POLLEN MORPHOLOGICAL STUDIES IN *CAPSICUM*-II. INTERSPECIFIC HYBRIDS, INDUCED TETRAPLOIDS AND DESYNAPTIC MUTANTS

O. ANIEL KUMAR, RAMESH C. PANDA, K. G. RAJA RAO & C. G. K. RAMANUJAM*

Department of Botany, Andhra University, Waltair, India. *Department of Botany, Post Graduate College of Science, Osmania University, Saifabad, Hyderabad, India.

Abstract

Detailed pollen morphology has been studied in eight F_1 interspecific hybrids, two induced autotetraploids and one desynaptic mutant. The pollen grains are prolate-spheroidal to subprolate, tricolporate to tetracolporate and psilate. The size of the grains showed variation at different ploidy levels. The variation in size and shape, apocolpial diameter, and mesocolpial diameter is higher in induced autotetraploids and desynaptic mutant, but intermediate in F_1 hybrids. Finally, the significance of pollen morphology in unravelling the evolutionary status of the above taxa vis-a-vis their parents has also been discussed.

Introduction

The importance of pollen morphology as an expression of the geographical distribution, genomal constitution, taxonomy and evolution of plants has been demonstrated in recent years. Palynology of species and varieties of the genus Capsicum has been studied and the interrelationships among them were deduced in part I. Information on the pollen morphology of species hybrids, induced polyploids and desynaptic mutants in angiospermic taxa is rather meagre. There are, however, a few reports on the pollen morphology of interspecific hybrids, viz., Sonneratia (Muller, 1969), Hibiscus (Nair, 1961), Amaranthus (see Nair, 1970; Pal & Khoshoo, 1966) and Physalis (Lydia Prasad et al., 1984). Similarly a few reports on pollen morphology of polyploids, viz., Rice (Sampath & Ramanathan 1951; Maurizio, 1956; Rangaswamy & Raman, 1973) and Physalis (Lydia Prasad et al., 1984) have been made. Also the pollen morphology in induced mutants has been investigated in Pimpinella (Joshi & Raghuvanshi, 1967) and Capsicum (Raghuvanshi, 1976). Still there is no information available on the palynology of the interspecific hybrids, induced polyploids and desynaptic mutants in the genus Capsicum. The present study documents for the first time the pollen morphology of species hybrids, induced polyploids and mutants of Capsicum and its significance in our understanding of the interrelationships of these taxa.

Materials and Methods

The eight interspecific F_1 hybrids, viz., C. annuum var. $G3 \times C$. frutescens, C. annuum var. cerasiformis x C. chinense var. mishme, C. annuum var. cerasiformis x C. pendulum, C. frutescens var. tabasco x C. annuum var. cerasiformis, C. chinense x C. frutescens, C. chacoense x C. annuum var. BOB, C. chacoense \times C. chinense var. mishme were obtained through artificial hybridization. Autotetraploids were obtained in two cultivars of C. annuum through colchicine administered to the seedlings by the following methods: (i) 10 day old seedlings of C.

Geophytology, 17(1): 1-11, 1987.

annuum var. cerasiformis were treated with aqueous solution of 0.3% colchicine by applying the chemical to the apical growing buls using a small cotton plug to keep the bud continuously wet for two days, (ii) the growing tips of three-week old seedlings of *C. annuum* var. *TNK* in the field at eight leaves stage were cut and 0.3% aqueous solution of colchicine in cotton swabs was applied to the cut end continuously for six hours. A induced desynaptic mutant was obtained through the seed treatment of 0.2% aqueous colchicine solution for 48 hours.

The fresh polliniferous material of interspecific hybrids, tetraploids and the desynaptic matants were collected from the adult plants. Pollen preparations were made by revised acetolysis method of Erdtman (1969). Observations have been recorded with the help of light microscopy (Carl Zeiss binocular). The measurements are based on an average of fifty non-chlorinated grains randomly selected but excluding the deviating or abortive pollen. The terminology used is that of Faegri and Iverson (1964) and Murry and Eshbaugh (1971) and the descriptions of pollen are in accordance with Erdtman (1952, 1969).

Description of Pollen Grains

I. Interspecific hybrids:

1. C. annuum var. $G3 \times C$. frutescens var. tabasco: Shape: prolate-spheroidal to subprolate, semi-angular in polar view, Size: $23.73 \pm 1.46 \ \mu$ (P) $\times 20.44 \pm 0.11 \ \mu$ (E) PEI = $109.25 \pm 0.53 \ \mu$ and PAI= $15.33 \pm 0.43 \ \mu$ size varied from very small to larger pollen grains. Structure: Exine tectate $1.24 \pm 0.01 \ \mu$ thick, ektexine thicker than endexine. A general thickening noticeable near the apertures. In polar view equatorial diameter $20.57 \pm 0.9 \ \mu$, mesocolpial diameter $12.56 \pm 0.14 \ \mu$ and apocolpial diameter $3.13 \pm 0.09 \ \mu$ in equatorial view. Apertures: Tricolporate, angulaperturate, inaperturate grains also observed rarely. Margos distinct, os lalongate, constricted, extends half the width of the grain. Sometimes the apertures irregularly or obliquely placed, surface psilate (Pl. 1, Figs. 1-3, Table 1).

