

POLLEN MORPHOLOGY AND TAXONOMY OF THE TRIBE SOPHOREAE (FABACEAE)

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Abstract

Pollen morphology of 40 species belonging to 4 genera of the tribe Sophoreae (Fabaceae) has been studied. Generic delimitations and interspecific relationship within the tribe have been assessed. Palynological findings have been correlated with related tribes of the family for better palynotaxonomical and phylogenetic understanding of the group.

Introduction

The Tribe Sophoreae comprises 48 genera (Hutchinson, 1964); Airy Shaw, 1973) and is taxonomically characterised by free stamens, non-jointed pods and odd pinnate to rarely simple leaves in addition to other common fabaceous features. Bentham (1865) treated the tribe next to Dalbergieae and placed as the last tribe of Papilionaceae before Caesalpineae.

Sophoreae is represented by 5 genera (s. l.) in India, viz., *Dalhousia* R. Grah, *Sophora* Linn., *Calpurnia* E. Mey., *Pericopsis* Thw. and *Ormosia* Jack. Except *Pericopsis* which is represented in India by single species *P. mooniana* Thw. all the other genera have been investigated.

The pollen morphology of the tribe Sophoreae has been meagrely worked out by Mohl (1835), Hassall (1842), Cranwell (1942), Selling (1947), Vishnu-Mitre and Sharma (1962), Huang (1968, 1972), Heusser (1971) and Ohashi, (1975) mostly in relation to regional flora. Recently Guinet (1981) and Ferguson and Skvarla (1981) gave only a synoptic information on the pollen morphology of the Leguminosae in general. Considering the lack of palynological informations of the tribe an attempt has been made to present a detail palynological data on the Indian and available foreign materials with emphasis on pollen morphology and its application in taxonomy.

Material and methods

The polliniferous material was collected from the authentic herbarium sheets of the Central National Herbarium (CAL). Pollen slides were prepared by acetolysis method (Erdtman, 1952) and have been deposited in the sporothek of palynological laboratory, CAL. For measurements, mean of 25 readings have been taken from acetolysed non-chlorinated pollen grains for each taxon. Terminology of Erdtman (1969) and Faegri and Iversen (1964) have been followed. Salient pollen morphological feature have been given in the Table 1. Only latest valid names have been cited in the table and basionyms as well as synonyms are given for reference in the list of materials studied. New names (*sensu stricto*) with the previous names are given in the text for better understanding of the taxon from palynotaxonomical point of view.

Material studied

Calpurnea aurea Baker, India: M. P., Panchmari, V. Narayanaswami—3503. Africa: Schimperi inter Alyssinicum. u. i.—1840, CAL—133241. *C. floribanda* Harv., S. Africa Peddie dist, R. D. A. Bayliss—5602. *C. intrusa* E. Mey, S. Africa: T. Cooper—856 (1862), Inanda, J. M. Wood—956. *C. lasiogyna* E. Mey, Africa Herb. Natal Bot. Gard. J. M. Wood—102; Natal,

Table 1—Summary of pollen morphological features of the tribe *Sophoreae* (Fabaceae)

Taxa	Outline in eq.view (contour)	SIZE	EXINE		Type	Ecto.	Endo.	Remarks		
			PXE μ_m	Thickness Mean	Pat.	Lumina size	(Colpus)	(Os=Pore)		
1	2	3	4	5	6	7	8	9	10	11
<i>Calpurnia aust.</i> Baker	Fro.Sph.	Comp.oval $22.5-31 \times 23-30$ 27×26	1-1.5	ret.	± 1	3-colpr.	slit.	lolong	Sex. thicker than nex., Ex. thicker at apocolpium. Endoap. rectangular type, foot layer and col. heads distinct.	
<i>C. floribunda</i> Harv. Pro.	Ellip.oval $23-31 \times 18-24$ 26.5×21	1.5	Obs.	—	3-colpr.	slit.	lolong	Sex. =nex.		
<i>C. intrusa</i> E. Mey Pro.	Ellip.oval $22-29 \times 16-23$ 35×19	1-1.2	Psi.	—	3-colpr.	slit.	lolong	Colpus constricted at equator, sex =nex., Ex. thicker at apocolpium.		
<i>C. lasiogyna</i> E. Mey Pro. Pr.Sph	Comp.oval $22.5-30 \times 22-28$ 26.5×25	1.5	ret.	1-1.25	3-colpr.	slit.	lolong	Sex.=nex., colpus constricted at the equator, ret. distinct at apocolpium.		
<i>C. sylvatica</i> E. Mey Pro.	Comp.oval $26-33 \times 24-32$ 31×28.5	± 2	ret.	1.2-1.5	3-colpr.	tap.	lolong	Sex. thicker than nex., eq. margin of the endoap. indistinct.		
<i>Dalhousia africana</i> S. Moore	Pro.Sph.	Comp.oval $28-36 \times 26-34$ 34×32.5	2	ret.	± 1	3-colpr.	tap.	Colpus margin smooth, endoap. granulated, margin indistinct, sex. thicker than nex.; foot layer, col. and col. heads distinct.		

