

Neogene macro- and palynoflora from Hungary and their relationship

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The macrofloral remains recorded from the Egerian, Eggenburgian, Ottnangian, Karpatian, Badenian, Sarmatian, Pannonian and Pontian of the Hungarian Neogene have been reviewed. The macroflora has been compared with the palynoflora wherever recorded from the above stages, and their relationship has been evaluated.

Key-words—Macroflora, palynoflora, Neogene, Hungary.

INTRODUCTION

THE microfossils of plant origin in many cases could be identified only to a phylum level, and rarely up to the generic and species levels. In our studies macrofossils along with microfossils (palynofossils) were instrumental for proper interpretations, primarily when the one and the same locality was to be evaluated. It is of great importance to compare palynological data with those of the macrofloras.

In recent years attempts have been made to perform an integrated study of macrofossils and palynological data (Nagy & Pálfalvy 1960, 1961, 1963).

The major studies on the Neogene macroflora of Hungary are linked with G. Andreánszky and his co-workers. Andreánszky (1966) studied the Egerian flora, the Middle and Upper Miocene and Pliocene floras, and the Sarmatian floras (Andreánszky, 1949, 1959). Pálfalvy (1964, 1967) studied the flora of different periods, ranging from Egerian to Lower Pliocene but was mainly involved in the Middle Miocene of Hungary. Hably (1982, 1985) studied many floral assemblages of the Egerian and the Lower Miocene flora of Ipolytarnóc. A summary on Neogene gymnospermous and angiospermous fossil woods can be seen in Greguss's work (1967, 1969). From these studies attempts have been made to compare the data with that of the Neogene palynoflora

of Hungary. For this only those macrofossils have been selected which are referred along with the palynological data, without attempting to identify them up to species level. The taxa not included in both the lists have not been used.

MACRO AND PALYNOFLORAL RELATIONSHIPS

Andreánszky (1966, p.23) mentions the exposure of the Eger Brickyard as containing the richest Upper Oligocene-Lower Miocene flora, listing 123 species. Included in the palynological data, 304 taxa are from the Egerian Formation. In addition to the holostratotype, borehole sections of other Egerian formations were also dealt with. Data on macroflora represent 40.4% of the palynological data. From the macrofloral fossils of the Egerian, 49 species are turned out to be correlatable with palynological data, without attempting to identify them beyond generic level. This means that the 54 spore-pollen species representing 17.8% of the total amount of spore-pollen species were encountered in the Egerian formations. The number of comparable species of fern and gymnosperms is very small (see Egerian Table 1), whereas the angiosperms are fairly good in number. Despite this, the macrofloral data yielded more information on floral relationships than those of microflora (Table 1).