2. C. annuum var. cerasiformis \pm C. chinense var. mishme: Shape: prolate-spheroidal, subtriangular in polar view, Size: $24.87 \pm 0.29 \mu$ (P) $\pm 22.48 \pm 0.29 \mu$ (E), PEI = 110.96 $\pm 1.27 \mu$ and PAI = $17.09 \pm 0.74 \mu$. Size also varies from very small to large, aborted grains very irregular. Structure: Exine tectate $1.16 \pm 0.01 \mu$ thick, ektexine and endexine neariy of same thickness. In polar view equatorial diameter $20.57 \pm 0.33 \mu$, mesocolpial diameter $12.44 \pm 0.28 \mu$ and apocolpial diameter $3.83 \pm 0.17 \mu$ in equatorial view. Apertures: mostly tricolporate, either irregular or obliquely placed, angulaperturate, os lalongate, constricted os extends half the width of the grain, outer ends tapering to slightly rounded. Surface psilate (Pl. 1, Figs. 4-6; Table 1).

3. C. annuum var. cerasiformis $\times C$. pendulum: Shape: prolate-spheroidal, subtriangular in polar view. Size: $25.53 \pm 0.25 \mu$ (P) x $22.68 \pm 0.18 \mu$ (E), PEI = $112.64 \pm 0.98 \mu$ and PAI = $13.38 \pm 0.67 \mu$, aborted grains irregular in sizes. Structure: Exine tectate $1.20 \pm 0.02 \mu$, thick, ektexine and endexine nearly equal in thickness. In polar view the equatorial diameter $22.36 \pm 0.21 \mu$, mesocolpial diameter $13.36 \pm 0.24 \mu$ and apocolpial diameter $4.12 \pm 0.14 \mu$ in equatorial view. Apertures: mostly tricolporate rarely tetracolporate, angulaperturate, in equatorial view furrows bordered by distinct margos, os lalongate, unconstricted, extends half the width of the grain, outer ends tapering. Surface psilate (Pl. 1, Figs. 7-9; Table 1).

