PALYNOLOGICAL INVESTIGATION OF A BORE CORE NEAR RATARIA, SOUTHERN KUTCH, GUJARAT

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ABSTRACT

Palynological fossils recovered from the bore core no. 27 drilled near Rataria in the district of Kutch, Gujarat have been presented here. The assemblage comprises microplanktons, fungal spores and microthyriaceous bodies, pteridophytic spores and gymnospermous and angiospermous pollen. Of 60 dispersed genera and 66 species, nine genera and seven identifiable species belong to algae and microplanktons, seven genera and seven species to fungi, 17 genera and 20 species to pteridophytes, three genera and two species to gymnosperms and 24 genera and 30 species to angiosperms. The pteridophytic spores (48%) are dominant followed by angiospermous pollen (35%). The following seven species are mostly encountered in the percentage count: Striatriletes susannae, S. multicostatus, Cheilanthoidspora enigmata, Polypodiaceaesporites strictus, Palmaepollenites kutchensis, Couperipollis kutchensis and Oligospharidium complex. The samples studied from this bore core have been dated as early Eocene by the Directorate of Geology & Mining, Gujarat while others have assigned Miocene age to the adjacent sedimentary exposures. The palynological assemblage has been compared with the known Lower Eocene, Middle Eocene, Oligocene and Miocene assemblages of Kutch and it has been concluded that the age of the assemblage should be in between middle Eocene and late Eocene.

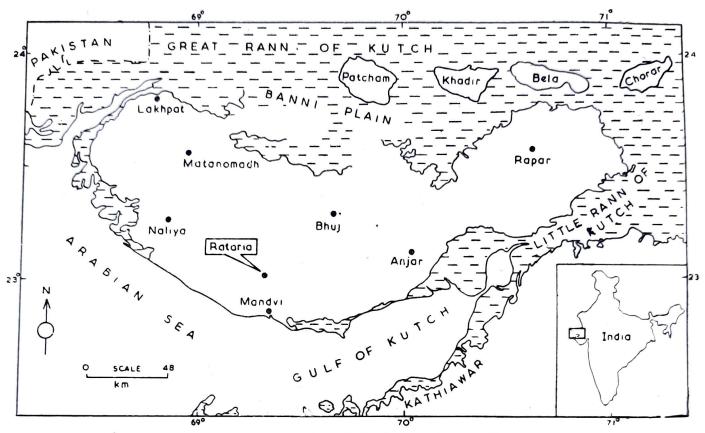
INTRODUCTION

Tertiary sediments occupy a large area of Kutch District of Gujarat, being well exposed in its southern and western parts. The earliest account on the geology of Kutch was published by Barnes (1834) which was followed by the publication of a geological map of Kutch by Grant (1840). Wynne (1872) made a detailed study of the geology of Kutch and published a map of this area; he divided the Tertiary succession into six units, viz. (i) Subnummulitic Group, (ii) Gypseous Shale, (iii) Nummulitic Group, (iv) Arenaceous Group, (v) Argillaceous Group and (vi) Upper Tertiary. Blanford (1878), Nuttal (1926), Vredenburg (1925), Tewari (1952, 1957), Nagappa (1959), Poddar (1956, 1963) and Sen Gupta (1964) have also contributed to the Tertiary stratigraphy and palaeontology of Kutch.

BISWAS (1965) instituted a new chronostratigraphic classification for the Tertiary sediments of Kutch and divided them into Madh, Berwali (Kakdi and Babia stages), Lakhpat, Khari (Waior and Khari stages) and Kankawati Series. BISWAS AND DESHPANDE (1970) revised this classification by changing the name and age of some units. They also published geological and tectonic maps of Kutch. BISWAS AND RAJU (1971, 1973) proposed a new lithestratigraphic classification of the Tertiary sequence of Kutch on the basis of the lithological characteristics and mappability of the proposed units.

The palynological studies on the Tertiary of Kutch were started, rather late, when Mathur (1963) described spores, pollen grains and microplanktons from the Gypseous shale (=Naredi Formation—Lower Eocene). This was followed by another paper by Mathur (1966) which deals with the palynoflora from the Supratrappeans (=Matanomadh Formation—Palaeocene). Recently, the palynology of this formation has been published by Kar and Saxena (1976) and Saxena (1978, 1979a, 1979b, 1980).

The palynofloral contributions to the Naredi Formation, beside that by Mathur (1963), are by Venkatachala and Kar (1968, 1969a, 1969b), Sah and Kar (1969, 1970) and Kar (1978). The Harudi Formation (Middle Eocene) palynoflora has been published by Kar (1978). The Maniyara Fort Formation (Oligocene) has been worked out by Chandra and Chatterjee (1973) and Kar (1977, 1979). The Sandhan Formation (Pliocene) has been palynologically investigated by Mathur and Mathur (1969).



Map 1. Location of the bore hole no. 27.

The present paper deals with the palynofloral investigation of the sediments encountered in bore core no. 27 near Rataria (Lat. 23°01': Long. 69°19') in southern Kutch drilled by the Directorate of Geology & Mining, Government of Gujarat (Map-1).

The geology of the area around Rataria (cf. Chota Ruttria) was briefly recorded by Wynne (1872, pp. 282-283). He described the lithology of traps and overlying sediments exposed in this area without commenting on their age and stratigraphic position. He, however, remarked that the rocks succeeding the laterite (altered traps) are most heterogeneously assembled group of lenticularly and falsely bedded, coarse, white and conglomeratic sandstones with large grains of white quartz and pale, purple, unctuous mottled, aluminous layers and lenticular masses, quite similar in character to the usual volcanic ashy beds associated with the laterites. The Directorate of Geology & Mining assigned an early Eocene age to the bore core sequence. But in the geological map published by Biswas and Deshpande (1970) the rock exposed around Rataria was shown under Vinjhan Stage (=Vinjhan Shale: Burdigalian).

MATERIAL

The base of the bore core no. 27 is constituted by altered traps (3.06m) and lithomargic clay (4.58m), followed by about 11 metre thick shale comprising carbonaceous shale (3.12m), grey shale (3.59m), carbonaceous shale (4.50m) and a band (0.01m) of siderite. Eleven samples (sample nos. 1-11) were collected from these sedi-

ments. Of these, sample no. 1 (36.8m) proved barren while rest yielded palynological fossils. Sample no. 2 was collected at the depth of 36 m, rest of them at the interval of 1m each, from bottom to top. Overlying sediments constitute 24.5 metre thick bed of variegated clay succeeded by 1.2 metre thick soil cover (Fig. 1). The slides and negatives were deposited in the repository of the Birbal Sahni Institute of Palaeobotany, Lucknow.