(Table 1—Contd.)

	1	2	3	4	5	6	7	8	9	10	11
<i>Dalhousia bracteata</i> Grah. ex tenth.	Pro.Sph. -Prc.	Comp.oval $18\text{-}23 \times 19\text{.8}$ 20×20.5	micr-ret.	± 0.5	3-colpr.	tap.	lalong	Copulus margin irregular, endo-ap. margin indistinct, sex. = nex., col. distinct with distinct heads, lumina size in apocolpium $\pm 1/\mu\text{m}$.			
<i>Fedorovia laxa</i> (Prain) Yakovlev	Pro.sph. -Prc.	27-35 \times 23.5-33 31×29	obs.	—	3-colpr.	tap.	lalong	Copulus constricted at equator, sex. thicker than sex. at apocolpium, col. indistinct.			
<i>F. striata</i> (Dunn.) Yakovlev	Pro.-sph.	Gomp.oval $24\text{-}32 \times 21\text{-}30$ 28.5×26	psi.	—	3-colpr.	tap.	lalong	Copulus constricted at equator, sex. thicker than sex. at apocolpium, col. indistinct.			
<i>Ormosia assamica</i> Yakovlev	Pro.	Ellip.oval $31\text{-}38 \times 27\text{-}35$ 36×32	ret.	1.5-2	3-colpr.	tap..	lalong	Pollen grains comparatively larger than the other spp. of the genus. Col. distinct. Double layer of columellae observed. Endoap. rectangular.			
<i>O. bancana</i> (Fig.) Merr.	Pro.Sph.	Comp.oval $26\text{-}34 \times 25\text{-}32$ 30×28.5	ret.	± 1	3-colpr.	tap.	lalong	Sex. thicker than nex., endo-ap. rectangular type, equatorial margin indistinct, copulus constricted at the equator.			
<i>O. dulce</i> Prain	Pro-Pro. sph.	Ellip.oval $23.5\text{-}32 \times 21\text{-}27$ 28×23.5	ret.	1	3-colpr.	tap.	lalong	Sex. = nex. Copulus constricted at the equator. Col. distinct.			

Table 1—(Contd.)

1	2	3	4	5	6	7	8	9	10	11
<i>O. emarginata</i> Benth.	Prc. Sph.	Comp.oval	25.34×23.31	2	ret.	1-1.5	3-colpr.	tap.	lolong	
			28.5×26							
<i>O. hoensis</i> Prain	Pro.	Ellip.oval	21.5-31×21-27	1.75-2	ret.	1	3-colpr.	slit.	lolong	
			27×23							
<i>O. macrodiscia</i> Baker	Pro.-Sph.	Comp.oval	21-27×21-26	1.75	micro-ret.	0.5-0.75	3-colpr.	tap.	lalong	
			24×22.5							
<i>O. microsterna</i> Baker	Pro. Sph.	Comp.oval	23-32×23-30.5	2	f. ret.	0.75-1	3-colpr.	tap.	lalong	
			29×28							
<i>O. pinnata</i> (Lour.) Merr.	Pro.	Ellip.oval	26-33×24-31	2	ret.	1-1.5	3-colpr.	tap.	lalong	
			29.5×25.5							
<i>O. baliita</i> Prain	Pro.-Pro. sph.	Comp.oval	24.5-33×26-32	1.5-2	f. ret.	0.75-1	3-colpr.	tap.	lolong	
			29.5×27.5							
<i>O. scandens</i> Prain	Pro-sph.	Comp.oval	26.5-34×26-32.5	1.5	f. ret.	0.75	3-colpr.	slit.	lolong	
			30.5×28							
<i>O. sumatrana</i> (Miq.) Prain	Poo.	Ellip.oval	24.31×21.30	1.5	obs.	—	3-colpr.	slit.	lolong	
			27+23.5							

Sex.=nex., colpus constricted at the equator. Eq. margin of the endoap. indistinct. Lumina larger at poles.