				some number				
Taxa	2n	Polar diameter (P)	Equatorial diameter (E) (equatorial view)	Polar equatorial index P/E X 100	Equatorial diameter (polar view)	Apocolpial diameter	Mesocolpial diameter	Polar area index A/E X 100
C. annuum var. G3 X C. frutescens	24	$\begin{array}{c} 23.73 \pm 1.46 \\ (20.01 - 26.72) \end{array}$	$\begin{array}{c} 20.44\pm \ 0.11 \\ (18.32-24.34) \end{array}$	$\begin{array}{cccc} 109.25 \pm & 0.53 \\ (91.20 \pm 123.63) \end{array}$	$\begin{array}{c} 20.57\pm \ 0.09\\ (17.79-26.02) \end{array}$	3.13 ± 0.09 (2.83.6.90)	$\begin{array}{c} 12.56\pm \ 0.14 \\ (10.62-2.44) \end{array}$	$\begin{array}{c} 15.33 \pm 0.43 \\ (11.72 - 29.06) \end{array}$
C. annum var. cerasi- formis × C. chinense var. mishme	24	$\begin{array}{c} 24.87 \pm \ 0.29 \\ (21.06 - 28.47) \end{array}$	22.48 ± 0.29 (17.35-26.46)	110.96 ± 1.27 (94.44—137.12)	$\begin{array}{c} 20.57 \pm \ 0.33 \\ (16.55 - 24.51) \end{array}$	3.83 ± 0.17 (0.00-5.34)	$\begin{array}{c} 12.44 \pm \ 0.28 \\ (9.03 - 15.84) \end{array}$	$\begin{array}{c} 17.09 \pm \ 0.74 \\ (11.95 - 24.60) \end{array}$
C. annum var. cerasiformis $ imes$ C. pendulum	24	$\begin{array}{c} 25.53 \pm 0.25 \\ (20.44 - 28.76) \end{array}$	$\begin{array}{c} 22.68 \pm \ 0.18 \\ (19.56 - 26.55) \end{array}$	$113.64\pm 0.98 (94.67-130.67) ($	$\begin{array}{c} 22.36\pm\ 0.21\\ 18.58-25.75) \end{array}$	$\begin{array}{c} 4.12 \pm 0.14 \\ (2.57 - 6.90) \end{array}$	$13.36 \pm 0.24 \\ (9.73 - 16.72)$	18.38 ± 0.67 (10.34-29.77)
C.frutescens var. tabasco × C. annuum var. cerasiformis	24	$\begin{array}{c} 23.50 \pm \hspace{0.1cm} 0.30 \\ (19.20 - 26.64) \end{array}$	$\begin{array}{c} 21.85\pm \ 0.32 \\ (17.70-26.81) \end{array}$	$\begin{array}{c} 108.80 \pm & 1.55 \\ (88.34 - 124.01) \end{array}$	$\begin{array}{c} 20.55\pm \ 0.33\\ (15.13-23.45)\end{array}$	3.85 ± 0.14 (2.48—5.49)	$\begin{array}{c} 12.39 \pm 0.27 \\ (8.58 - 18.58) \end{array}$	$\begin{array}{c} 18.36 \pm 0.66 \\ (12.03 - 38.41) \end{array}$
C. chinense var. mishme × C. frutescens var. tabasco	24	$\begin{array}{c} 22.73 \pm \ 0.20 \\ (20.88 - 28.41) \end{array}$	$\begin{array}{c} 20.96\pm \ 0.15 \\ (18.85-24.16) \end{array}$	108.62 ± 0.81 (96.33 -124.78) ($\begin{array}{c} 20.58\pm \ 0.25\\ 13.72-27.88) \end{array}$	3.09 ± 0.11 (1.77-5.04)	$13.43 \pm 0.23 \\ (8.78 - 16.46)$	14.76 ± 0.51 (8.77-23.73)
C. chacoense × C. annuum var. BOB	24	$\begin{array}{c} 2.24 \pm \ 0.26 \\ (17.79 - 25.66) \end{array}$	$\begin{array}{c} 7.18 \pm \hspace{0.1cm} 0.27 \\ (15.66 - 24.07) \end{array}$	105.44 ± 1.23 (91.53—120.72) ($\begin{array}{c} 18.67 \pm \ 0.34 \\ 10.53 - 24.78 \end{array}$	$\begin{array}{c} 2.89 \pm 0.09 \\ (1.06 - 4.51) \end{array}$	$\frac{11.55 \pm 0.30}{(8.32 - 19.38)}$	$\begin{array}{c} 13.81 \pm \ 0.51 \\ (8.47 - 23.38) \end{array}$
C. chacoense × C. frutescens var. labasco	24	27.93 ± 0.19 (24.69 - 29.91)	$\begin{array}{c} 25.35 \pm \ 0.20 \\ (22.12 - 27.43) \end{array}$	$110.34 \pm 0.86 \\ (98.72 - 125.62) $	26.09 ± 0.16 24.34 - 29.20	$\begin{array}{c} 4.58\pm0.11\\ (3.09-6.28) \end{array}$	$\begin{array}{c} 19.34 \pm \ 0.45 \\ (13.09 - 23.98) \end{array}$	$\begin{array}{c} 18.03 \pm \ 0.46 \\ (11.72 - 24.21) \end{array}$
C. chcoense × C. chinense var. mishme	24	$\begin{array}{c} 22.37 \pm \ 0.16 \\ (19.38 - 23.98) \end{array}$	19.5 ± 0.12 (17.70-12.33) ($\begin{array}{c} 114.27\pm \ 0.71\\ (104.57-125.99) \end{array}$	$\begin{array}{c} 19.55\pm \ 0.17\\ (15.84-21.33)\end{array}$	2.97 ± 0.07 (1.86 -3.81)	$\begin{array}{c} 12.70 \pm 0.15 \\ (10.62 - 15.33) \end{array}$	15.26 ± 0.40 (10.01-22.99)
C. annuum var. cerasiformis	48	36.91 ± 0.21 (34.07 - 39.39)	31.89 ± 0.26 (26.28-35.40)	$\begin{array}{c} 116.03 \pm 0.99 \\ (98.41 - 134.02) \end{array}$	33.02 ± 0.16 (30.80-35.13)	5.22 ± 0.14 (2.74—6.99)	23.65 ± 0.20 (18.58-25.75)	16.43 ± 0.44 (8.10-25.18)
C. annuum var. TNK	48	$\begin{array}{c} 38.19 \pm \ 0.31 \\ (31.59 - 42.57) \end{array}$	$\begin{array}{c} 31.67\pm \ 0.21 \\ (28.14-33.72) \end{array}$	$\begin{array}{c} 121.35\pm 1.15\\ (94.68-134.89)\end{array}$	32.94 ± 0.32 (26.99 -39.82)	5.60 ± 0.11 (4.78-7.61)	22.46 ± 0.24 (18.58–25.75)	$\begin{array}{c} 19.01 \pm \hspace{0.1cm} 0.34 \\ (14.92 - 25.13) \end{array}$
C. annuum var cerasiformis desynaptic mutant	24	$\begin{array}{c} 23.42 \pm \ 0.22 \\ (18.14 - 26.37) \end{array}$	$\begin{array}{c} 20.00 \pm 0.21 \\ (15.75 - 22.30) \end{array}$	117.57 ± 1.34 (103.91-136.07)	20.08 ± 0.24 (16.72-23.89)	2.17 ± 0.19 (0.00-4.34)	13.61 ± 0.14 (11.41—15.13)	10.83 ± 0.97 (0.00-17.34)

Table 1—Mean frequency of pollen parameters of the hybrids, induced autotetraploids and desynaptic mutant of Capaian and their chromo-

4. C. frutescens var. tabasco \times C. annuum var. cerasiformis: Shape: Prolate-spheroidal, subtriangular in polar view. Size: $23.50 \pm 0.30 \mu$ (P) $\times 21.85 \pm 0.32 \mu$, PEI=108.80 \pm 1.55μ and PAI=18.36 $\pm 0.66 \mu$. Structure: Exine tectate $1.13 \pm 0.03 \mu$ thick, ektexine thicker than endexine. A general thickening present near the apertures. In polar view the equatorial diameter $20.55 \pm 0.33 \mu$, mesocolpial diameter $12.39 \pm 0.27 \mu$ and apocolpial diameter $3.85 \pm 0.14 \mu$ in equatorial view. Apertures: Tricolporate, angulaperturate, syncolporate, colpae bordered by distinct margos, os lalongate, deeply constricted extends more than half the width of the grain, outer ends slightly rounded. Surface psilate (Pl. 1, Figs. 10-12; Table 1).