Table 1. Egerian

Macroflora	Extant taxa	Microflora
<i>Osmunda lignitum</i> Gieb. <i>O. parschlungiana</i> /Ung./Andreánszky	<i>Osmunda javanica</i> B. <i>O. regalis</i> L.	<i>Osmundacidites nanus</i> <i>O. primarius</i> <i>O. primarius crassiprimarius</i>
<i>Blechnum braunii</i> Ett.	<i>Blechnum brasiliense</i> Desv.	<i>Laevigatosporites discordatus discordatus</i> <i>L. pseudodiscordatus</i>
<i>Asplenium egedense</i> Andr.	<i>Asplenium marinum</i>	<i>L. nitidus</i> <i>Punctatisporites crassieximus</i>
<i>Pinus taedaeformis</i> /Ung./Heer	<i>Pinus taeda</i> L.	<i>P. crassimaximus</i> <i>Pinuspollenites latisaccatus</i>
<i>P. palaeostrobis</i> Ett.	<i>P. strobus</i> L.	<i>P. labdacus</i>
<i>Sequoia langsdorffi</i> /Brgt/Heer	<i>Metasequoia</i>	<i>Taxodiaceapollenites</i> sp.
<i>S. couttsiae</i> Heer	<i>Glyptostrobus</i>	<i>Taxodiaceapollenites</i> sp.
<i>Callitrites brongniarti</i> Endl.	<i>Callitris</i>	<i>Cupressacites</i> sp.
<i>Magnolia</i> cf. <i>M. grandiflora</i> L.	<i>Magnolia</i>	<i>Magnoliaepollenites simplex</i>
<i>Arbutus praenedo</i> Andr.	<i>Arbutus unedo</i> L.	<i>Ericipites callidus</i> <i>E. discretus</i> <i>E. ericius</i> <i>Proteacidites egerensis</i>
<i>Lomatites aquensis</i> Sap.	<i>Lomatia longifolia</i> A. Br./Proteaceae	<i>Alnipollenites verus</i>
<i>Alnus</i> cf. <i>A. nepalensis</i> Don	<i>Alnus nepalensis</i> Don	<i>Carpinipites carpinoides</i>
<i>Carpinus grandis</i> Ung.	<i>Carpinus japonica</i> Bl.	<i>Tricolporopollenites cingulum fusus</i>
<i>Castanopsis ? furcinervis</i> Rossm./Kr. et Wld.	<i>Castanopsis</i> sp.	<i>T. cing. pusillus</i>
<i>Lithocarpus colchica</i> Kolak.	<i>Lithocarpus/Pasania/pseudomolucca</i> Rehd.	<i>T. cing. oviformis</i>
<i>Quercus tenerrima</i> Web.	<i>Quercus chrysolepis</i> Liebm.	<i>Quercopollenites granulatus</i>
<i>Q. gigantum</i> Ett.	<i>Q. pagodaefolia</i> /Aske/Elliott	<i>Tricolporopollenites henrici</i> <i>T. microhenrici</i> <i>T. porasper</i> <i>T. villensis</i> <i>T. minimus</i> <i>Engelhardtoides microcoryphaeus</i> <i>Pterocaryapollenites stellatus</i>
<i>Q. crassipetiolata</i> Andr. et Kov.		
<i>Engelhardtia brongniarti</i> Sap.	<i>Engelhardtia</i> sp.	
<i>Pterocarya denticulata</i> /O. Web./Heer	<i>Pterocarya fraxinifolia</i> / Lam/Spach.	
<i>Juglans</i> cf. <i>J. regia</i> L.	<i>Juglans regia</i> L.	<i>Juglanspollenites verus</i>
<i>Juglans</i> cf. <i>J. cinerea</i> L.	<i>J. cinerea</i> L.	<i>J. maculosus</i>
<i>Carya felcata</i> And.	<i>Carya olivaeformis</i> Nutt.	<i>Caryapollenites simplex simplex</i>
<i>Myrica/Comptonia/acutiloba</i> Btg.	<i>Comptonia asplenifolia</i> /L./Spreng.	
<i>Myrica</i> cf. <i>M. javanica</i> Bl.	<i>Myrica javanica</i> Bl.	<i>Myricipites bituitus</i>
<i>Myrica</i> cf. <i>M. longifolia</i> Teysm. et Binn.	<i>M. longifolia</i> Teysm. et Binn.	<i>M. rurensis</i>
<i>M. lignitum</i> /Ung./Saporta	?	<i>M. myricoides</i>
<i>Salix levateri</i> Heer	<i>Salix russelliana</i> Sm.	
<i>S. arcinervia</i> O. Web.	<i>Salix</i> sp.	<i>Salixipollenites densibaculatus</i>
<i>S. varians</i> Goepp.	<i>S. fragilis</i> L.	<i>S. helveticus</i>
<i>Ulmus</i> sp. I.	<i>Ulmus americana</i> L.	<i>Ulmipollenites miocaenicus</i>
<i>Ulmus</i> sp. II.	?	<i>U. polyangulus</i>
<i>Ulmus</i> sp. III.	?	<i>U. stillatus</i>
<i>Ulmus</i> IV.	?	<i>U. undulosus</i>
<i>Ulmus</i> V.	<i>Ulmus levis</i> Pall.	
<i>Rhus succedaneoides</i> Andr.	<i>Rhus succedanea</i> Sieb. et. Zucc.	<i>Rhoipites</i> sp.
<i>Rhus.</i> cf. <i>R. glabra</i> L.	<i>R. glabra</i> L.	<i>Rhoipites pseudocingulum</i>
<i>Acer hungaricum</i> Andr.	<i>A. pennsylvanicum</i> L.	
<i>A. trilobatum</i> /Sternbg/ A. Br.	<i>A. rubrum</i> L.	<i>Aceripollenites reticulatus</i>
<i>A. agriense</i> Andr.	?	
<i>Rhamnus</i> cf. <i>R. purshiana</i> DC.	<i>R. purshiana</i> DC.	<i>Rhamnaceapollenites triquetrus</i>
<i>R. deletus</i> Heer	<i>R. confluens</i> Bolss.	
<i>Symplocos</i> cf. <i>S. phanerophlebia</i> Meer.	<i>S. phanerophlebia</i> Meer.	<i>Porocolpopollenites stereoformis</i>

<i>Symplocos</i> cf. <i>S. rubiginosa</i> Wall.	<i>S. rubiginosa</i> Wall.	<i>P. triangulus</i>
<i>Cornus büchii</i> Heer	<i>C. paniculata</i> L' Hérít.	<i>Tricolporopollenites hedwigae</i>
<i>Tuzsonia hungarica</i> Andr.	?	<i>Dicolpopollenites calamoides</i>
<i>Phoenicites leganyii</i> Andr.	?	<i>Sabalpollenites papillosus</i>
<i>Phoenicites</i> sp.	?	<i>S. retareolatus</i>
		<i>Monocolpopollenites tranquillus</i>

The macrofloral data of the Eggenburgian Stage are rather scarce. A few finds are mentioned by Jablonszky (1914), Rásky (1959) and Pálfalvy (1976) from the Ipolytarnóc Sandstone Slab with footprint records. Sandstone is less suitable for preserving pollen grains as well as macrofossils. Comparing the floral list compiled by Jablonszky and Pálfalvy with of borehole sections Püspökhatvan 4, (rich in pollen); it can be stated that all the taxa that can be botanically identified are included therein, except for species of Lauraceae and Cyperaceae families and the *Araceites hungaricus* species described by Rásky (1959).

Hably (1985) repeatedly studied the flora of the sandstone and mentions that *Ulmus pyramidalis* Göepp

occurs only in the sandstone. It is also noted that the flora cannot be separated from that of the overlying tuffs. The Eggenburgian flora of the sandstone was largely preserved by tuff beds corresponding to the Ottnangian, substage (Rásky, 1959 & Hably, 1985).