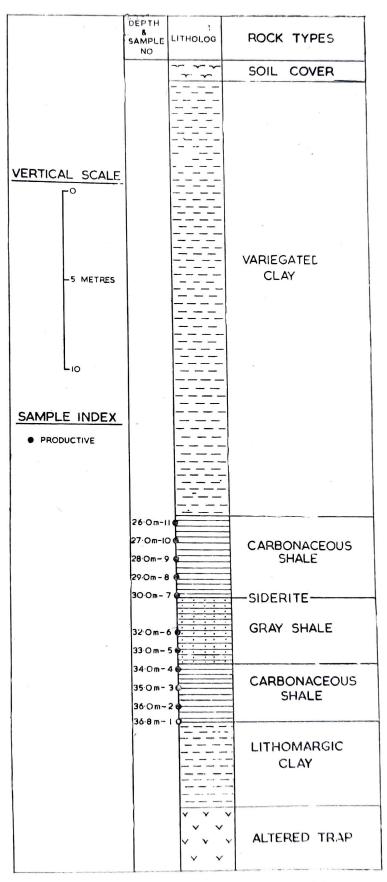


Fig. 1. Lithology of the core in bore hole no. 27 and the position of the samples studied.

The following spore-pollen species are present in the assemblage:

*Cyathidites australis Couper, 1953 (Pl. 1, Fig. 1), Biretisporites convexus Sah & Kar, 1969 (Pl. 1, Fig. 2), *Todisporites kutchensis Sah & Kar, 1969 (Pl. 1, Fig. 3), Intrapunctisporis apunctis Krutzsch, 1959 (Pl. 1, Fig. 4), *Intrapunctisporis sp., Stereisporites assamensis Dutta & Sah, 1970 (Pl. 1, Fig. 11), Lygodiumsporites lakiensis Sah & Kar, 1969 (Pl. 1, Fig. 6), Dandotiaspora plicata (Sah & Kar) Sah, Kar & Singh, 1971, *Dandotiaspora verrucata sp. nov., *cf. Osmundacidites sp., *Lophotriletes tertiarus sp. nov., *Foveosporites splendus sp. nov., *Foveosporites sp., Striatriletes susannae van der Hammen emend. Kar, 1979, (Pl. 1, Fig. 14), *Striatriletes multicostatus sp. nov., *Striatriletes microverrucosus sp. nov., *Striatriletes sp., Cheilanthoidspora enigmata Sah & Kar, 1974 (Pl. 2, Fig. 22), Cheilanthoidspora monoleta Sah & Kar, 1974 (Pl. 2, Fig. 23), *Tetrad type-1, Laevigatosporites lakiensis Sah & Kar, 1969 (Pl. 2, Fig. 25), *Biswasiaspora baculata sp. nov., *Biswasiaspora pseudoreticulata sp. nov., *Polypodiac aesporites strictus sp. nov., *Polypodiisporites sp., Seniasporites verrucosus Sah & Kar, 1969 (Pl. 2, Fig. 33), *Callialasporites trilobatus (Balme) Dev, 1961 (Pl. 2, Fig. 34), *Parasaccites sp., *Podocarpidites khasiensis Dutta & Sah, 1970 (Pl. 2, Fig. 37), Palmaepollenites kutchensis Venkatachala & Kar, 1969a, Palmaepollenites nadhamunii Venkatachala & Kar, 1969a, *Liliacidites reticulatus Sah & Kar, 1974 (Pl. 2, Fig. 36), *Palmidites naviculus sp. nov., Arecipites bellus Sah & Kar, 1970 (Pl. 2, Fig. 40), *Arecipites intrapunctatus sp. nov., *Arecipites sp., Couperipollis kutchensis Venkatachala & Kar, 1969a (Pl. 3, Fig. 44), *Psiloschizosporis psilata sp. nov., *Psiloschizosporis punctata sp. nov., Proxapertites microreticulatus Jain, Kar & Sah, 1973 (Pl. 3, Fig. 50), *Assamialetes reticulatus sp. nov., Tricolpites reticulatus Cookson, 1947 (Pl. 3, Fig. 55), Tricolpites crassireticulatus Dutta & Sah, 1970 (Pl. 3, Fig. 56), Tricolpites retibaculatus Saxena, 1979a, (Pl. 3, Fig. 63), *Retitrescolpites robustus sp. nov., Marginipollis kutchensis (Venkatachala & Kar) Kar, 1979, *Trisyncolpites sp., Umbelliferoipollenites ovatus Venkatachala & Kar, 1969a (Pl. 3 Fig. 47), Cupuliferoipollenites ovatus Venkatachala & Kar. 1969a (Pl. 3, Fig. 62), Paleosantalaceaepites primitiva Biswas, 1962 (Pl. 3, Fig. 57), Symplocoipollenites kutchensis Venkatachala & Kar, 1969a (Pl. 3, Fig. 61), Araliaceoipollenites matanamadhensis Venkatachala & Kar, 1969a (Pl. 3, Fig. 51), Striacolporites ovatus Sah & Kar, 1970 (Pl. 3, Fig. 64), Striacolporites cephalus Sah & Kar, 1970 (Pl. 3, Fig. 65), Pellicieroipollis langenheimii Sah & Kar, 1970 (Pl. 3, Fig. 66), Lakiapollis ovatus Venkatachala & Kar, 1969a (Pl. 4, Fig. 67), Lakiapollis matanamadhensis Venkatachala & Kar, 1969a (Pl. 4, Fig. 68), *cf. Triorites bellus Sah & Kar, 1970 (Pl 4, Fig. 69), *Myricipites vulgaris Dutta & Sah, 1970 (Pl 4, Fig. 70), *Myricipites globatus sp. nov., *Myricipites sp., *Semitectatriporites ratariaensis sp. nov., *Malvacearumpollis rudis Kar, 1969 (Pl. 4, Fig. 76), Phragmothyrites eocaenica Edwards emend. Kar & Saxena, 1976, *Parmathyrites sp., *Kutchiathyrites Kar, 1979, Notothyrites amorphus Kar & Saxena, 1976, Inapertisporites kedvesii Elsik, 1968, Dyadosporonites constrictus Kar, 1979, Lacrimasporonites longus Kar, 1979, *Tetraporina sp., *Hystrichosphaeridium tubiferum (Ehrenberg) Davey & Williams, 1966 (Pl. 4, Fig. 78), *Oligosphaeridium complex (White) Davey & Williams, 1966 (Pl. 4, Fig. 79), *cf. Perisselasphaeridium pannosum Davey & Williams, 1966 (Pl. 4, Fig. 80), *Cordosphaeridium gracilis Eisenack emend. Davey & Williams, 1966 (Pl. 4, Fig. 82), *Cleistosphaeridium heteracanthum (Deflandre & Cookson) Davey, Downie, Sarjeant & Williams, 1966 (Pl. 4, Fig. 83), *Hystrichokolpoma eisenacki Williams & Downie, 1966 (Pl. 4, Fig. 84), Cryptosphaera valvata Sah & Kar, 1974 (Pl. 4, Fig. 85), Cornplanktona fracta Sah & Kar, 1974.

106

The asterisk marks indicate that the species have either been described or commented upon here. Plate and figure numbers given in the above list in parenthesis denote the illustration of the present paper.

Cyathidites australis Couper, 1953 Pl. 1, Fig. 1

Remarks—The spores studied here are subtriangular with ±convex inter-apical margins while those described by Dettmann (1963) from the Mesozoic sediments of southeastern Australia have, on the contrary, concave inter-apical margins.

Todisporites kutchensis Sah & Kar, 1969

Pl. 1, Fig. 3

Remarks—Specimen assigned to this species is bigger $(78 \times 70~\mu\text{m})$ than those described by Sah and Kar (1969) from the Eocene sediments of Kutch. The trilete rays are, however, ill-developed, and do not extend more than half the radius; exine is laevigate and irregularly folded.