5. C. chinense var. mishme \times G. frutescens var. tabasco: Shape: Prolate-spheroidal to subprolate, semiangular in polar view. Size: $22.73 \pm 0.20 \mu$ (P) $\times 20.96 \pm 0.15 \mu$ (E) PEI=108.62±0.81 μ and PAI=14.76±0.51 μ . Structure: Exine tectate $1.24\pm0.01 \mu$ thick, ektexine thicker than endexine. A general thickening present near the apertures. In polar view the equatorial diameter $20.58\pm0.25 \mu$, mesocolpial diameter $13.43\pm0.23 \mu$ and apocolpial diameter $3.09\pm0.11 \mu$ in equatorial view. Apertures: Mostly tricolporate occasionally tetracolporate, angulaperturate, inaperturate grains also observed. Margos distinct, os lalongate, constricted, extends half the width of the grain, outer ends slightly tapering to rounded. Rarely syncolporate condition was also observed. Surface psilate (Pl. 2, Figs. 13-15; Table 1).

6. C. chacoense \times C. annuum var. BOB: Shape: Prolate spheroidal to subprolate, semiangular in polar view. Size: 22.24±0.26 μ (P) \times 21.18±0.27 μ (E), PEI=105.44± 1.23 μ and PAI=13.81±0.51 μ . Structure: Exine tectate 1.26±0.03 μ thick, ektexine and endexine nearly equal in thickness. A general thickening noticeable near the apertures. In polar view the equatorial diameter 18.67±0.34 μ , mesocolpial diameter 11.55±0.30 μ , and apocolpial diameter 2.89±0.09 μ in equatorial view. Apertures: Tricolporate, occasionally tetracolporate, angulaperturate, rarely fossaperturate, occasionally inaperturate grains also observed. Os lalongate, deeply constricted, extends half the width of the grain, outer ends pointed to slightly rounded. Surface psilate (Pl. 2, Figs. 16-18; Table 1).

7. C. chacoense \times C. frutescens var. tabasco: Shape: Prolate-spheroidal to subprolate, semiangular in polar view. Size: $27.93 \pm 0.19 \mu$ (P) $\times 25.35 \pm 0.20 \mu$ (E) PEI=110.34 \pm 0.86 μ and PAI=18.03 $\pm 0.46 \mu$. Structure: Exine tectate $1.43 \pm 0.09 \mu$ thick, ektexine thicker than endexine. A general thickening noticeable near the apertures. In polar view the equatorial diameter 26.09 ± 0.16 , mesocolpial diameter $19.34 \pm 0.45 \mu$ and apocolpial diameter $4.58 \pm 0.11 \mu$ in equatorial view. Apertures: Mostly tricolporate, occasionally tetracolporate, angulaperturate or fossaperturate, rarely syncolporate, margos indistinct, os lalongate, deeply constricted to non-constricted, extends three fourth width of the grain, outer ends pointed to slightly rounded. Surface psilate (Pl.2, Figs. 19-21; Table 1).

8. C. chacoense \times C. chinense var. mishme: Shape: Prolate-spheroidal to subprolate, angular to semiangular in polar view. Size: $22.37 \pm 0.16 \ \mu$ (P) $\times 19.55 \pm 0.12 \ \mu$ (E), PEI=114.27±0.71 μ and PAI=15.26±0.40 μ . Structure: Exine tectate $1.08\pm0.03 \ \mu$ thick, ektexine and endexine nearly equal in thickness. A general thickening present near the apertures. In polar view, equatorial diameter $19.55\pm0.17 \ \mu$ m, mesocolpial diameter $12.70\pm0.15 \ \mu$, and apocolpial diameter $2.97\pm0.07 \ \mu$ in equatorial view. Apertures: Mostly tricolporate rarely tetracolporate, angulaperturate or fossaperturate, less frequently inaperurate grains also noticed. Margos distinct to indistinct, os lalongate, deeply constricted, extends half the width of the grain, outer ends pointed to slightly rounded. Surface psilate (Pl. 2, Figs. 22-24; Table 1).

II. Colchicine induced variants of C. annuum:

(a) Autotetraploids:

1. var. TNK: Shape: Subprolate, tri or quadrilobed, subtriangular in polar view. Size: $38.19 \pm 0.31 \ \mu$ (P) $\times 31.67 \pm 0.21 \ \mu$ (E), PEI= $121.35 \pm 1.15 \ \mu$ and PAI= $19.01 \pm 0.34 \ \mu$. Structure: Exine tectate $1.140 \pm 0.02 \ \mu$ thick, ektexine and endexine equal in thickness. In polar view equatorial diameter $32.94 \pm 0.16 \ \mu$, mesocolpial diameter $22.46 \pm 0.24 \ \mu$ and apocolpial diameter $5.60 \pm 0.11 \ \mu$ in equatorial view. Apertures: Tricolporate to tetracolporate, angulaperturate, colpae bordered by distinct margos, cs lalongate constricted to non-constricted, extends half the width of the grain, outer ends slightly rounded. Surface psilate (PI. 3, Figs. 25-30; Table 1).

2. var. cerasiformis: Shape: Subprolate, tri or quadrilobed or hexalobed, semiangular in polar view. Size: $36.91 \pm 0.21 \ \mu$ (P) × $31.89 \pm 0.26 \ \mu$ (E) PEI=116.03±0.99 μ and PAI=16.43±0.44 μ . Structure: Exine tectate $1.32\pm0.02 \ \mu$ thick, ektexine thicker than endexine. A general thickening noticeable near the opertures. In polar view the equatorial diameter $33.02\pm0.16 \ \mu$, mesocolpial diameter $23.65\pm0.20 \ \mu$ and apocolpial diameter $5.22\pm0.14 \ \mu$ in equatorial view. Apertures: Tricolporate or tetracolporate, angulaperturate or fossaperturate, colpae bordered by distinct margos, os lalongate non-constricted, outer ends pointed, extends half the width of the grain. Surface psilate (Pl. 3, Figs. 31-33; Table 1).