The Ottnangian flora of Ipolytarnóc was published by Hably (1985) which includes 64 species. It is recommended not to compare the macroflora on a quantitative basis, which is indicative of the lower part of the Ottnangian, with the palynological data having 209 species recorded from a number of boreholes from all over the country. However, the macrofloral list was compared with the palynological data in the following manner (Table 2).

Table 2. Ottnangian

Macroflora	Microflora
<i>Dryopteris</i> sp.	<i>Leiotriletes</i> sp.
<i>Asplenium</i> sp.	<i>Laevigatosporites</i> sp.
<i>Pinus saturni</i> Ung.	<i>Pinuspollenites labdacus</i>
<i>Pinuxylon tarnocziense</i> /Tuzson/Greguss	<i>Magnoliaepollenites simplex</i>
<i>Magnolia</i> (5 species)	<i>Tricolporopollenites henrici</i>
<i>Quercus apocynophyllum</i> Ettings.	<i>T. microhenrici</i>
" <i>Quercus</i> " <i>cruciata</i> A. Braun	<i>Engelhardtoidites microcoryphaeus</i>
<i>Engelhardtia orsbergensis</i>	<i>Caryapollenites</i> sp.
<i>Cyclocarya cyclocarpa</i> /Schlech./Knobl.	<i>Pterocaryapollenites</i> sp.
<i>Myrica hakeafolia</i> /Ung./Sap.	<i>Myricipites</i> sp.
Ebenaceae	<i>Sapotaceoidaepollenites</i> sp. Ebenales
Caesalpiniaceae : <i>Podogonium</i>	<i>Tricolporopollenites</i>
<i>oehningensis</i> /koning/kirchh.	<i>caesalpiniaceaeformis</i>
<i>Myrtophyllum</i> sp.	<i>Myrtaceidites myrtiformis</i>
<i>Acer tricuspidatum</i> Bronn	<i>Aceripollenites rotundus</i>
<i>Oreopanax protomulticaulis</i> Rásky/Hably	<i>Araliaceoipollenites edmundi</i>
<i>Schefflera gaudini</i> /Sap./Rásky	<i>A. edmundi</i> f. <i>reticulatus</i>
<i>S. protolucescens</i> Rásky	<i>A. euphorii</i>
<i>Tricalsya protojavanica</i> Rásky	Rubiaceae sp.
aff. <i>Andromeda</i> sp.	<i>Ericipites baculatus</i>
	<i>E. ericius</i>
	<i>E. discretus</i>
	<i>Sabalpollenites retareolatus</i>
<i>Sabal major</i> /Ung./Heer	

The representative of *Dryopteris* and *Asplenium* species mentioned by Hably as belonging to the Polypodiaceae are likely to be encountered in our material in great number with small *Leiotriletes* and *Laevigatosporites* species. The *Pinus saturni* Ung., *Pinus* sp., detrital fossils of conifers and the world wide-

known *Pinuxylon tarnocziense* (Tuzson) Greguss can also be correlated with *Pinuspollenites labdacus*. Hably has described 5 species of *Magnolia* species. As for *Magnolia diana* Unger, Hably reported it from the Egerian flora and also from the Ottnangian, Karpatian and Sarmatian beds. The pollen *Magnoliaepollenites*

simplex has so far been found in Egerian, Ottnagian, Middle and Upper Badenian sediments.

Two species, namely *Quercus apocynophyllum* and *Q. cruciata* assigned to the genus *Quercus* are partly of uncertain botanical relationship and can be correlated with *Tricolporollenites henrici* and *T. microhenrici*. *Engelhardtia orsbergensis* can be compared with *Engelhardtoidites microcoryphaeus*. *Cyclocarya cyclocarpa* is regarded as *Carya* or *Pterocarya* species, and thus it can represent, on palynological basis, the genera *Caryapollenites* and *Pterocaryapollenites*.

According to Hably 1985 the "*Myrica*" *hakeaefolia* is assigned to the Myricaceae but is said to be of uncertain taxonomic position. Its comparison with *Myricipites* species, which occur in significant amount throughout the Neogene and particularly in the Ottnagian, is uncertain.

Podogonium oehningense (Koenig) Kirchheimer assigned to the Caesalpiniaceae family is present in the Lower Miocene. A species described as *Tricolporopollenites caesalpiniaceaeformis* assigned to the Caesalpiniaceae has been identified from the Ottnagian in borehole Zengővarkony 45.

Leaf fossils of *Myrtophyllum* sp. assigned to Myrtaceae can be correlated with the pollen of *Myrtaceidites myrtiformis* Sim., described by Simoncsics from the Salgóterján Brown Coal Formation. This pollen has been found in the Egerian (borehole Föt 1) and Ottnagian part (911.0 m) of borehole section Nagygörbo 1.