Intrapunctisporis sp.

Pl. 1, Fig. 5

Description—Subcircular, $64\times60~\mu m$. Trilete rays distinct, extend up to three-fourths radius. Exine 1.5 μm thick, laevigate, intrapunctate.

Comparison—Intrapunctisporis apunctis Krutzsch (1959) is distinguished by its triangular shape.

Genus—Dandotiaspora Sah, Kar & Singh, 1971

Type species—Dandotiaspora dilata (Mathur) Sah, Kar & Singh, 1971

Dandotiaspora verrucata sp. nov.

Pl. 1, Figs. 7, 8

1978—Dandotiaspora dilata (Mathur) Sah, Kar & Singh, in Saxena pl. 1, fig. 11 Holotype—Pl. 1, Fig. 7; Size $67\times60~\mu\mathrm{m}$; Slide no. 6360/11.

Type Locality-Bore core no. 27, depth 36 m, Rataria, Kutch.

Diagnosis—Triangular-subtriangular, $55-70\times50-64~\mu m$. Trilete rays distinct, extending three-fourths of the radius to margin, associated with a protuberance on distal side. Exine verrucose, verrucae being sometimes fused.

Comparison—Dandotiaspora dilata (Mathur) Sah, Kar & Singh (1971) closely resembles the present species in shape, size and presence of protuberance at ray ends, but the latter is distinguished by its verrucose ornamentation. D. dilata has three warts at the ray ends while D. verrucata has many verrucae.

cf. Osmundacidites sp.

Pl. 1, Fig. 9.

Description—Subcircular, $98 \times 77~\mu m$. Trilete not discernible. Exine about 1.5 μm thick, grana up to 1 μm high, closely placed in middle.

Genus—Lophotriletes (Naumova) Potonié & Kremp, 1954 Type species—Lophotriletes gibbosus (Ibrahim) Potonié & Kremp, 1954

Lophotriletes tertiarus sp. nov.

Pl. 1, Figs. 16, 17

Holotype—Pl. 1, Fig. 16; Size $28 \times 22 \ \mu \text{m}$; Slide no. 6370/5.

Type Locality—Bore core no. 27, depth 33 m, Rataria, Kutch.

Diagnosis—Spores triangular, $25-30\times20-27~\mu\mathrm{m}$; inter-apical margins concave. Trilete rays distinct, extending up to margin. Exine up to 2 μm thick, coni ± 1 μm high, closely placed.

Comparison—Lophotriletes rectus Bharadwaj & Salujha (1964) described from Permian of India resembles present species in shape and size range but is distinguished by its bigger coni which are about 2 μm long and are sparsely placed. The trilete rays in this species also extend up to the margin.

Genus—Foveosporites Balme, 1957 Type species—Foveosporites canalis Balme, 1957

Foveosporites splendus sp. nov.

Pl. 1, Figs. 10, 12

Holotype—Pl. 1, Fig. 12; Size $77 \times 71~\mu\mathrm{m}$; Slide no. 6351/7.

Type Locality-Bore core no. 27, depth 27 m, Rataria, Kutch.

Diagnosis—Spores triangular—subtriangular, $60-80 \times 50-75~\mu m$. Trilete extending up to three-fourths radius. Exine 1-2 μm thick; foveolae closely placed.

Comparison-Foveosporites triangulus Dutta & Sah (1970) is distinguished from the present species by its smaller size (35-48 μ m) and the foveolae are comparatively more distinct. F. pseudoreticulatus Dutta & Sah (1970) comes near to the present species in subtriangular shape but is also easily separated by its smaller size range (35-43 μm) and proximal reticulation.

Foveosporites sp.

Pl. 1, Fig. 13

Description—Triangular, $46 \times 43 \mu m$. Trilete not traceable, exine about 2 μm thick; foveolae closely placed.

Comparison—Foveosporites triangulus is bigger and possesses distinct trilete mark.

Genus—Striatriletes van der Hammen emend. Kar. 1979

Type species—Striatriletes susannae van der Hammen emend. Kar, 1979.

Striatriletes multicostatus sp. nov.

Pl. 1, Figs. 15, 18

Holotype—Pl. 1, Fig. 15; Size $78 \times 70 \mu \text{m}$; Slide no. 6358/6.

Type Locality—Bore core no. 20, depth 35m, Rataria, Kutch.

Diagnosis—Spores triangular-subtriangular in polar view, $70-80 \times 68-78 \mu m$, anisopolar, distal side distinctly convex. Apices bluntly rounded, inter-apical margins straight to convex. Trilete rays distinct extending half of radius. Exine laevigate, costate, costae 6-9, arising at ray-ends extending on distal side.

Comparison—From Striatriletes susannae the present species is distinguished by its extension of trilete rays up to half of the radius and commencement of costae at rayends.

Striatriletes microverrucosus sp. nov.

Pl. 1, Figs. 19, 20

1979-Striatriletes sp. Kar, pl. 1, figs. 16-17

Holotype—Pl. 1, Fig. 20 ; Size $80 \times 76 \ \mu \text{m}$; Slide no. 6352/11.

Type Locality-Bore core no. 27, depth 27m, Rataria, Kutch.

Diagnosis—Spores triangular-subtriangular, $68-82 \times 54-79 \mu m$. Trilete rays distinct-indistinct, extending up to two-thirds radius. Costate, microverrucose, 1-2 sets of costate generally present in inter-radial areas, others originating at ray-ends extending distally in three concentric rings.

Comparison—S. microverrucosus is distinguished from Striatriletes susannae by its microverrucose costae. S. multicostatus is also separated by its laevigate costae. Striatriletes cf. S. susannae van der Hammen emend. Kar (1979) has flattened, laevigate costae.

Striatriletes sp.

Pl. 2, Fig. 21

Description—Triangular, $52 \times 50~\mu m$, apices rounded, inter-apical margins \pm straight. Trilete rays extending half of radius. Exine 1.5 μm thick, costate, costae narrow, arising at ray-ends extending distally in three concentric rings.

Comparison—The specimen is smaller than the species described here.

Tetrad type-1

Pl. 2, Fig. 24

Description—Tetrahedral tetrad, $80 \times 78~\mu m$, three spores present, individual spore up to 60 μm , laevigate.

Remarks—Droseridites Cookson (1947), a tetrahedral tetrad, is spinose. Quadrisporites (Hennelly) Potonié & Lele (1961) is also sculptured.

Genus—Biswasiaspora gen. nov.

Type species—Biswasiaspora baculata sp. nov.

Generic Diagnosis—Spores oval-subcircular. Distinct monolete mark absent, but spores generally splitting open along longitudinal axis. Exine baculate, bacula closely or sparsely placed. Incipient inner body may be present.

Generic Description—Spores mostly oval, occasionally a few subcircular spores found, size $60-85\times50-60~\mu m$. Well marked haptotypic mark not observed, but exine generally folding longitudinally simulating monolete mark. Sometimes exine also folded to provide a pseudotrilete mark and minutely folded to look wrinkled. Exine thin, not more than $2~\mu m$ in most of the specimens. Bacula 2-10 μm long, 1-7 μm broad, mostly juxtaposed, rare on equatorial margin; tip of bacula straight or slightly lacerated; a few pila may also be interspersed. In some specimens bacula 2-4 μm high, while in others $6-10~\mu m$. Exine in some specimens (Pl. 2, Figs. 27, 28) splitting open partially in middle region providing an appearance of a colpus.