(b) Desynaptic mutant of *C. annuum* var. cerasiformis: Shape: Subprolate, semiangular in polar view. Size: $23.42 \pm 0.22 \ \mu$ (P) × $20.00 \pm 0.21 \ \mu$ (E), PEI= $117.57 \pm 1.34 \ \mu$ and PAI= $10.83 \pm 0.97 \ \mu$. Structure: Exine tectate $1.30 \pm 0.02 \ \mu$ thick, ektexine thicker than endexine. A general thickening present near the apertures. In polar view the equatorial diameter $20.08 \pm 0.24 \ \mu$ m, mesocolpial diameter $13.61 \pm 0.14 \ \mu$ and apocolpial diameter $2.17 \pm 0.19 \ \mu$. Apertures: Mostly tricolporate, occasionally tetracolporate, angulaperturate or fossaperturate, inaperturate grains also observed. Margos indistinct, os lalongate, deeply constricted, extends half the width of the grain, outer ends pointed to slightly rounded. Surface psilate (Pl. 3, Figs. 34-36; Table 1).

Discussion

The pollen morphology of the species and varieties of Capsicum which were involved in the hybridization programme is rather fairly homogeneous. Generally the pollen of Capsicum is subprolate or prolate-spheroidal in shape (Erdtman, 1952; Murry & Eshbaugh, 1971; Basak, 1967; Nair & Kapoor, 1974; Raghvanshi, 1976). The shape of pollen grains in species and varieties currently involved in the hybridization work was found to be subprolate to prolate-spheroidal and oblate spheroidal (Aniel Kumar, 1984). In polar view the shape markedly varies with species and ploidy and is considered to be valuable as was observed by Muller (1969) in Sonneratia species and hybrids and also in interspecific hybrids of Physalis (Lydia Prasad et al., 1984). The pollen grains in the eight interspecific F1 hybrids of the present study showed irregular shapes due to reduced pollen fertility. Triangular and occasionally tetralobed grains were found in all the F_1 hybrids except in C. annuum var. cerasiformis \times C. chinense var. mishme hybrid. Shrunken, aberrant and inaperturate condition was found in two F_1 hybrids (C. annuum var. cerasiformis $\times C$. chinense var. mishme and C. chacoense \times C. chinense var. mishme). The shape of the pollen grains in the F_1 hybrids of C. annuum var. cerasiformis \times C. chinense var. mishme, C. chinense $\times C$. frutescens was found to be intermediate between the respective parents, while

in the remaining six F_1 hybrids (C. annuum var. $G3 \times C$. frutescens var. tabasco, C. annuum var. cerasiformis \times C. pendulum, C. frutescens var. tabasco \times C. annuum var. cerasiformis, C. chacoense \times C. frutescens var. tabasco and C. chacoense \times C. chinense var. mishme) they were generally prolate-spheroidal. In contrast, Muller (1969) has recorded the shape of the pollen grains in the F_1 hybrids of Sonneratia alba \times S. ovata and S. alba \times S. cascolaris as either dominant or recessive but never intermediate. He suggested that the shape subprolate is dominant over prolate-spheroidal type. Lydia Prasad et al. (1984) also encountered variation in pollen grains shape in the interspecific hybrids of Physalis.

Raw colchiloids C. annuum var. cerasiformis and C. annuum var. TNK generally suffer from low pollen production and reduced fertility compared to the corresponding diploid genotypes. Both tricolporate (30.00%) and tetracolporate (70.00%) grains were encountered in the two tetraploids. The shape of the pollen grains in them is subprolate, having a mean PEI of $121.35 \pm 1.15\mu$ and $116.30 \pm 0.99 \mu$ respectively. Similar variation in pollen shapes was recorded in Sisymbrium irio complex (Nair & Sharma, 1966-67). Shrunken and aberrant grains not uncommon in newly synthesized polyploids were encountered in the two tetraploids of the present study and also in the induced autotetraploid Physalis pubescens (Lydia Prasad et al., 1984).

Pollen grains of the desynaptic mutant were by and large subprolate; however, prolate-spheroidal and oblate-spheroidal grains were also recorded though in a low frequency thus exhibiting variation in pollen shapes. The occurrence of shrunken and aberrant grains in the interspecific hybrids, raw colchiploids and desynaptic mutant of the present study was attributed to the irregular meiosis and polyad formation. Similar variation in pollen shape was attributed to higher pollen sterility. Such variation in pollen shapes was reported in the induced mutants of *Pimpinella* (Joshi & Raghuvanshi, 1967) and *Capsicum* (Raghuvanshi, 1976).

Pollen size seems to be an important character in *Capsicum*. The size variation was more evident in all the eight F_1 hybrids of the present study showing micro and macro pollen grain formation. Similar micro and macro pollen grains were reported in the F_1 hybrid of *Amaranthus dubius* $\times A$. *spinosus* (Pal & Khoshoo, 1966), and the variation in the sizes of the pollen has been attributed to the presence of lagging univalents resulting in fewer chromosomes or more chromosomes in the microspores. The variation encountered in pollen sizes of interspecific hybrids and mutants of the present investigation may be attributed to the occurrence of univalents and irregular meiosis in them.