Sex. thicker than nex. Ex. thicker at poles. Endoap. o. nate type.

Sex.=nex., ret. finer towards aperture, endoap. rectangular.

Sex.=nex. colpus constricted at the equator. Eq. margin of endoap. indistinct.

Sex.=nex., ret. heterobrochate, finer towards aperture. Colpus constricted at the equator. Col. distinct.

Sex. thicker than nex., Ex. thin at mesocelium, lumina larger at poles and finer towards aperture.

Endoap. area granulated.

Endoap. area indistinct.

Table 1—(Contd.)

1	2	3	4	5	6	7	8	9	10	11
<i>S. japonica</i> Linn. var. <i>japonica</i>	Pro.	Ellip.oval $23\text{-}30 \times 21\text{-}25.5$ 26×23	1.5-2 $24.5\text{-}31 \times 21\text{-}29$ 26×23.5	ret.	1-1.25 ± 1	3-colpr.	tap.	1olong		
<i>S. littoralis</i> Schrad	Pro.	Ellip.oval $24.5\text{-}31 \times 21\text{-}29$ 26×23.5	1.5-2 30×27.5	ret.	3-colpr.	tap.	1olong			
<i>S. mollis</i> Grah. var. <i>griffithii</i> (Stock) Tsoong	Pro.-pro. sph.	Ellip. oval $21\text{-}30.5 \times 21\text{-}27$ 27.5×24	± 2	ret.	1-1.25 1.5	3-colpr.	slit	1olong		
<i>S. mollis</i> Grah. var. <i>mollis</i>	Pro.-Pro. sph.	Ellip.oval $25.5\text{-}32 \times 23\text{-}29$ 30×27.5	± 2	ret.	1-1.5 1.5	3-colpr.	slit	1olong		
<i>S. macrostoma</i> Benth.	Pro.- Pro. sph.	Comp.oval $23\text{-}30.5 \times 22\text{-}28.5$ 27×25.5	1.5 1.5	ret.	1-1.25 1.5	3-colpr.	slit	1olong		
<i>S. pachycarpa</i> Schrenk ex C. A. Mayer	Pro.sph.	Comp.oval $(17\text{-})24\text{-}32 \times 22\text{-}30$ 30×28.5	2	ret.	1-1.75 1.75	3-colpr.	slit	1olong		
<i>S. prazeri</i> Prain var. <i>prazeri</i>	Pro.sph.	Comp.oval $22\text{-}28 \times 19\text{-}23$ 25.5×23	± 2	f. ret.	± 1	3-colpr.	tap.	1olong		
<i>S. tomentosa</i> Linn. var. <i>tomentosa</i>	Pro.	Comp.oval $26\text{-}35 \times 18\text{-}29$ 31.5×26.5	1.5 1.5	f. ret.	0.5-0.75 $0.5-0.75$	3-colpr.	slit	1olong		
										Endoap. rectangular type, eq. margin indistinct, heterobrochate.

Sex.=nex., Ex.
thicker at
poles, col.
indistinct.

Sex.=nex., col.
indistinct at
mesocolpium,
colpus consti-
tuted at the
equator.

Sex.=nex., lumina larger
at poles.
Pollen grain size
is the only
difference with
other var. studied.

Sex.=nex., ccl.
indistinct,
lumina size
larger at poles.

Sex.=nex., Col.
indistinct at
mesocolpium,
lumina larger
at poles. Rarely
ob. sph. grains
observed.

Sex. thicker than
nex., colpus con-
stricted at the
eq., eq. margin
of the endocap.
indistinct.

Table 1—(Contd.)