Comparison—Polypodiaceaesporites Thiergart (1938) is characterized by a distinct monolete mark, bean-shaped outline and generally laevigate exine. Monolites (Erdtman) Potonié (1956), Laevigatosporites Ibrahim (1933) and Luenites Bose & Kar (1967) are distinguished from Biswasiaspora proposed here by their unsculptured exine. Polypodiisporites Potonié (1934) is bean-shaped and verrucose. Verrucatosporites Pflug (1952) is also profusely verrucose. Crassimonoletes Singh, Srivastava & Roy (1964) is differentiated

by its laevigate exine and well-developed, wavy monolete mark. Foveomonoletes (van der Hammen) Mathur (1966) is foveclate and the monolete mark is also discernible in most of the specimens. Verrucososporites (Knox) Potonié (1970) and Tuberculatosporites Imgrund (1952) are verrucose and spinose, respectively. Biswasiaspora instituted here is distinguished from all the comparable genera by its oval-subcircular shape and baculate sculptural elements.

Derivation of name—After Dr. S. K. Biswas, Oil & Natural Gas Commission, Dehra Dun, for his outstanding work on the geology of Kutch.

Biswasiaspora baculata sp. nov.

Pl. 2, Figs. 26, 27

Holotype—Pl. 2, Fig. 27; Size $78 \times 64 \mu m$; Slide no. 6363/12.

Type Locality-Bore core no. 27, depth 34 m, Rataria, Kutch.

Diagnosis—Spores oval, $60\text{-}84\times52\text{-}60~\mu\text{m}$. Haptotypic mark not discernible but spores generally split open along the longitudinal axis to look like a colpus. Exine up to $2~\mu\text{m}$ thick, baculate; bacula 6-10 μm long and 3-7 μm broad, closely placed. Incipient inner body may be present in some spores.

Biswasiaspora pseudoreticulata sp. nov.

Pl. 2, Figs. 28, 29

Holotype—Pl. 2, Fig. 28; Size $77 \times 60 \ \mu m$; Slide no. 6363/12.

Type Locality-Bore core no. 27, depth 34 m, Rataria, Kutch.

Diagnosis—Spores oval, $60\text{-}72\times52\text{-}60~\mu\text{m}$. Distinct monolete absent; spores sometimes splitting longitudinally. Exine sometimes wrinkled, up to $2~\mu\text{m}$ thick, baculate; bacula $2\text{-}4~\mu\text{m}$ high, closely placed providing pseudoreticulate appearance in surface view.

Comparison—In B. baculata, the bacula are 6-10 μ m long whereas in B. pseudoreticulata, the bacula are only 2-4 μ m long.

Genus-Polypodiaceaesporites Thiergart, 1938

Type species-Polypodiaceaesporites haardti Thiergart, 1938.

Polypodiaceaesporites strictus sp. nov.

Pl. 2, Figs. 30, 31

Holotype—Pl. 2, Fig. 30; Size $70 \times 44 \mu m$; Slide no. 6366/14.

Type Locality-Bore core no. 27, depth 34 m, Rataria, Kutch.

Diagnosis—Spores oval, $55-70\times40-51~\mu\mathrm{m}$. Monolete distinct-indistinct, extending up to three-fourths radius. Exine 1-2 $\mu\mathrm{m}$ thick, laevigate; intrastructure appearing to be intrabaculate.

Comparison—Polypodiaceaesporites tertiarus Sah & Dutta (1966) is distinguished by its scabrate exine. P. levis Sah (1967) is bean-shaped and P. chatterjii Kar (1979) is devoid of any intrastructure.

Polypodiisporites sp.

Pl. 2, Fig. 32

Description—Spore bean-shaped, $59 \times 38~\mu m$, ends unequally broad. Monolete distinct, extending up to three-fourths along longitudinal axis. Verrucae 2-4 μm high, closely placed, giving false reticulate appearance on surface view.

Comparison—Polypodiisporites constrictus Kar (1979) from the Oligocene of Kutch resembles this specimen by its close sculptural elements, but the latter is bigger and is not constricted in middle. *P. repandus* Takahashi (1964) and *P. mawkmaensis* Dutta & Sah (1970) have robustly built, sparsely placed verrucae.

Callialasporites trilobatus (Balme) Dev, 1961 Pl. 2, Fig. 34

Remarks—Callialasporites—a typical Mesozoic genus, is found in abundance in the Mesozoic sediments of Kutch (Singh, Srivastava & Roy, 1964; Venkatachala & Kar 1969c, 1969d, 1972; Venkatachala, Kar & Raza 1969a, 1969b). In the material studied here, only a solitary specimen was obtained. It may be mentioned here that Sah and Kar (1970) also reported this species from the Lower Eocene sediments around Jhulrai, Baranda and Panandhro in the district of Kutch. The pollen grain seems to be reworked as in the adjacent areas there are Mesozoic rocks. It is, however, surprising that only this genus is frequently encountered in the Lower Eocene sediments while the other characteristic Mesozoic genera are absent.

Parasaccites sp.

Pl. 2, Fig. 35

Remarks—Only a single specimen has been recovered. This is a reworked Permian pollen grain. Such monosaccate pollen grains have been earlier reported by Venkata-chala and Kar (1969e) and Kar (pers. obs.) from the other Tertiary horizons of India.

Podocarpidites khasiensis Dutta & Sah, 1970

Pl. 2, Fig. 37

Remarks—The species is represented by a single specimen in the assemblage. It is $78 \times 60 \ \mu m$ and is smaller in size than those described by DUTTA AND SAH (1970) from the Cherra Sandstone of Assam. The diploxylonoid nature is marked and the central body is distinct.

Liliacidites reticulatus Sah & Kar, 1974

Pl. 2, Fig. 36

Remarks—The specimens studied here have more or less same size range and shape, but the mesh-size is more or less uniform.

The extant pollen grains of *Metroxylon salomonense* illustrated by Thanikaimoni (1970, pl. 21, fig. 423) resemble this species in general organisation and shape but possess thicker exine and coarser reticulation.

Genus—Palmidites Chitaley ex Couper, 1953

Type species—Palmidites maximus Couper, 1953

Palmidites naviculus sp. nov.

Pl. 2, Figs. 38, 39

Holotype—Pl. 2, Fig. 38; Size $110 \times 56 \mu \text{m}$; Slide no. 6358/11.

Type Locality—Bore core no. 27, depth 35 m, Rataria, Kutch.

Diagnosis—Pollen grains monocolpate, one end broader than other, $80-110 \times 40-56 \mu m$, colpus well-developed, extending from end to end, broader at one side. Exine 1-2 μm , thick, laevigate.

Comparison—Palmidites plicatus Singh (in SAH & SINGH, 1974) described from the Tura Formation of Assam approximates the present species in size and psilate exine but has associated folds along the colpus.

Genus—Arecipites Wodehouse, 1933

Type species—Arecipites punctatus Wodehouse, 1933

Arecipites intrapunctatus sp. nov.