Acer tricuspdatum can be correlated with *Aceripollenites routundus*. Of the Araliaceae family, 3 macrofossils, viz., *Oreopanax protomulticaulis* and 2 species of *Schefflera* (*S. gaudini* and *S. protolucescens*,

are included. In the Ottnagian Formation of Hungary, 3 pollen species assigned to the Araliaceae family are included, viz., *Araliaceopollenites edmundi*, *A. edmundi* cf. *raticulatus* and *A. euphorii*. However, I do not mean that they actually correspond to the aforesaid macrospecies. Hably (1982) also mentions the occurrence of the genus *Oreopanax* in the Oligocene of South France. The *Araliaceopollenites* species are of Paleogene origin.

Rásky (1955) described *Tricalysia protojavanica*, a macrofossil belonging to the Rubiaceae family, which also includes pollen from the Ottnagian of borehole section Várpálotá 133.

A fossil leaf, *Andromeda* sp. of uncertain affinity of the Ericaceae family was compared with the genus *Vaccinium* by Hably (1985). Three species of *Ericipites* are included in the Ottnagian flora, namely, *E. baculatus*, *E. ericius* and *E. discretus*. Of Palmae, *Calamus noszky* and *Sabal major* are found at Ipolytarnóc. *Calamus* is not included in the palynoflora but two other palm species, namely *Monocolpopollenites tranquillus* and *Sabalpollenites retareolatus* are included therein.

A comprehensive study of the Karpatian-Badenian flora in the Mecsek Mountains (Pálfalvy, 1964) has provided a basis for the correlation of the Karpatian-Badenian macroflora with the palynoflora. Pálfalvy does not differentiate the species belonging to the Karpatian (Helvetion) from those of the Badenian (Tortonian) beds. However, no other comprehensive study on this aspect is available. Pálfalvy's work has been used in the present study (Table 3).

Table 3

Karpatian-Badenian	Karpatian	Badenian
<i>Osmunda heeri</i> Gaud.	<i>Osmundacidites primarius crassiprimar.</i>	<i>O. primarium crassiprimarius</i> + 4 species
<i>Lygodium gaudini</i> Heer	<i>Leiotriletes</i> species	<i>Leiotriletes</i> species
<i>Gleichenites hungaricus</i> Pálf.	<i>Gleicheniidites rimosus</i>	<i>Gleicheniidites rimosus</i>
<i>Ginkgo adiantoides</i> Ung.	<i>Ginkgoretectina neogenica</i>	<i>G. neogenica</i>
<i>Pinus palaeoistobus</i> Ett.	<i>Pinuspollenites labdacus</i>	<i>P. labdacus</i>
<i>P. preasilvestris</i> Ett.	<i>P. labdacus</i>	<i>P. labdacus</i>
<i>P. teadaeformis</i> /Ung./Heer	<i>P. latisaccatus latisaccatus</i>	<i>P. latisaccatus latisaccatus</i>
<i>Keteleeria rhenana</i> Kr.	<i>Keteleeriaepollenites komloënsis</i>	<i>Keteleeriaepollenites komloënsis</i>
<i>Cedrus miocaenica</i> Lauby	<i>Cedripites hidasensis</i>	<i>Cedripites hidasensis</i>
<i>Taxodium dubium</i> /Stembg./Heer	<i>Taxodiaceapollenites</i> sp.	<i>Taxodiaceapollenites</i> sp.
<i>Sequoia langsdorfii</i> /Brgt./Heer	<i>Sequoiapollenites polyformosus</i>	<i>Sequoiapollenites polyformosus</i>
<i>Glyptostrobus europaeus</i> /Brgt./Heer	<i>Taxodiaceapollenites</i> sp.	<i>Taxodiaceapollenites</i> sp.
<i>Ephedrites sotzkianus</i> Ung.	8 <i>Ephedripites</i> species	12 <i>Ephedripites</i> species
<i>Magnolia diannae</i> Ung.		<i>Magnoliaepollenites simplex</i>
<i>Liquidambar europaea</i> A. Br.	<i>Liquidambarpollenites styracifluaeformis</i>	<i>L. styracifluaeformis</i>
<i>Acacia parschlugiana</i> Ung.	<i>Acaciapollenites varpalotaënsis</i>	
<i>Nyssa disseminata</i> /Ludw./Kirchh.	<i>Nyssapollenites contortus</i>	<i>N. contortus</i>
<i>Myrtus</i> sp.	<i>Myrtaceidites myrtiformis</i>	<i>M. mesoneus</i>