	1	2	3	4	5	6	7	8	9	10	11
In the aperture area nexine departed from the sexine and intruded inward forming vestibulum (Faegri and Iversen, 1964).											
<i>S. velutina</i> Lin. var. <i>velutina</i>	Pro. Pro. Sph.	Comp. oval	21-26 × 18-24.5 <u>24 × 22</u>	1-1.5	rug.- ret.	(1)-1.5	3-colpr.	tap.	lolong	Colpus constri- cted at the eq., endoap. area gr., rarely gr. extends through colpus length, Sex.=nex. ret. finer tow- ards aperture.	Sex.=nex., colpus constricted at the eq.
<i>S. wightii</i> Baker	Pro.	Ellip. oval	24-29.5(-36.5) × 19.5-23.5(-31) <u>26 × 22</u>	2	f. ret	± 1	3-Colpr.	tap.	lolong	Colpus constri- cted at the eq., endoap. rectang- ular type, Sex. thicker than nex., hetero brochate, ret. finer towards aperture. Ap. membrane gr.	Sex.=nex., Col. indistinct. Ex. thicker at poles. Some sma- ller grains also observed.
<i>Trichocarymus pachycarpa</i> (Camp. ex Benth.) Yakovlev	Pro.- Pro.sph.	Comp. oval	26.5-35 × 24-32 <u>31 × 28</u>	2.5	ret.	(0.75-) 1-1.5	3-colpr.	tap.	lolong	Double layer of exine stratification is observed.	
<i>Vicia alopaeoides</i> (Spach. ex Jaub. et Spach.)	Pro.- Pro.sph.	Ellip.oval	(18-) 23-31 × 21- <u>29.5</u>	± 2	ret.	1-1.5	3-color.	slit	lolong		
Slit like Sph..—Spheroidal; Tap.—Tapering; var.—variety.											

Abbreviations used:

Ap.—Aperture; Col.—collumella; comp. oval—compressed oval; colpr.—colporate; Ellip.—Elliptic oval; Eq.—Equatorial diameter; Ectoap.—Ectoaperture; Endoap.—Endoaperture; f.—finesely reticulate; f. ret.—equatorial; f. ret.—finely reticulate; gr.—grains granulated; laalong.—laalongate; lolong.—lolongate; Micro-ret.—Micro-reticulate; nex.—nexine; obs.—obscure; Ob.—Oblique; P.—Polar axis length; Pat.—Pattern; Pro.—Prolate; Pro. Sph.—Prolate-spheroidal; Psi.—Psilate; ret.—reticulate; Sex.—Sexine; Slit.—Slit like Sph..—Spheroidal; Tap.—Tapering; var.—variety.

J. M. Wood—6125. *C. sylvatica* E. Mey,
Africa: Natal, R. Schlechter—6158.
Dalhousia africana S. Moore, Africa:
Congo C. Evrad—5040, CAL—567580. *D.
bracteata* Grah. ex Benth., India: Herb.
Hort. Cal., J. C. Prazer—75, CAL—133783;
Assam, Khasia, Jenkins—262. Bangladesh:
Sylhet, NIL—7162, (24-5-1868), CAL—
133181.

Fedorovia laxa (Prain) Yakovlev = *Ormosia laxa* Prain, Burma: Kachin Hills, Shaik Mokim, s. n. CAL-133679. *F. striata* (Dunn) Yakovlev = *Ormosia striata* Dunn., China: Yunan, Aug. Henry-41886.

Ormosia assamica Yakovlev = *O. robusta* Baker, Assam: Badal Khan—10; NIL, CAL—133806. *O. bancana* (Miq.) Merr. = *O. parviflora* Baker. Malaya Peninsula: NIL, s. n., Det.—Yakovlev (1967). *O. dulse* Prain, Burma: Shaik Mokim—s. n. Det.—Yakovlev (1967). *O. emerginata* Benth. Hongkong: B. Garden, NIL, s. n. CAL—133675. *O. hoaensis* Prain, Cochin—China: L. Pierre, s. n., CAL—133680. *O. macrodisca* Baker, Singapore: HNR—2103. *O. microsperma* Baker, Malacc: R. Derry—1090. *O. pinnata* (Lour.) Merr. = *O. hainanensis* Gagnep. China: Hainan, F. C. How—72995. *O. polita* Prain = *O. nitida* Prain, Burma: Shaik Mokim, s. n. Det. Yakovlev (1967). *O. scandens* Prain, Malay peninsula: Dr. King's Col.—3560. *O. sumatrana* (Miq.) Prain, China: Yunnan, Aug. Henry—12885. Det.—Yakovlev (1967).