Pl. 3, Figs. 41, 42

Holotype—Pl. 3, Fig. 41; Size $70 \times 47 \mu m$; Slide no. 6369/1.

Type Locality—Bore core no. 27, depth 34 m, Rataria, Kutch.

Diagnosis—Pollen grains oval, $62-75 \times 40-51 \ \mu m$. Monocolpate, colpus end to end, distinct-indistinct. Exine 1-2 μm thick, intrapunctate.

Comparison—Arecipites bellus Sah & Kar (1970) comes close to this species in shape and size range but the latter is separated by its intrapunctate ornamentation. A. bellus is punctate and the puncta are uniformly distributed.

Arecipites sp.

Pl. 3, Fig. 43

Description—Pollen grain monocolpate, $97 \times 60 \mu m$, colpus distinct, extends from one end to other. Exine about 2 μm thick, closely punctate.

Comparison—A. bellus Sah & Kar (1970) is much smaller in size (58-66 \times 48-55 μ m) than the present species. A. intrapunctatus has intrapunctate structure.

Genus—Psiloschizosporis Jain, 1968

Type species—Psiloschizosporis cacheutensis Jain, 1968

Psiloschizosporis psilata sp. nov.

Pl. 3, Figs. 45, 46

Holotype—Pl. 3, Fig. 46; Size $87 \times 50 \mu m$; Slide no. 6369/6.

Type Locality—Bore core no. 27, depth 27 m, Rataria, Kutch.

Diagnosis—Pollen grains oval, $64-100 \times 40-60 \ \mu m$. Exine 1-2 μm thick, laevigate. Furrow distinct, often splits pollen grains in two equal halves.

Comparison—Psiloschizosporis cacheutensis Jain (1968) is similar to the present species in size range and laevigate exine; however, the former is distinguished by its circular-subcircular shape. P. psilata proposed here is oval in shape.

Psiloschizosporis punctata sp. nov.

P1. 3, Figs. 48, 49

Holotype—Pl. 3, Fig. 48; Size $104 \times 48 \mu m$; Slide no. 6355/2.

Type Locality—Bore core no. 27, depth 26 m, Rataria, Kutch.

Diagnosis—Pollen grains oval-elliptical, $80\text{-}110\times45\text{-}62~\mu\mathrm{m}$. Exine up to $2~\mu\mathrm{m}$ thick, punctate, puncta closely or sparsely placed. Furrow distinct, pollen grains sometimes divide into two equal parts.

Comparison—Psiloschizosporis cacheutensis Jain (1968) is laevigate whereas the species proposed here has characteristic punctate ornamentation.

Genus—Assamialetes Singh, 1975

Type species-Assamialetes emendatus (Sah & Dutta) Singh, 1975

Assamialetes reticulatus sp. nov.

Pl. 3, Figs. 53, 54

Holotype—Pl. 3, Fig. 53; Size $48 \times 46 \mu m$; Slide no. 6364/16.

Type Locality-Bore core no. 27, depth 34 m, Rataria, Kutch.

Diagnosis—Pollen grains subcircular, $40-52 \times 36-50 \mu m$, sometimes split equatorially in two halves, reticulate, meshes may be bigger in size in equatorial and smaller in middle region.

Comparison—Assamialetes emendatus (Sah & Dutta) Singh (1975) has bigger and uniform reticulation. In A. dubius (Sah & Dutta) Singh (1975), the reticulation is imperfect. The present species is distinguished from all the known species by its two types of meshes.

Genus—Retitrescolpites Sah, 1967

Type species-Retitrescolpites typicus Sah, 1967

Retitrescolpites robustus sp. nov.

Pl. 3, Figs. 59, 60

Holotype—Pl. 3, Fig. 60; Size $62 \times 60 \mu m$; Slide no. 6364/17.

Type Locality-Bore core no. 27, depth 34 m, Rataria, Kutch.

Diagnosis—Pollen grains subcircular in equatorial view, $53-70 \times 49-68~\mu m$. Tricolpate, colpi narrow, funnel-shaped. Exine baculate-pilate; sculptural elements robustly built, closely placed forming negative reticulum in surface view.

Comparison—Retitrescolpites africanus Sah (1967) is distinguished by circular shape; R. minor Dutta & Sah (1970) is smaller in size-range (22-28 μ m); R. assamicus Dutta & Sah (1970) has not so strongly built sculptural elements.

Trisyncolpites sp.

Pl 3, Fig 52

Description—Pollen grain subcircular, $30 \times 28 \mu m$. Trisynmargocolpate; colpi uniformly broad, joined together to provide the appearance of a triradiate ridge. Exine granulose-pilate.

Comparison—Trisyncolpites ramanujamii Kar (1979) is separated by its bigger size-range.

cf. Triorites bellus Sah & Kar, 1970

Pl. 4, Fig. 69

Remarks—Only one specimen could be recovered. Two ora are distinct, one seems to be abortive, but these are not placed in the apices as in T. bellus Sah & Kar (1970). The exine is granulose.

Genus—Myricipites Wodehouse, 1933

Type species—Myricipites dubius Wodehouse, 1933

Myricipites vulgaris Dutta & Sah, 1970

Pl. 4, Fig. 70

Remarks—The specimens are bigger in size (40-50 μ m) than those described by Dutta and Sah (1970) from the Tertiary sediments of Assam. The pores are strongly

113

built with thickened outer margin. The granulose ornamentation is discernible in some specimens.

Myricipites globatus sp. nov.

Pl. 4, Figs. 71, 72

Holotype—Pl. 4, Fig. 72; Size $74 \times 68 \mu m$; Slide no. 6368/2.

Type Locality-Bore core no. 27, depth 34 m, Rataria, Kutch.

Diagnosis—Pollen grains subcircular, $64-78\times56-70~\mu m$. Triorate, margin thickened, interoral margin convex. Exine 1.5-2.5 μm thick, generally granulose, sometimes laevigate.

Comparison—Myricipites vulgaris Dutta & Sah (1970) compares with the present species in shape, nature of the ora and psilate exine, but is, smaller in size range (31-36 μ m) than the present species.

Remarks—According to Praglowski (1962) the sculptural elements in Myrica arise from a suprategillar protrusion and thus they do not stand on crest-like suprategillar ridges as in Betulaceae, Casuarinaceae and Corylaceae.

Myricipites sp.

Pl. 4, Fig. 73

Description—Pollen grains triangular, $68-80 \times 60-74~\mu m$. Triorate, ora not protruded. Exine 1-1.5 μm thick, laevigate, irregularly folded.

Comparison—Myricipits sp. resembles M. vulgaris Dutta & Sah (1970) in size range and shape but the specimen described here has thinner exine and nonprotruding ora. M. globatus sp. nov. is subcircular in shape and possesses convex interoral margin.

Genus—Semitectotriporites Guzmán, 1967

Type species-Semitectotriporites gratus Guzmán, 1967

Semitectotriporites ratariaensis sp. nov.

Pl. 4, Figs. 74, 75

Holotype—Pl. 4, Fig. 75; Size $60 \times 58 \mu \text{m}$; Slide no. 6352/9.

Type Locality-Bore core no. 27, depth 27m, Rataria, Kutch.