<i>Pteleacarpum bronni</i> Ung./Wld.	<i>Rutacearumpollenites komloensis</i>	<i>R. komloensis</i>
<i>Rhus heufleri</i> Heer	<i>Rhoipites pseudocingulum</i>	<i>R. pseudocingulum</i>
<i>R. pyrrhae</i> Ung.		
<i>Acer decipiens</i> A. Br.	<i>Aceripollenites reticulatus</i>	<i>A. reticulatus</i>
<i>A. inaequilobum</i> Kov.	<i>A. rotundus</i>	<i>A. rotundus</i>
<i>A. mecsekense</i> Andr.		
<i>Ilex berberidifolia</i> Heer	<i>Ilexpollenites iliacus</i>	<i>I. iliacus</i>
<i>Cyrilla hungarica</i> Pálf.	<i>Cyrillaceapollenites exactus</i>	<i>C. exactus</i>
	<i>C. megaexactus</i>	<i>C. megaexactus</i>
<i>Tetrastigmophyllum hungaricum</i> Andr.		<i>Rubiaceae</i> sp.
<i>Cornus graeffi</i> /Heer/Hantk.	<i>Tricolporopollenites edmundi</i>	<i>T. edmundi</i> f. <i>major</i>
	f. <i>major</i>	
<i>Tilia milleri</i> Ett.	6 <i>Intratrisporopollenites</i> spp.	6 <i>Intratrisporopollenites</i> species
<i>Leucothoe protogaea</i> Ung.	5 <i>Ericipites</i> species	4 <i>Ericipites</i> species
<i>Vaccinium rottense</i> Ung.		
<i>Achras pitecobroma</i> Ung.	11 <i>Sapotaceoidaepollenites</i> species	12 <i>Sapotaceoidaepollenites</i> species
<i>Mimusops hungarica</i> Andr.	3 <i>Porocolpopollenites</i> species	7 <i>Porocolpopollenites</i> species
<i>Symplocos lignitarum</i> /Quenstedt/Kirchh.	3 <i>Persicarioipollenites</i> species	2 <i>Persicarioipollenites</i> species
<i>Polygonum cardiocarpum</i> Heer		<i>Moraceae</i> sp.
<i>Ficus giebleri</i> Heer	3 <i>Ulmipollenites</i> species	4 <i>Ulmipollenites</i> species
<i>Ulmus carpinoides</i> Goepf.		
<i>U. longifolia</i> Ung.	<i>Celtipollenites komloensis</i>	<i>C. komloensis</i>
<i>Celtis hungarica</i> Pálf.	<i>Zelkovaepollenites potoniei</i>	<i>Z. potoniei</i>
<i>Zelkova ungeri</i> /Ett./Kov.	<i>Z. thiergarti</i>	<i>Z. thiergarti</i>
<i>Zelkova praelonga</i> Ung.	<i>Carpinipites carpinoides</i>	<i>C. carpinoides</i>
<i>Carpinus grandis</i> Ung.		
<i>C. kisseri</i> Berger /fruct./		<i>Triporopollenites coryloides</i>
<i>Corylus insignis</i> Heer		<i>B. betuloides</i>
<i>Betula prisca</i> Ett.	<i>Betulaepollenites betuloides</i>	
<i>B. subpubescens</i> Goepf.		
<i>Alnus feroniae</i> /Ung./Cz.	<i>Alnipollenites verus</i>	<i>A. verus</i>
<i>A. gracilis</i> Ung.		
<i>A. attenuata</i> Goepf.	5 <i>Faguspollenites</i> species	6 <i>Faguspollenites</i> species
<i>Fagus deucalionis</i> Ung.	<i>Tricolporopollenites</i>	<i>T. cingulum oviformis</i>
<i>Castanea</i> cf. <i>C. alavia</i> Ung.	<i>cingulum oviformis</i>	
	<i>T. cingulum pusillus</i>	<i>T. cingulum pusillus</i>
<i>Castanopsis decheni</i> /Web./Kr. et Wld.		
<i>Quercus böckhi</i> Staub	<i>Quercopollenites granulatus</i>	<i>Q. granulatus</i>
<i>Q. kubinyi</i> /Kov./Cz.	<i>Q. robur typus</i>	<i>Q. robur typus</i>
<i>Q. mediterranea</i> Ung.	<i>Q. petrea typus</i>	<i>Q. petrea typus</i>
<i>Q. neriifolia</i> A. Br.	<i>Juglanspollenites verus</i>	<i>J. maculosus</i>
<i>Juglans acuminata</i> A. Br.		<i>J. verus</i>
<i>J. sieboldiana</i> Max.	<i>Pterocaryapollenites stellatus</i>	<i>P. mecsekensis</i>
<i>Pterocarya denticulata</i> /Web./Heer		<i>P. rotundiformis</i>
<i>P. castaneaefolia</i> /Goepf./Schlecht	<i>Caryapollenites simplex simplex</i>	<i>C. simplex simplex</i>
<i>Carya bilinica</i> Ung.	<i>Engelhartioidites microcoryphaeus</i>	<i>E. microcoryphaeus</i>
<i>Engelhardtia marcoptera</i> /Brongt./Ett.	<i>Myricipites myricoides</i>	<i>M. myricoides</i>
<i>Myrica acuminata</i> /Ung./Sap.	<i>M. rurensis</i>	<i>M. rurensis</i>
<i>M. integerrima</i> Kr. et Weyl.		
<i>M. lignitum</i> /Ung./Sap.	<i>Salixipollenites densibaculatus</i>	<i>S. densibaculatus</i>
<i>Salix angusta</i> A. Br.	<i>S. helveticus</i>	<i>S. helveticus</i>
<i>S. media</i> A. Br.	<i>Cyperaceapollenites neogenicus</i>	<i>C. neogenicus</i>
<i>Cyperites</i> sp.	<i>Graminidites media</i>	<i>G. crassiglobosus</i> ?
<i>Phragmites oeningensis</i> A. Br.	<i>Tetradomonoporites typhoides</i>	
<i>Typha latissima</i> A. Br.	<i>Arecipites trachycarpoides</i>	<i>A. trachycarpoides</i>
<i>Trachycarpus raphifolia</i> /Stembg./Takht.	<i>Monocolpopollenites</i>	
<i>Phoenicites daemonorops</i> /Ung./Heer	<i>tranquillus</i>	
	<i>Sabalpollenites retareolatus</i>	<i>S. retareolatus</i>
<i>Sabal major</i> Ung.		