Placolobium gracilis (Prain) Yakovlev =
Ormosia gracilis Prain, Malay peninsula:
 Perak, King's Col.—4234. *P. travancorica*
 (Bedd.) Yokovlev = *Ormosia travancorica* Bedd.
 Tamil Nadu: Tinnevelly, C. A. Barber—
 3127, Det.—G. S. Gamble.

Ruddia fordiana (Oliv.) Yakovlev—*Ormosia fordiana* Oliv. China: Hainan, F. C. How
—72898.

Sophora ardexa Grah. America: Ex. Horto. bot. Petropolitani, s. n. CAL—133547. *S. benthami* V. Steen.=*S. acuminata* Benth. ex Baker, Meghalaya: *G. gallatly*—258. Sikkim: H. F. Green—835. *S. flaves-cens* Ait. var. *flavescens*=*S. flavescens* Ait; *S. angustifolia* S. & Z.: *S. flavescens* var. *Stenophylla* Hayata; *S. angustifolia* S. & Z. var. *senophylla* Mak. et Nemoto. Japan: Yokohama, Ex. Herb. Bot. Petropolitani, CAL—133475; T.bet: Tali vally, George Forest—4233; China: Zimmermann—201—*S. heptaphylla* Linn. Srilanka: C. P. Tha-waites—570; Herb. Sulp. Kurz—s. n.

CAL—133333. *S. interrupta* Bedd. Madras: Beddome, s. n. CAL—133458, Det. J. G. Gamble (1917). *S. japonica* Linn. var. *japonica*=*S. japonica* L. Tamil Nadu: Yarkand, J. Scully—s. n. CAL—133490. China: Yunnan, Fr. Ducloux—7617. *S. littoralis* Schrad, Australia: Communicatum ereliquis Martianis (1875), Aquisitions —Journal Nr. 229, CAL—133580. *S. mollis* Grah. var. *griffithii* (Stock) Tsoong—*S. griffithii* Stocks, *S. mollis* subsp. *griffithii* (Stock) Ali, *Keyserlingia griffithii* (Stock) Bunge ex Boiss. Baluchistan: J. F. Duthie—8642. *S. mollis* Grah. var. *mollis*, N. W. Himalaya—H. B. Royle, s. n. CAL—133427; Afghanistan: H. A. Deane, s. n. Herb. Hort. Bot. Calcuttensis. *S. moocroftiana* Benth. Tibet: King's Col.—I, CAL—133280. *S. pachicarpa* Schrenk ex C. A. Mayer, Afghanistan: J. E. T. Aitchinson—435. *S. prazeri* Prain var. *prazeri*, Burma: Gilbert Rogers—1043; Abdul Khalil—s. n. (1896), CAL—133467, *S. tomentosa* Linn. var. *tomentosa*=*S. tomentosa* Linn. Australia: Ferd Mueller, s. n. CAL—133564. *S. velutina* Lindl. var. *velutina*=*S. glauca* Lesch. Madras: Coimbatore, C. E. C. Fischer—1056, Det.—G. P. Yakovlev (1967). *S. wightii* Baker—*S. heptaphylla* Auct. non Linn. Wight, Maharashtra: Herb. Hort. Bot. Calcuttensis, NIL, 273B (1879).

Trichocyamus pachycarpa (Camp. ex Benth.) Yakovlev = *Ormosia pachycarpa* camp. ex Benth. China: NIL. CAL—133684.

Vexibia alopecuroides (Spach. ex Jaub. et Spach) Yakovlev—*Sophora alopecuroides* Linn. Baluchistan: Stocks—1001, CAL 133390.