Larger size of the pollen grains often provides a reliable index for the detection of autotetraploids in the experimental populations. In the present study the pollen grains in induced autotetraploids of C. annuam var. cerasiformis and C. annuam var. TNK were found to be larger than those of the diploid progenitors. A test for significance revealed that they were highly significantly different from each other (Table 2). Though there was no overall correlation between the chromosome number and pollen size it is presumed that the plants with lower chromosome number have smaller pollen and increase in the chromosome number of the same taxa is generally accompanied by larger pollen and with more defectives, more apertural numbers, and changes in aperture disposition etc. Such variations in the pollen size that accompanied an increase in chromosome number was also recorded in Oryzi (Sampath & Ramanathan, 1951; Rangaswamy & Raman, 1973), Plantago (Basset & Crompton, 1968) and in Physalis pubescens (Lydia Prasat et al., 1984). Variation in pollen size ranging from small to large was encountered in the desynaptic mutant of the present study and this is attributed to irregularities pre-

Paired comparison	Polar diameter	Equatorial diameter	Polar equatorial index	Apocolpial diameter	Polar area index
C. annvum var. cerasiformis (diploid) and C. annuum var. cerasiformis (tetraploid)	44.30	28.21	4.28	4.68	4.95
C. annuum var. TNK (diploid) and C. annuum var. TNK (tetraploid)	36.98	19.66	11.39	6.19	0.87*

Table 2-Comparison of various pollen characters in the diploids and their corresponding tetraploids of Capsicum L. ('t' values)

*=Not Significant at p=0.05.

vailing in the genome. Similar variation in the pollen sizes were also reported in taxa treated with chemicals and physical mutagens such as in *Pimpinella* (Joshi & Raghuvanshi, 1967) and *Capsicum* (Raghuvanshi, 1976) respectively. Therefore, the variation in the pollen sizes observed in interspecific hybrids, tetraploids and the desynaptic mutant of *Capsicum* is attributed to the unstable nature of the newly synthesized plants.

The pollen size was measured along the polar axis. Based upon sizes the pollen grains may be placed either in small (10-25 μ) or in medium (25-50 μ) classes (Erdtman, 1952; Murry & Eshbaugh, 1971). Accordingly six out of the eight F₁ hybrids fit into the small class whose mean ranges from 22.24 to 24.80 μ . The remaining two F₁ hybrids (*C. annuum* var. cerasiformis x *C. pendulum* and *C. chacoense* x *C. frutescens* var. tabasco) and the two tetraploids of *C. annuum* varieties cerasiformis and *TNK* and one desynaptic mutant come under medium class with mean ranges of 25.53 to 38.19 μ .

The relative size of the polar area based on polar area index (PAI) has been suggested by Faegri and Iverson (1954) and Murry and Eshbaugh (1971) as a quantitative character of diagnostic value. On the basis of the polar area index the following two classes are suggested pertaining to the taxa of the present study.

- (1) smaller polar area index (PAI) between 10-15 μ , eg: desynaptic mutant, C. chacoense x C. annuum var. BOB and C. chinense x C. frutescens.
- (2) Medium size polar area index (PAI) between 15-20 μ , eg: two induced autotetraploids of *C. annuum* varieties of cerasiformis and *TNK* and six interspecific hybrids, viz., *C. annuum* var. G3 x *C. frutescens*, *C. annuum* var. cerasiformis x *C. pendulum* and *C. chinense* var. mishme, *C. frutescens* var. tobasco x *C. annuum* var. cerasiformis, *G. chacoense* x *C. frutescens* var. tobasco and *C. chacoense* x *C. chinense* var. mishme.

In all the interspecific hybrids, two induced autotetraploids and one desynaptic mutant, the PAI was less than that of the corresponding parents due to syncolporate condition. Similar results were reported in the induced autotetraploid, interspecific hybrids and the mutants of *Physalis* (Lydia Prasad *et al.*, 1984). Smaller PAI due to syncolporate condition was also observed in some species of *Capsicum* (Murry & Eshbaugh, 1971) and in the induced mutants of *C. annuum* (Raghuvanshi, 1976).

The higher number of apertures and the occasional inaperturate condition observed in the induced autotetraploids, desynaptic mutant and interspecific hybrids could be attributed to the meiotic irregularities in the newly synthesized genome. The multiaperturate and occasional inaperturate condition was also reported in the interspecific hybrids, induced polyploids, and polyhaploid of *Physalis* (Lydia Prasad *et al.*, 1984) and polyhaploids of *Hordeum* (Rajendra *et al.*, 1978), *Solanum* species (Erdtman, 1952; Anderson & Gensel, 1976) and induced mutants of *Capsicum annuum* (Raghuvanshi, 1976). Injecting colchicine to the flower buds Dover (1972) was able to induce multiaperturate condition in *Triticum aestivum* containing alien addition chromosomes in the genomes.

The endoaperture (os) was found to be lalongate, deeply constricted to non-constricted nature and extending half to three fourths of the width in all the eight F_1 hybrids, two autotetraploids and the desynaptic mutant. Irregular os with tapering ends was recorded in 4 F_1 hybrids of the present study (C. annuum var. G3 x C. frutescens, C. annuum var. cerasiformis x C. pendulum, C. chacoense x C. frutescens var. tabasco and C. chacoense x C. chinense var. mishme). Irregular os with outer ends rounded was observed in the F_1 hybrids of C. annuum var. cerasiformis x C. chinense var. mishme, two autotetraploids of C. annuum and the desynaptic mutant. Circular os was observed in C. frutescens var. tabasco x C. annuum var. cerasiformis hybrid and bifurcated os was observed in C. chinense x C. frutescens and C. chacoense x C. annuum var. BOB hybrids. Irregular os formation could be attributed to the meiotic irregularities and higher pollen sterility present in the genome. Similar irregular os was also reported in interspecific hybrids and induced polyploids of Physalis (Lydia Prasad et al., 1984), species hybrids of Sonneratia (Muller, 1969), induced mutants of C. annuum (Raghuvanshi, 1976) and in some species of Solanum (Sharma, 1974).