A total of 78 macroflora species can be compared with 109 taxa of the Karpatian-Badenian palynoflora. This quantitative difference itself points to the fact that the taxa have usually been correlated to the generic level.

In his study on the Sarmatian macroflora Andreánszky (1959) gives summary of comprehensive

studies and also presents a number of data from Greguss's work. Of macrofossils, those that can be correlated with the palynological data are listed in Table 4. one hundred one species of macroflora could be correlated with 85 palynological species.

Table 4. Sarmatian

Macroflora	Microflora
<i>Riccia</i> cf. <i>R. frostii</i> Aust.	<i>Ricciaesporites hungaricus</i>
<i>Osmunda parrishiana</i> /Ung./Andreánszky	<i>Osmundacidites nanus</i>
	<i>O. primarius primarius</i>
<i>Pteris paleoaurita</i> É. Kovács	<i>Polypodiaceoisporites spiniverrucatus</i> / <i>Pteris pellucida</i>
<i>Ginkgo adiantoides</i> /Ung./Heer	<i>Ginkgoretectina neogenica</i>
<i>Cedroxylon</i> sp. Andreánszky	4 <i>Cedripites</i> species
<i>Abies</i> sp.	<i>Abiespollenites absolutus</i>
<i>Picea</i> /vel <i>Tsuga</i> /	5 <i>Tsugaepollenites</i> species
<i>Picea</i> sp.	<i>Piceapollenites neogenicus</i>
<i>Pinus/Taeda/rigios</i> Ung.	<i>Pinuspollenites latisaccatus media</i>
<i>Pinus kotschyana</i> Ung.	<i>P. labdacus</i>
<i>Pinus</i> cf. <i>P. halepensis</i> Mill.	
<i>Taxodium distichum miocaenicum</i> Heer	<i>Taxodiaceapollenites</i> sp.
<i>Taxodioxyton taxodii</i> Gothan	<i>Taxodiaceapollenites</i> sp.
<i>Sequoia langsdorfii</i> /Brgt./Heer	<i>Taxodiaceapollenites</i> sp.
<i>Taxodioxyton sequiadendri</i> Andreánszky	<i>Sequiapollenites polyformosus</i>
<i>Tetracentron hungaricum</i> Andreánszky	
<i>Tetracentronites hungaricus</i> Greguss	<i>Tetracentracearumpollenites minimus</i>
<i>Liquidambar europaea</i> A. Br.	<i>Liquidambarpollenites styracifluaeformis</i>
<i>Nyssa hungarica</i> Andr.	<i>Nyssapollenites contortus</i>
	<i>N. pseudocruciatus</i>
	<i>Slovakipollis elaeagnoides</i>
	<i>Rhoipites pseudocingulum</i>
	<i>Acerpollenites rotundus</i>
	<i>A. reticulatus</i>
	<i>Ilexpollenites iliacus</i>
	<i>I. margaritatus</i>
	<i>I. propinquus</i>
	<i>Vitipites sarmaticus</i>
	<i>Caprifoliipites andreanszkyi</i>
	<i>Viburnum rhytidophyllum</i> Hemsl.
	<i>Caprifoliipites gracilis</i>
	<i>Tricolporopollenites edmundi major</i>
	<i>T. hedvigae</i>
	<i>Loniceraipollis gallwitzii</i>
	5 <i>Intratrisporopollenites</i> species
	<i>Reevesiapollis triangulus</i>
	<i>Oleoidearumpollenites chinense</i>
	5 <i>Ericipites</i> species
	<i>S. rotundus</i>
	<i>S. sapotoides</i>
	<i>Ulmipollenites stillatus</i>
	<i>U. undulosus</i>
	<i>Zelkovaepollenites potonieii</i>
<i>Elaeagnus acuminata</i> Web.	
<i>Rhus</i> cf. <i>R. glabra</i> L.	
<i>Acer</i> 87 species	
<i>Ilex parrishiana</i> Ung.	
<i>Ilex oreadum</i> Ett.	
<i>I.</i> cf. <i>I. mexopaca</i> Ait.	
<i>Vitis</i> cf. <i>V. aestivalis</i> Mchx.	
<i>V. tokayensis</i> Stur	
<i>V. teutonica</i> A. Br.	
<i>Viburnum hungaricum</i> Andr.	
<i>Viburnum</i> cf. <i>V. dentatum</i> L.	
<i>Viburnum</i> cf. <i>V. tinus</i> L.	
<i>Cornus</i> cf. <i>C. sanguinea</i> L.	
<i>Cornus</i> cf. <i>C. ralba</i>	
<i>C. praeamomum</i> É. Kovács	
<i>Lonicera</i> cf. <i>L. chrysantha</i> Turcz.	
<i>L. liphayana</i> Andr.	
<i>Tilia vindobonensis</i> Stur	
<i>T. sarmatica</i> Andreánszky	
<i>Sterculiaceae</i> 2 species	
<i>Ligustrum</i> sp.	
<i>Ericaceae</i> species 3	
<i>Sapotacites minor</i> Ett.	
<i>Ulmus</i> 12 species	
<i>Zelkova</i> 6 species	