Diagnosis—Pollen grains subcircular, $55-62 \times 50-60~\mu m$, triporate, pore margin thickened. Exine up to $2~\mu m$ thick, baculate, interbacular space granulose.

Comparison—Semitectotriporites gratus Guzmán (1967) compares well with the present species in shape, but is distinguished by its smaller size and semitectate nature which simulates 'a foveolate or verrucate sculpture. The present species has baculate-granulose ornamentation.

Malvacearumpollis rudis Kar, 1979

Pl. 4, Fig. 76

Remarks—This species is rarely found. The pollen grains are more or less of same size as previously described by Kar (1979) from the Oligocene sediments of Kutch. Panporate nature is discernible and the spines are strongly built with bulbous base and pointed tip.

Parmathyrites sp.

Pl. 4, Fig. 77

Description—Dimidiate ascostroma, with processes $50 \mu m$, pseudoparenchymatous, cells in middle region dark, non-ostiolate. Marginal cells spinose, spines radiating.

Comparison—Parmathyrites robustus Jain & Kar (1979) described from the Neogene sediments of Kerala comes close to the present specimen in general organisation but the former is distinguished by its larger size range and pseudoparenchymatous cells in middle region.

Kutchiathyrites eccentricus Kar, 1979

Remarks—This species was originally described by KAR (1979) from the Oligocene sediments of Kutch. Jain and KAR (1979) reported the same species from the Miocene of Kerala. In Eocene, this species is rare but is bigger in size (90 × 87 µm) than the latter two species.

INCERTAE SEDIS

Tetraporina sp.

Pl. 4, Fig. 81

Description—Spore folded, 44 µm. Tetraporate, pore distinct, exine laevigate. Remarks—Only one specimen could be recovered. Naumova (1950) first described this genus from the Carboniferous of USSR and since then it has been reported in different geological horizons.

MICROPLANKTON

Hystrichosphaeridium tubiferum (Ehrenberg) Davey & Williams, 1966

Pl. 4, Fig. 78

Description—Body subcircular, $41 \times 38 \mu m$, composed of two layers, the periphragm bearing the processes; processes tubiform, more or less of equal length with serrated tips, more than twenty; reflected tabulation not very clear; apical archaeopyle not discernible.

Oligosphaeridium complex (White) Davey & Williams, 1966

Pl. 4, Fig. 79

Description—Central body subcircular-circular, 30-40 μ m. Wall two-layered, inner endophragm and outer periphragm which bears processes. Processes hollow, cylindrical, branched at tips, 20-30 μ m long, more or less of same length; cingular processes absent; number of processes not more than 18 in intact specimens.

cf. Perisselasphaeridium pannosum Davey & Williams, 1966

Pl. 4, Fig. 80

Remarks—Only a single specimen could be recovered. The central body is subcircular, $50 \times 48 \ \mu m$, made up of two layers; the inner endophragm and the outer periphragm, the latter bears the processes. The processes are of two kinds—one is broad open and tubular and the other is slender, closed processes. The reflected tabulation is not distinct and the apical archaeopyle not detectable.

Cordosphaeridium gracilis Eisenack emend. Davey & Williams, 1966

Pl. 4, Fig. 82

Description—Central body subcircular, $54 \times 45 \mu m$, two layered, the outer layer bearing processes; processes few, more or less of same size, cylindrical, hollow, branching

115

at tips with characteristic Y-shaped pattern. Archaeopyle apical, made up of one plate; partially ruptured plate seen in the figured specimen.

Remarks—According to Davey and Williams (1966), the specimens assignable to this species from the London Clay have the size range of the central body 50-71 μ m. The specimen figured here is 54 μ m in size.

Cleistosphaeridium heteracanthum (Deflandre & Cookson) Davey, Downie, Sarjeant & Williams, 1966

Pl. 4, Fig. 83

Remarks—The central body is subcircular, folded, 50 μ m. The periphragm bears many, slender, small processes of various shapes. Besides, one large, broadly rectangular process is also clearly visible. Davey et al. (1966) think that this big process is apical in position and the archaeopyle, when developed, is also apical in position. By the nature of the processes, it seems that there are more than one process per plate.

Hystrichokolpoma eisenacki Williams & Downie, 1966

Pl. 4, Fig. 84

Description—Central body not clearly distinguishable due to robustly built processes. Periphragm thin bearing two kinds of processes—the larger ones with very broad bases and slightly tapering ends and the other slender and small. Antapical process slightly bigger than the rest.

DISCUSSION

The palynological fossils reported here belong to algal cysts and other bodies, fungal spores, hyphae and microthyriaceous ascostromata, pteridophytic spores, gymnospermous and angiospermous pollen grains. In total, 60 dispersed genera comprising 66 species have been recovered. Of them nine genera and seven species belong to algae and seven genera and seven species to fungi. Pteridophytes are represented by 17 genera and 20 species, of which 12 genera and 15 species are of triletes and five genera and five species are of monoletes. Only three genera and two identifiable species of gymnosperms could be recognized. Out of them two genera, viz. *Callialasporites* and *Parasaccites* are supposed to be reworked. Twenty-four genera and 30 species of angiospermic pollen are found in the assemblage. This is the highest generic representation amongst all the groups present in the assemblage.

Of the yielding samples, sample nos. 2, 3, 4, 5 and 10 are rich in palynological fossils, and were counted to find out the percentage of different species. The other samples, i.e. nos. 6, 7, 8, 9 and 11, are poor and hence not counted. The following 41 species have been found in the percentage count: Cyathidites australis, Biretisporites convexus, Todisporites kutchensis, Intrapunctisporis apunctis, Stereisporites assamensis, Lygodiumsporites lakiensis, Dandotiaspora plicata, Striatriletes susannae, S. multicostatus, Cheilanthoidspora enigmata, S. monoleta, Laevigatosporites lakiensis, Biswasiaspora baculata, Polypodiaceaesporites strictus, Seniasporites verrucosus, Palmaepollenites kutchensis, P. nadhamunii, Liliacidites reticulatus, Palmidites naviculus, Arecipites intrapunctatus, Couperipollis kutchensis, Proxapertites microreticulatus, Tricolpites reticulatus, T. crassireticulatus, Cupuliferoipollenites ovatus, Araliaceoipollenites matanamadhensis, Pellicirroipollis langenheimii, Lakiapollis ovatus, I. matanamadhensis, Myricipites vulgaris, M. globatus, Malvacearumpollis rudis, Phragmothyrites eocaenica, Notothyrites setiferus, Inapertisporites kedvesii, Dyadosporonites constrictus, Lacrimasporonites longus, Hystrichosphaeridium

Table 1. Showing the percentage of the species in samples studied.