Analysis of variance was also employed in the eight F_1 hybrids for quantitative meassurements utilising parameters such as polar diameter (P) (F=79.93P>0.01), equatorial diameter (E) (F=65.23, P>0.01), polar equatorial index (PEI) (F=6.70, P>0.01), apocolpial diameter (A) (F=27.48, P>0.01), and polar area index (PAI) (F=10.30, P>0.01). The 'F' values indicated highly significant differences among them.

Inter-relationships

The inter-relationships between the five species of *Capsicum* have been explored through the crossability relationships and a cytogenetic analysis of the eight F_1 interspecific hybrids. Hybrids were obtained easily in the following combinations, viz., *C. frutescens* var. tabasco x *C. annuum* var. cerasiformis, *C. annuum* var. cerasiformis x *C. pendulum*, *C. annuum* var. cerasiformis x *C. chinense* var. mishme, *C. chacoense* x *C. annuum* var. BOB and *C. chinense* x *C. frutescens*. Cytogenetic studies revealed that the hybrids differ from each other at least in one or two translocations in their genomes. They are partly pollen fertile indicating that there are considerable homologies or homologies among the genomes of the respective parents and hence they are probably closely related to each other. The pollen morphological evidence deduced through the application of the two parameters (PEI and PAI) also supports the cytogenetical evidence.

Evolutionary significance

In general the plant morphologists agree that similarity in a large number of characters of different taxa indicate close relationships, whereas diversity among the characters shows distant relationship. In recent years a few palynologists like Erdtman (1952, 1961), Saad (1961), Nair (1965), Punt (1971), Sharma (1969) and Muller (1969) have attempted to trace the evolutionary significance of some pollen morphological characters. In the genus *Capsicum* it is difficult to express any definite opinion about the evolutionary significance of pollen morphological characters. However, the present study highlights that some pollen characters showed evolutionary significance in species hybrids, induced autotetraploids and desynaptic mutant than in the respective parents.

- (1) Shape: Prolate-spheroidal grains are considered to be advanced over the subprolate, eg: All the eight F_1 interspecific hybrids.
- (2) Multiaperturate condition is an advanced character over tricolporate eg: induced autotetraploids of C. annuum, desynaptic mutant and F₁ hybrids of C. annuum var. cerasiformis x C. pendulum, C. chinense x C. frutescens, C. chacoense x C. annuum var. BOB.
- (3) Circular os is considered to be an advanced feature over bifurcated and tapering outer ends of the os,
 eg: F₁ hybrid of C. frutescens var. tobasco x
 C. annuum var. cerasiformis.
- (4) Bifurcated os is advanced than tapering outer ends of the os, eg: F_1 hybrids of C. chinense x C. frutescens and C. chacoense x C. annuum var. BOB.

The utilization of the above pollen morphological characters for the purpose of interpreting the relative advancement and the primitiveness of the taxa revealed that the interspecific hybrids, induced autotetraploids and desynaptic mutant showed advanced features over the corresponding parental species and varieties.

Acknowledgements

We express our thanks to Mr K. Prakasa Rao for his assistance in photographic work. Two of us (OAK and RCP) are grateful to the C. S. I. R. and U. G. C. New Delhi for providing the financial assistance.

References

- ANDERSON, G. J. & GENSEL, P. G. (1976). Pollen morphology and the systematics of Solanum., section Basanthrum. Pollen Spores, 18: 533-552.
- ANIEL KUMAR, O. (1984). Cytogenetical and palynological studies in the genus Capsicum. Ph.D. Thesis. Andhra University, Waltair.
- BASAK, R. K. (1967). Pollen grains of Solanaceae. Bull. Bot. Soc. Bengal, 21(1): 49-58.
- BASSET, I. J. & CROMPTON, C. W. (1968). Pollen morphology and chromosome numbers of the family Plantaginaceae in North America. Can. J. Bot., 46: 349-361.
- DOVER, G. A. (1972). The organisation and polarity of pollen mother cells of Triticum aestivum. J. Cell Sci., 11: 697-711.
- ERDTMAN, G. (1952). Pollen morphology and plant taxonomy. An Introduction to Palynology I. Angiosperms. Chronica Botanica Company, Waltham, Massachusetts.
- ERDTMAN, G. (1961). Pollen wall and angiosperm phylogeny. Recent advances in Botany, I: 675-678 (Canada).
- ERDTMAN, G. (1969). Hand Book of Palynology, Morphology, Taxonomy and Ecology. An Introduction to the study of pollen grains and spores. Hafner Publishing Co., New York.
- FAEGRI, K. & IVERSON, J. (1964). Text book of pollen analysis. Hafner Publishing Company, New York.
- JOSHI, S. & RAGHUVANSHI, S. S. (1969). Studies on chemically induced change in expression of gene (S) controlling the pollen snapers in *Pimpinella monoica*. Grana Palynol. 7: 378-384.
- LYDIA PRASAD, Y., RAJA RAO, K. G. & RAMANUJAM, C. G. K. (1984). Pollen morphology of some diploid, polyploid species and hybrids of *Physalis* L. *New Botanist*, **11**(2) : 145-160.