<i>Celtis occidentaloïdes</i> É. Kov.	<i>Celtipollenites komloënsis</i>
<i>Carpinus</i> 7 species	<i>Carpinipites carpinoides</i>
<i>Carpinoxylon hungaricum</i> Greguss	
<i>Ostrya</i> 4 species	<i>Ostryapollenites rhenanus</i>
<i>Corylus</i> 4 species	<i>Triporopollenites coryloides</i>
<i>Betula</i> 14 species	<i>Betulaepollenites betuloides</i>
<i>Alnus</i> 23 species	<i>Alnipollenites verus</i>
<i>Fagus</i> cf. <i>F. orientalis</i> Lipsky	<i>Faguspollenites gemmatus</i>
<i>Fagus</i> cf. <i>F. grandifolia</i> Ehrh.	<i>F. minor</i>
<i>Fagus</i> 6 species	<i>Faguspollenites</i> 4 species
<i>Castanea</i> 5 species	<i>Tricolporopollenites cingulum oviformis</i>
<i>Quercus</i> 41 species	<i>Quercopollenites</i> 3 species
	<i>Tricolporopollenites microhenrici</i>
	<i>Juglanspollenites maculosus</i>
<i>Juglans</i> 5 species	<i>J. verus</i>
	<i>Caryapollenites simplex</i>
<i>Carya</i> 5 species	<i>Pterocaryapollenites mecsekensis</i>
<i>Pterocarya denticulata</i> /O. Web./Heer	<i>P. rotundiformis</i>
<i>P. castaneifolia</i> /Gepp./Schlecht.	<i>P. stellatus</i>
<i>Pterocaryoxylon</i> sp.	<i>Engelhardioidites microcoryhaeus</i>
<i>Engelhartia brogniarti</i> Sap.	
<i>Myrica longifolia</i> Ung.	<i>Myricipites myricoides</i>
<i>Myrica</i> cf. <i>M. microcarpa</i> Benth.	<i>M. rurensis</i>
<i>M. deperdita</i> Ung.	
<i>M. integrifolia</i> Ung.	
<i>Salix angusta</i> A. Br.	<i>Salixipollensites densibaculatus</i>
<i>S.</i> cf. <i>S. fragilis</i> L.	<i>S. helveticus</i>
<i>S. arcinervia</i> O. Web.	
<i>S. pentandra miocaenica</i> Kubát	<i>Arceipites chamaedoriformis</i>
<i>Phoenicites</i> sp. I.	
<i>Phoenicites</i> sp. II.	
<i>Eucaryoxylon crystallophorum</i> Müller-Stoll et Madel.	

The Pannonian macrofloras are poorly studied. The macroflora of Rudabanya localities assigned to the Pannonian from the Vilmos opencast by Nagy and Pálfalvy (1961) is the older one, whereas the opencast mine Andrásy III is the younger probably (*pers. comm.* L. Kardos). The beds of the Vilmos opencast mine have a

poor macro- and microfloral assemblages. Nevertheless, this is the only one to be reckoned since the other one is uncertain. Fossilized tree-trunk described by Horváth (1954) from Megyaszó contributes to the knowledge of the macroflora (Table 5).

Table 5. Pannonian

Macroflora	Microflora
<i>Osmunda porschlugiana</i> /Ung./And.	<i>Osmundacidites primarius primarius</i>
<i>Ginkgo adiantoides</i> /Ung./Heer	<i>Ginkgoretectina neogenica</i>
<i>Pinus</i> sp.	<i>Pinuspollenites labdacus</i>
<i>Taxodium dubium</i> /Stembg./Heer	
<i>Glyptostrobus europaeus</i> /Brgt./Heer	<i>Taxodiaceapollenites</i> sp.
<i>G. tenerum</i> /Kraus./Cosw.	
<i>Sequoioxylon gypsaceum</i> /Goeppl./Greg.	
<i>Liquidambar europae</i> A Br.	<i>Liquidambarpollenites styracifluaeformis</i>
<i>Liquidambaroxylon speciosum</i> Felix	
<i>Nyssa disseminata</i> /Ludw./Kirchh.	<i>Nyssapollenites contortus</i>
	<i>N. pseudocruciatus</i>
<i>Myriophyllum</i> sp.	<i>Myriophyllumpollenites balatonensis</i>
	<i>M. minimus</i>
	<i>M. quadratus</i>
<i>Byttneriophyllum tiliaefolium</i> /A.Br./Knobl. et Kvac.	<i>Reevesiapollis triangulus</i> /Sterculiaceae
<i>Celtis</i> sp.	<i>Celtipollenites komloënsis</i>