Name of the species	Samp. no. 2	Samp.	Samp. no. 4	Samp.	no. 6 (few	no. 7 (few	Samp. no. 8 (few spores)	no. 9 (few	Samp. no. 10	Samp. no. 11 (few spores)	Aver- age
Cyathidites australis	3			1	v						1
Biretisporites convexus				2							0.5
Todisporites kutchensis	2			4					5		2
Intrapunctisporis											
apunctis	1			1							0.5
Stereisporites assamensis	1			2							0.5
Lygodiumsporites											
lakiensis	8	2		4					2		3
Dandotiaspora plicata	5	1	1	3							2
Striatriletes susannae		18							40		. 12
Striatriletes multicostatus									33		7
Cheilanthoidspora											
enigmata	6	1	41	3							10
Cheilanthoidspora											
monoleta			4								1
Laevigatosporites											
lakiensis	2			3							1
Biswasiaspora baculata			1								
Polypodiaceaesporites											+
strictus	12	1	1	11							5
Seniasporites verrucosus	2		-								
Palmaepollenites	_										0.5
kutchensis	3			21							_
Palmaepollenites	J			-1							5
nadhamunii	1			3							
Liliacidites reticulatus	3			3							1
Palmidites naviculus	Ü	4	1						0		0.5
Arecipites intrapunctatus		-	•						2		1
Couperipollis kutchensis	8	2	28	9					1		+
Proxapertites	Ü	-	_0	3							9.5
microreticulatus	3		3	8							
Tricolpites reticulatus	12	1	3	7							3
Tricolpites	12	•		,							4
crassireticulatus									15		3
Cupuliferoipollenites										•	3
ovatus	1										1
Pellicieroipollis											+
langenheimii				1							
Lakiapollis ovatus	2	6		8							+ 3
Lakiapollis							*				3
matanamadhensis	3		1	. 1							14
Myricipites vulgaris	1		2	1							1
Myricipites globatus	1		1	3							1
Malvacearumpollis rudis	S	1									1
Notothyrites setiferus	1										+
Inapertisporites kedvesii	15	2	16								+
Geophytology, 11(2)	•										7

117

Table 1—(Contd.)

Dyadosporonites					
constrictus	1				+
Lacrimasporonites longus	1				+
Hystrichosphaeridium					_
tubiferum	11				2
Oligosphaeridium					0
complex	45				9
Cleistosphaeridium					
heteracanthum	2	3		1	1
Cryptosphaera valvata	2				0.5

Of the 41 species which appeared in the percentage count, none of them is consistently represented in all the samples. In sample no. 2, Lygodiumsporites lakiensis (8%), Cheilanthoidspora enigmata (6%), Polypodiaceaesporites strictus (12%), Couperipollis kutchensis (8%), Tricolpites reticulatus (12%) and Inapertisporites kedvesii (15%) are found as major taxa. But in sample no. 3, Striatriletes susannae (18%), Lakiapollis ovatus (6%), Hystrichosphaeridium tubiferum (11%) and Oligosphaeridium complex (45%) are the dominant species. Cheilanthoidspora enigmata (41%), Couperipollis kutchensis (28%) and Inapertisporites kedvesii (16%) are mostly found in sample no. 4. In sample no. 5, Polypodiaceaesporites strictus (11%), Palmaepollenites kutchensis (21%), Couperipollis kutchensis (9%), Proxapertites microreticulatus (8%), Tricolpites reticulatus (7%) and Lakiapollis ovatus (8%) are commonly met with. The sample no. 10 is rich in Striatriletes susannae (40%), S. multicostatus (33%) and Tricolpites crassireticulatus (15%). However, taking the average of all the samples the following seven species are found in good percentage. Striatriletes susannae (12%), S. multicostatus (7%), Cheilanthoidspora enigmata (10%), Polypodiaceaesporites strictus (5%), Palmaepollenites kutchensis (5%), Couperipollis kutchensis (9.5%) and Oligosphaeridium complex (9%).

The pteridophytic spores are the dominant elements in the assemblage as they contribute 48 per cent to the assemblage. Of them, triletes (41%) are more common than monoletes (7%). Angiospermous pollen grains come second (35%) in spite of the fact that they have better generic and specific representation. The fungal elements are rare (4%) but the microplanktons (13%) are common.

COMPARISON WITH OTHER TERTIARY ASSEMBLAGES IN KUTCH

Naredi Formation—The Naredi Formation is represented by Triorites triangulus Cenozone with the characteristic species of Cupuliferoipollenites ovatus, Lakiapollis ovatus, Cyathidites minor, Tricolpites reticulatus, T. levis, T. brevis, Rhoipites kutchensis, Symplocoipollenites constrictus, Meliapollis ramanujamii and Pseudonothofagidites kutchensis.

The following species are common in both the assemblages: Dandotiaspora plicata, Palmaepollenites kutchensis, Proxapertites microreticulatus, Tricolpites reticulatus, Araliaceoi-pollenites matanamadhensis, Cupuliferoipollenites ovatus and Lakiapollis ovatus.

The present assemblage differs considerably from the Naredi Formation by its dominance of pteridophytic spores. The genus *Striatriletes* which is found in significant percentage is conspicuous by its absence in the Naredi Formation. Moreover, *Triorites triangulus* which is most common in Naredi Formation is not at all found in the Rataria bore core. Besides, the present material has also good percentage of microplankton.

Harudi Formation—This formation in Kutch has been dated as Middle Eocene (Lutetian) on the basis of Nummulites by Biswas and Raju (1971, 1973) and planktonic

foraminifera by Mohan and Soodan (1970). The miospore assemblage worked out by Kar (1979) is dominated by Proxapertites microreticulatus together with Palmaepollenites kutchensis, P. ovatus, Cyathidites minor, Couperipollis kutchensis, Scantigranulites sparsus, Seniasporites verrucosus, Laevigatosporites cognatus and Tricolpites levis.

The assemblage recovered from the present bore core has very insignificant percentage of *Proxapertites microreticulatus*. In fact, except *Palmaepollenites kutchensis* and *Couperipollis kutchensis* the other important taxa of both the assemblages are not common to each other. *Striatriletes* which is abundant in the present bore core is totally absent in the Harudi Formation.

Maniyara Fort Formation—This formation has been dated as Oligocene by BISWAS (1965), BISWAS AND DESHPANDE (1970) and BISWAS AND RAJU (1971). KAR (1977, 1979) recorded Polysphaeridium microtriainum Cenozone, Trisyncolpites ramanujamii Cenozone and Aplanosporites robustus Cenozone from this formation.

In sample no. 3 of the present bore core microplanktons are dominant and mostly represented by Oligosphaeridium complex and Hystrichosphaeridium tubiferum. It may be mentioned here that the small spiny varieties which are abundant in the Maniyara Fort Formation are rare in the present assemblage. On the contrary, dinoflagellate cysts with open, tubular processes are found in abundance. Trisyncolpites ramanujamii is also not found in the present assemblage and only one pollen grain, assignable to this genus, has been described as Trisyncolpites sp. The next dominant element of the Maniyara Fort Formation, viz. Aplanosporites robustus, is also absent in this assemblage. However, Striatriletes susannae, the parkeriaceous spore, is found as one of the dominant elements in both the assemblages. Besides, Inapertisporites kedvesii, Phargmothyrites eocaenica, Kutchiathyrites eccentricus, Laevigatosporites lakiensis, Dyadosporonites constrictus and Lacrimasporonites longus are also found as common species in the two assemblages.