- MURIOZIO, A. (1956). Pollengestaltung bee einigen polyploiden kulturpflanzen. Granz Palynol., 1(2): 59-69.
- MULLER, J. (1969). A palynological study of the genus Sonneratia (pollen variability in five species and two hybrids of Sonneratia). Pollen Spores, 11: 223-298.
- MURRY, L. E. & ESHBAUGH, W. H. (1971). A palynological study of the Solainae (Solanaceae). Grana, 11: 65-78.
- NAIR, P. K. K. (1961). Pollen grains of cultivated plants-II. Bougainvillaea, Hibiscus and Euphorbia. J. Indian bot. Soc., 40: 365-381.
- NAIR, P. K. K. (1965). Trends in morphological evolution of pollen and spores. J. Indian bot. Soc., 44: 468-478.
- NAIR, P. K. K. (1970). Pollen morphology of Angiosperms (A historical and phylogenetic study). Scholar Publishing House, Lucknow.
- NAIR, P. K. & KAPOOR, S. K. (1974). Pollen morphology of Indian vegetable crops, in Glimpses in Plant Research, vol. II.
- NAIR, P. K. K. & SHARMA, M. (1966-67). Cytopalynological observations on the Sysymbrium irio complex.
 J. Palynol., 2 & 3:33-40.
- PUNT, W. (1971). Pollen morphology of the genera Norantia, Souroubea and Rhyschia (Marcgraviaceae). Pollen Spores, 13: 199-232.
- RAGHUVANSHI, R. K. (1976). Palynological studies in Capsicum L. J. Palynol., 12: 81-86.
- RAJENDRA, B. R., TOMB, A. S. MUJEEB, K. A. & BATES, L. S. (1978). Pollen morphology of selected Triticeae and two intergeneric hybrids. *Pollen Spores*, **20**: 145-156.
- RANGASWAMY, S. R. & RAMAN, V. S. (1973). Pollen production in diploids and autotetraploids of Rice (Oryza sativa L). Pollen Spores, 15: 189-193.
- SAAD, S. I. (1981). Phylogenetic development in the apertural mechanism of *Linum* pollen grains. *Pollen Spores*, 3: 33-43.

SAMPATH, S. & RAMANATHAN, K. (1951). Pollen grain sizes in Oryza. J. Indian bot. Soc., 30: 40-48.

- SHARMA, B. D. (1969). Studies of Indian pollen grains in relation to plant taxonomy-Sterculiaceae. Proc. Nat. Inst. Sci. India, 35B(4): 320-359.
- SHARMA, B. D. (1974). Contribution to the palynotaxonomy of the genus Solanum Linn. 7. Palynol., 10(1): 51-68.

Explanation of Plates

(For all figures, magnification bar represents 10 μ)

PLATE 1

- 1-3. Pollen grains of C. annuum var. $G3 \times C$. frutescens var. tabasco
- 1. Equatorial view showing well separated colpae and mesocolpium.
- 2. Equatorial view showing ill-defined Os.
- 3. Polar view showing margos.
- 4-6. Pollen grains of C. annuum var. cerasiformis \times C. chinense var. mishme
- 4 Equatorial view showing colpae.
- 5. Equatorial view showing irregular, constricted Os.
- 6. Polar view showing well developed margos.
- 7-9. Pollen grains of C. annuum var. cerasiformis \times C. pendulum
- 7. Equatorial view showing colpae.
- 8. Equatorial view showing nonconstricted Os.
- 9. Polar view showing margos.
- 10-12. Pollen grains of C. frutescens var. tabasco \times C. annum var. cerasiformis

10. Equatorial view showing colpae.

- 11. Equatorial view showing deeply constricted circular Os.
- 12. Polar view showing syncolporate condition.



Kumar et al.-Plate 1

















18





23





PLATE 2

- 13-15. Pollen grains C. chinense var. mishme \times C. frutescens var. tabasco
- Equatorial view showing well separated colpae. 13.
- Equatorial view showing irregular, bifurcated Os. 14.
- Polar view showing tetracolporate condition. 15.
- Pollen grains of C. chacoense \times C. annuum var. BOB 16.18.
- Equatorial view showing well separated colpae. 16.
- Equatorial view showing irregular, constricted Os. 17.
- Polar view showing syncolporate condition. 18.
- Pollen grains of C. chacoense \times C. frutescens var. tabasco 19-21.
- Equatorial view showing colpae. 19.
- 20.Equatorial view showing constricted Os.
- Polar view showing margos. 21.
- 22-24. Pollen grains of C. chacoense \times C. chinense var. mishme
- 22. Equatorial view showing colpae.
- 23.Equatorial view showing long narrow irregular Os.
- 24. Polar view showing syncolporate condition.

PLATE 3

- 25-30. Pollen grains of induced autotetraploid of C. annuum var. TNK 25.
- Equatorial view showing enlarged colpae. 26.
- Equatorial view showing irregular constricted Os. 27.
- Polar view showing distinct margos. 28.
- Polar view showing pores situated between the angles. 29.
- Polar view showing irregular shape. 30.
- Polar view showing tetracolporate condition. 31-33.
- Pollen grains of induced autotetraploid of C. annuum var. cerasiformis 31.
- Equatorial view showing colpae 32.
- Polar view showing pores situated between the angles. 33.
- Polar view showing well developed margos. 34-36.
- Pollen grains of induced desynaptic mutant of C. annuum var. cerasiformis 34. Equatorial view showing colpae.
- 35.
- Equatorial view showing irrgular deeply constricted Os. Polar view showing margos. 36.