Observation and discussion

Pollen morphology of Sophoreae reveals that it is a stenopalynous tribe. Pollen isopolar, 3 zonocolporate, medium sized (exceptionally small), prolate or prolate spheroidal (very rarely oblate spheroidal) in equatorial view, polar axis length ranging from 18 to 38.5 μm and equatorial axis diameter ranging from 18-36 μm , either compressed oval or elliptic oval in meridional forms, circular in polar view. Colpi distinct, either narrow slit like or tapering. When tapering sometimes with a median constriction at the equator. Colpus membrane generally smooth, sometimes granulated in the endoaperture area and rarely granules extend through the colpus length. OS generally distinct, either lolon-

gate or lalongate type. When lalongate sometimes it is rectangular. Sometimes equatorial margin of the endoapertures are indistinct. Endoaperture area sometimes provided with randomly distributed granules. In *Ormosia hoensis* endoaperture is ornate type. Exine ornamentation in the tribe is generally reticulate type—either microreticulate or coarsely reticulate (Praglowski *et al.*, 1973); rarely psilate, obscure or ruguloreticulate. Lumina homobrochate or heterobrochate, when heterobrochate, finer towards aperture and/or coarser at apocolpium region. Exine layers differentiated into tectum, columella and endexine (Erdtman, 1969) (=sexine and nexine—Faegri & Iversen 1964)). In most of the species foot layer is not distinguishable. Exine thickness varies from 1 to 2.5 μm in the tribe. Generally exine is of uniform thickness, sometimes thicker at apocolpium than the mesocolpium region. Columella in most of the species of *Sophora* are indistinct, but distinct with distinct columella heads in the species of *Ormosia Calpurnia* and *Dalhousia*. In most of the taxa sexine is as thick as nexine, but sometimes sexine is thicker than nexine. Exine stratification in the tribe is important because of the fact that in some species of *Ormosia* (s.l.) double layer of columellae are evident. This observation reminds the similar observation of Ferguson and Skvarla (1981) in *Castanospermum* (Leguminosae). A detail SEM observation on the exine stratification of the Tribe, preferably from the ontogenetic point of view, is necessary for the knowledge of such anomalous double layer exine stratification. In *Sophora tomentosa* Linn. var. *tomentosa*, nexine splitted from the sexine in the endoaperture area and bend inward forming a vestibulum (Faegri & Iversen, 1964) which is a specialized pollen character rarely observed in legumeous taxa. Detailed pollen morphological characters of individual species studied are given in the Table 1.

Palynological trends in the tribe Sophoreae

Shape (oblate spheroidal)—Prolate spheroidal—Prolate (rare)
Size (Small) (rare)—Medium—Large
Aperture—Exo: Slit like—Tapering—Tapering, constricted at the equator.
Endo: (Circular)—Lolongate—Lalongate.

Membrane: Smooth—Granulated—random—Ornate.

Exine ornamentation—Psilate/Obscure—Reticulate

- a) finely reticulate/microreticulate (lumina upto 1 μm)
- b) Coarsely reticulate/(Lumina greater than 1 μm)
- c) ruguloreticulate

Columella layer—Single layer—double layer (most anomalous). Indistinct distinct, with distinct head

Palynotaxonomic consideration and affinities

Review of taxonomic literature (Yakovlev, 1972, 1973a, b, 1978; Tsoong, 1980; Tsoong & Machi, 1981) reveals that the nomenclature and identity of a number of taxa of the tribe have been changed from time to time by the taxonomists. As a result 4 genera and 40 species (*sensu lato*) studied palynologically are now validated as 9 genera and 40 species (*sensu generis* and species nova). Three species of *Ormosia* have been given the generic status by Yakovlev, viz., *Trichcyamus*, *Placolobium* and *Ruddia* but palynology has nothing to do in support of the creation of new taxa except for *Trichocynamus pachycarpa* (camp. ex Benth.) Yakovlev (=*O. pachycarpa*) which is distinct for its double layer of columellae in the exine. But similar important as well as anomalous feature is also observed in *O. assamica* Yakovlev (=*O. robusta*), *Placolobium travancorica* (Bedd.) Yakovlev (=*O. travancorica*), *P. gracilis* (Prain) Yakovlev (=*O. gracilis*). *Fedorovia laxa* (Prain) Yakovlev (=*O. laxa*), *F. striata* (Dunn) Yakovlev (=*O. striata*) and *Ruddia fordiana* (Oliv.) Yakovlev (=*O. fordiana*). all these taxa have been given new rank in taxonomy but they do not show any distinct palynological character in support of their new taxonomic status. There are a number of overlapping characters with other species of *Ormosia*. Similarly *Vexibia alepecuroides* (Spach. ex Jaub. et Spach.) Yakovlev (=*Sophora jaubertii*) shows no additional palynological supporting character in favour of its new status except a few exceptionally smaller pollen grain size, which I believe is more phytogeographical criterion rather than a stable and important specific character. Some exceptionally small pollen grains (20.5 \times 18 μm) were also observed in