<i>Celtixylon</i> cf. <i>Celtis occidentalis</i> L.	<i>Zelkovaepollenites potoniei</i>
<i>Zelkova zelkovaefolia</i> /Ung./Buz. et Kotl.	
<i>Zelkovoxyton</i> cf. <i>Zelkova serrata</i> L.	
<i>Ulmus pyramidalis</i> Goebb.	<i>Ulmipollenites polyangulus</i>
<i>Ulmoxylon</i> cf. <i>Ulmus americana</i> L.	<i>U. stillatus</i>
	<i>U. undulosus</i>
	<i>Carpinipites carpinoides</i>
<i>Carpinus grandis</i> Ung.	
<i>Betula alba</i> L./ <i>pubescens</i> /	
<i>Betula macrophylla</i> /Goebb./Heer	<i>Betulaepollenites betuloides</i>
<i>Betula</i> cf. <i>B. lanta</i> L.	
<i>Betuloxyton priscum</i> Felix	
<i>Alnus crebrinervis</i> É. Kov.	<i>Alnipollenites verus</i>
<i>Alnus</i> cf. <i>A. incana</i> /L./Inch.	
<i>Alnus</i> cf. <i>A. japonica</i> /Thunbg./Steud.	
<i>Fagus</i> sp./cf. <i>F. orientalis</i> Lipsky/	<i>Faguspollenites gemmatus</i>
<i>F. haidingeri</i> Kov. sensu Knobl.	5 <i>Faguspollenites</i> species
<i>Quercus</i> sp.	3 <i>Quercopollenites</i> species
<i>Quercinium böckhianum</i> Felix	
<i>Juglans acuminata</i> A. Br.	<i>Juglanspollenites verus</i>
<i>Pterocarya castanaefolia</i> /Goebb./Schl.	<i>Pterocaryapollenites mecsekensis</i>
	<i>P. rotundus</i>
	<i>P. stellatus</i>
<i>Carya denticulata</i> /Web./Schim p.	<i>Caryapollenites simplex simplex</i>
<i>Engelhardtia macroptera</i> /Brgt./Ett	<i>Engelhardtoidites microcoryphaeus</i>
<i>Myrica</i> sp.	<i>Myricipites myricoides</i>
	<i>M. rurensis</i>
<i>Salix varians</i> Goebb.	<i>Salixipollenites densibaculatus</i>
<i>S. palaeo-purpurea</i> Fr. Mey.	
<i>Phragmites oeningensis</i> A. Br.	<i>Graminidites</i> sp.
<i>Palmoxylon</i> sp.	<i>Sabalpollenites retareolatus</i>
<i>Typha latissima</i> A. Br.	<i>Tetradomonoporites typhoides</i>

A total of 41 macrofossil species were compared with 41 spore-pollen species that might correspond to other genera (see Table 5). This comparison can hardly be used as a tool for dating since all the concerned species can

also be found in Pontian formations. Perhaps, this fact allows to make it perceptible that the number of finds is much smaller here than in the Pontian (Table 6).

Table 6. Pontian

Macroflora	Microflora
<i>Ginkgo biloba</i> /lignite/	<i>Ginkgoretectina neogenica</i>
<i>Sequoia</i> sp.	<i>Sequoiapollenites polyformosus</i>
<i>Taxodioxyton sequoianum</i> /Mercklin/Gothan	
<i>Taxodium distichum miocaenicus</i> Heer	<i>Toxodiaceapollenites</i> sp.
<i>Taxodioxyton taxodii</i> Gothan	<i>Toxodiaceapollenites</i> sp.
<i>Glyptostrobus europaeus</i> /Brgt./Heer	
<i>Pinus</i> sp.	<i>Pinuspollenites labdacus</i>
<i>Pinus</i> cf. <i>P. cembra</i> L.	
<i>Trapa</i> cf. <i>T. natans</i> L.	<i>Sporotrapoidites hungaricus</i>
<i>Acer campestre</i> L. foss.	<i>Aceripollenites reticulatus</i>
<i>A. monspessulanum</i> L. foss. <i>plioaenicus</i> Mädlar	<i>A. rotundus</i>
<i>A. opulifolium plioaenicum</i> Sap.	
<i>A. polymorphum plioaenicum</i> Sap.	
<i>Ulmus</i> sp. /fol. et carp./	<i>Ulmipollenites maculosus</i>
	<i>U. polyangulus</i>
	<i>U. stillatus</i>
	<i>U. undulosus</i>
<i>Zelkova ungeri</i> Kov.	<i>Zelkovaepollenites potoniei</i>
<i>Carpinus betulus</i> L. foss.	<i>Carpinipites carpinoides</i>

Alnus cf. *A. incana* Mnch.
Fagus orientalis Lipsky
Quercus drimeia Ung.
Quercus cf. *sessiliflora* Salisb.
Quercus sp.
Engelhardtia brogniarti Sap.
Salix sp.
Typha sp.
T. latifolia L.
T. angustifolia
 Leguminosae/foliola/

Alnipollenites verus
Faguspollenites gemmatus
Quercopollenites granulatus
Q. petrea typus
Q. robur typus
Engelhardtites microcoryphaeus
Salixipollenites densibaculatus

Tetradomonoporites typhoides

 Leguminosae sp.

A brief summary of the macroflora of Pontian Formation, in relation to the flora of Rózsaszentmárton was made partly by Pálfalvy (1952) and partly by Vörös (1955). Some 25 macroflora species were within reach for correlating them with 23 pollen species.

Through this contribution I wish to express my respects to Professor Birbal Sahni on the occasion of 14th November 1991, his one hundredth birth anniversary.

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