahmachari. S.K. 1983. Temperature dependent $\mathrm{Z}=\mathrm{B}$ transitions in poly (dG-dC). Curr. Sci. 52: 907 909.

Marx. J.L. 1985. Z-DNA: Still searching for a function. Science 230 794-796.
Mc Campbell. C.R.. Wartell. R.M. \& Plaskon, R.R. 1989. Inverted repeat sequences can influence the melting transitions in linear DNAs. Biopolymers 28: 1745-1758.
Narasimhan, V. \& Bryan, A.M. 1984 Conformational flexibility of poly ( dG -me ${ }^{\text {r. }} \mathrm{dC}$ ) under very low salt concentrations. Experimentia 40 : 827-828.
Ramesh. N \& Brahmachari. S.K. 1983. Critical cation balance in $\mathrm{B}=2$ transition : Role of Li. FEBS Letters 164: 33-37.
Ramsing, N.B., Rippe, K. \& Jovin, T.M. 1989. Helix-coil transition of parallel stranded DNA. Thermodynamics of hairpin and linear duplex oligonucliotides. Biochemistry 28: 9528-9535.
Rich, A.. Nordhim, A \& Wang, A.H. 1984. The chemistry and biology of left-handed Z-DNA. Ann Rev. Biochem. 53: 791-846.
Souche. Y.S.. Latha. P.K.. Ramesh, N.. Madyan, V. \& Brahmachari. S.K. 1985. Left-handed DNA in synthetic and topologically constrained form $V$ DNA and its implications in protein recognition. J. Biosci. 8 : 5633-578.

Wang. A.H., Quigley. G.H., Kolpak. F.J., Carford, J.L., Van Boom. J.H.. Vander marel, G. \& Rich, A. 1979. Molecular structure of a left-handed double helical DNA fragment at atomic resolution. Nature 282 : 680-686.
Zacharias, W., Larson, J.E., Klysic. J., Stirdivant. S.M. \& Wells. R.D. 1982. Conditions which cause the right-handed to left-handed DNA conformational transitions. J. Biochem. 257: 2775-2782.
Zimm. B.H. \& Bragg J.K. 1959. Helix-coil transition in polypeptides. J.Chem. Phys. 31 : 526-553.