Sophora flavescens Ait. var. *flavescens* while exceptionally large ($36.5 \times 31 \mu\text{m}$) in *Sophora wightii* Baker. A few oblate-spheroidal pollen grains have been observed in contrary to the prolate-spheroidal normal ones in *Sophora pachicarpa* Schrenk ex C. A. Mayer. Some new species and variety of *Sophora* created by different authors from time to time have overlapping palynological characters with other species of the genus (Mitra *et al.*, 1979; Mitra & Mondal, 1982) published a detail pollen morphology of the tribe *Hedysareae* and *Desmodium*. Pollen morphologically *Sophora Ormosia* (some spp.) and *Calpurnia* show similarity with those of the tribe *Dalbergieae* and genera *Dalhousia* and *Ormosia* (some spp.) with the tribe *Hedysareae* and *Desmodieae* (sensu Polhill, 1981). Pollen grains of the tribe *Amherstiae* and *Detarieae* of the subfamily *Caesalpinoideae* (Okolo & Gill, 1987) show some similarity in shape, aperture and exine character. Bentham and Hooker's placement of the tribe *Sophoreae* as last tribe of *Papilionoideae* and just before *Caesalpinoideae* is justified in this regard. Similarity of pollen morphological features of the tribes of *Leguminosae* (Vishnu—Mitre & Sharma, 1962; Mitra *et al.*, 1969; Mitra & Mondal, 1982; Okolo & Gill, 1989) with *Connaraceae* (Dickison, 1979; Mondal, 1983, 1986) strengthen their possible common ancestry (Takhtajan, 1966, 1969, 1973).

Conclusion

Sophoreae is a stenopalynous tribe and supports natural grouping in Bentham and Hooker's (1862-1883) system of classification.

The taxa show a gradual trend from unspecialised to derived types of pollen morphoforms.

Creation of new genera is not supported palynologically.

Present study supports the recent phylogenetic scheme proposed by Polhill (1981), though through some bidirectional way. Palynologically *Sophoreae* links with *Dalbergieae* on one hand and *Hedysareae* (s.l.) on the other.

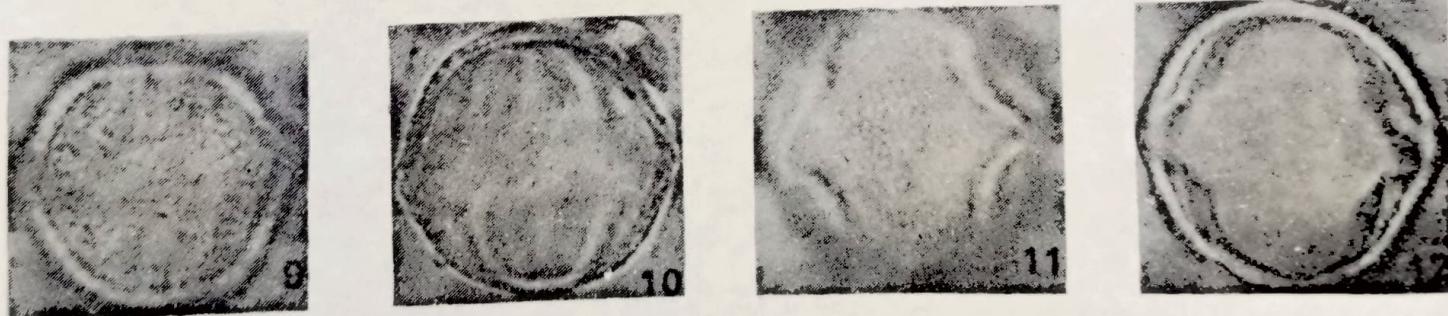
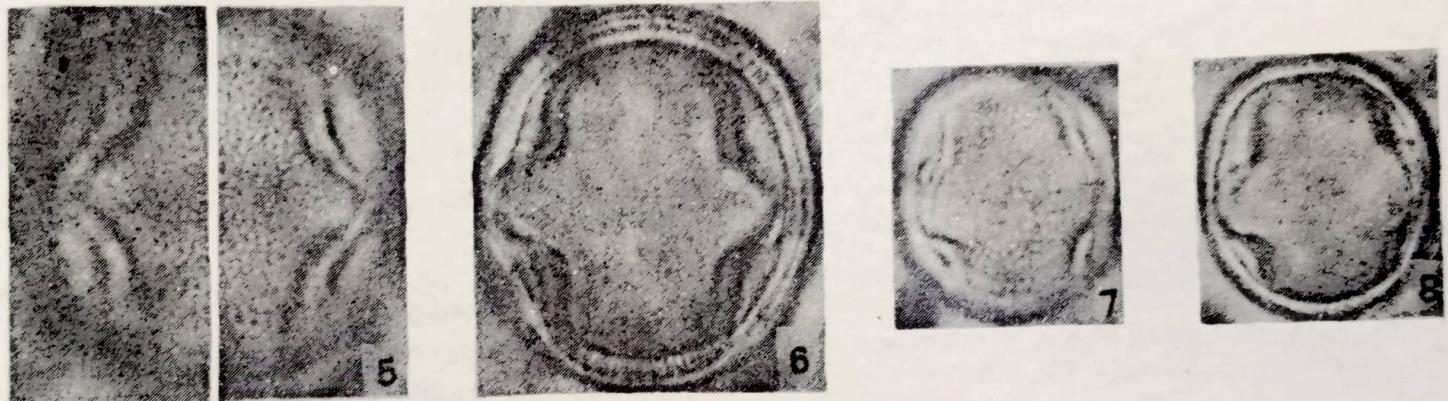
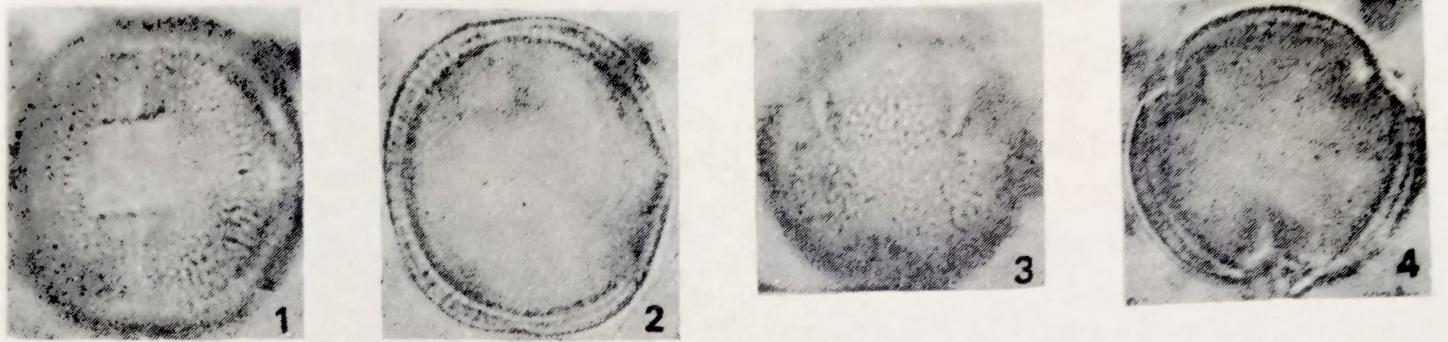
A detail SEM study is needed on the ontogeny of the pollen wall stratification of *Ormosia* for its unique development of double layer of columellae.

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Explanation of Plate

Plate 1

(All figures $\times 1000$)

- 1—4. *Trichocymus pachycarpa* (Camp. ex Benth.) Yakovlev. 1. Aperture and details of exine; 2. Optical section meridional showing double layer of columellae; 3. Polar view showing exine in apocolpium; 4. Optical section equatorial.
- 5—6. *Ormosia assamica* Yakovlev—5. Exine in 1st and 2nd focus; 6. Optical section meridional showing double layer of columellae.
- 7—8. *Sophora velutina* Lindl. var. *velutina*. 7. Details of exine; 8. Optical section meridional and aperture profile.
- 9—10. *Calpurnia auria* Baker. 9. Aperture and details of exine; 10. Optical section meridional.
- 11—12. *Dalhousia africana* S. Moore. 11. Details of exine; 12. Optical section meridional and aperture profile.