Khari Nadi Formation—Biswas and Raju (1971) assigned Aquitanian age for this formation on the basis of fauna. No published palynological information is available. However, work done by Kar (unpublished data) shows that this formation is characterized by the dominance of pteridophytic spores, gymnospermous pollen grains and microplanktons. The pteridophytic spores are mostly represented by Striatriletes, Cyathidites and Lycopodiumsporites. The gymnospermous pollen have been previsionally identified as Podocarpidites and Tsugaepollenites. Amongst angiosperms, Malvacearumpollis is common. The microplanktons are represented by dinoflagellate cysts and the small spiny types are prolifically represented than the cysts with open, tubular processes.

The present assemblage resembles that of the Khari Nadi by its representation of Striatriletes, but the rarity of gymnospermous pollen distinguishes the Rataria assemblage from the previous one. In addition, the microplanktons of the two assemblages are also distinct from one another.

Age of the present assemblage

The Directorate of Geology and Mining has assigned early Eocene age to the bore core because the carbonaceous shale rests on the lithomargic clay and laterites. Biswas and Deshpande (1970), however, thought that in the neighbourhood there is no Eocene exposure and marked the local exposures as Vinjhan Shale, dating them Burdigalian in age.

The Lower Eccene rocks represented by Naredi Formation have been thoroughly worked out and it has already been pointed out that the present assemblage differs

Geophytology, 11(2)

considerably from the former. The palynological assemblage from Harudi Formation (middle Eocene) is also quite distinct. The Maniyara Fort Formation (Oligocene) resembles the present assemblage by the presence of *Striatriletes*. It may be mentioned here that the genus *Striatriletes*, according to the present state of knowledge, does not appear in India before Upper Eocene (Kopili Formation) and finds its maximum development during Oligocene and Miocene.

On the basis of Striatriletes, the assemblage may be put anywhere between Upper Eocene to Miocene. But the presence of Cheilanthoidspora enigmata and C. monoleta in the assemblage indicates a Lower Eocene affinity as both of them are characteristic species of Palana lignites, Rajasthan which are of early Eocene in age. Taking all these data into consideration, it seems that the assemblage described here may be of middle Eocene—late Eocene in age. The presence of Oligosphaeridium complex and Hystrichosphaeridium tubiferum in good percentage in one sample also points towards Eocene as both these specimens have been recorded from the London Clay by Davey and Williams (1966).

The palynological assemblage from the Quilon and Warkalli beds (Miocene) of Kerala worked out by RAO AND RAMANJUAM (1978) and JAIN AND KAR (1979) differs from the present one by the presence of Crassoretitriletes, Margocclporites, Ctenolophonidites, Verrutriporites and Chenopodipollis.

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EXPLANATION OF PLATES

(All figures are ca ×500)

PLATE 1

- 1. Cyathidites australis Couper, Slide no. 6351/13.
- 2. Biretisporites convexus Sah & Kar, Slide no. 6361/5.
- 3. Todisporites kutchensis Sah & Kar, Slide no. 6351/4.
- 4. Intrabunctisporis abunctis Krutzsch, Slide no. 6365/5.
- 5. Intrapunctisporis sp., Slide no. 6352/10.
- 6. Lygodiumsporites lakiensis Sah & Kar, Slide no. 6352/4.
- 7, 8. Dandotiaspora verrucata sp. nov., Slide nos. 6360/11, 6364/1.
- 9. cf. Osmundacidites sp., Slide no. 6364/6.
- 10, 12. Foveosporites splendus sp. nov., Slide nos. 6370/3, 6351/7.
- 11. Stereisporites assamensis Dutta & Sah, Slide no. 6371/6.
- 13. Foveosporites sp., Slide no. 6365/1.
- 14. Striatriletes susannae van der Hammen emend. Kar, Slide no. 6356/2.
- 15, 18. Striatriletes multicostatus sp. nov., Slide nos. 6358/6, 6357/11.
- 16, 17. Lophotriletes tertiarus sp. nov., Slide no. 6370/5.
- 19, 20. Striatriletes microverrucosus sp. nov., Slide nos. 6351/11, 6352/11.

PLATE 2

- 21. Striatriletes sp., Slide no. 6357/11.
- 22. Cheilanthoidspora enigmata Sah & Kar, Slide no. 6367/7.
- 23. Cheilanthoidspora monoleta Sah & Kar, Slide no. 6366/3.
- 24. Tetrad type-1, Slide no. 6353/2.
- 25. Laevigatosporites lakiensis Sah & Kar, Slide no. 6368/22.
- 26, 27. Biswasiaspora baculata sp. nov., Slide nos. 6364/17, 6363/12.
- 28, 29. Biswasiaspora pseudoreticulata sp. nov., Slide nos. 6363/12, 6354/1.
- 30, 31. Polypodiaceaesporites strictus sp. nov., Slide nos. 6366/14, 6359/11.
- 32. Polypodiisporites sp., Slide no. 6364/10.
- 33. Seniasporites verrucosus Sah & Kar, Slide no. 6367/6.
- 34. Callialasporites trilobatus (Balme) Dev, Slide no. 6371/2.
- 35. Parasaccites sp., Slide no. 6368/18.
- 36. Liliacidites reticulatus Sah & Kar, Slide no. 6368/4.
- 37. Podocarpidites khasiensis Dutta & Sah, Slide no. 6368/16.
- 38, 39. Palmidites naviculus sp. nov., Slide nos. 6358/11, 6366/9.
- 40. Arecipites bellus Sah & Kar, Slide no. 6364/19.

PLATE 3

- 41, 42. Arecipites intrapunctatus sp. nov., Slide nos. 6369/1, 6361/6.
- 43. Arecipites sp., Slide no. 6367/15.
- 44. Couperipollis kutchensis Venkatachala & Kar, Slide no. 6367/8.
- 45, 46. Psiloschizosporis psilata sp. nov., Slide nos. 6370/2, 6369/6.
- 47. Umbelliferoipollenites ovatus Venkatachala & Kar, Slide no. 6370/2.
- 48, 49. Psiloschizosporis punctata sp. nov., Slide nos. 6355/2, 6363/9.
- 50. Proxapertites microreticulatus Jain, Kar & Sah, Slide no. 6368/20.
- 51. Araliaceoipollenites matanamadhensis Venkatachala & Kar, Slide no. 6373/5.
- 52. Tisyncolpites sp., Slide no. 6370/7.
- 53, 54. Assamialetes reticulatus sp. nov., Slide nos. 6364/16, 6364/13.
- 55. Tricolpites reticulatus, Cookson, Slide no. 6371/11.
- 56. Tricolpites crassireticulatus Dutta & Sah, Slide no. 6351/7.
- 57. Paleosantalaceaepites primitiva Biswas, Slide no. 6362/9.
- 58. Marginipollis kutchensis (Venkatachala & Kar) Kar, Slide no. 6372/9.
- 59, 60. Retitrescolpites robustus sp. nov., Slide nos. 6364/12, 6364/17.
- 61. Symplocoipollenites kutchensis Venkatachala & Kar, Slide no. 6367/8.
- 62. Cupuliferoipollenites ovatus Venkatachala & Kar, Slide no 6372/2.
- 63. Tricolpites retibaculatus Saxena, Slide no. 6351/5.
- 64. Striacolporites ovatus Sah & Kar, Slide no. 6367/13.
- 65. Striacolporites cephalus Sah & Kar, Slide no. 6366/2.
- 66. Pellicieroipollis langenheimii Sah & Kar, Slide no. 6372/3.

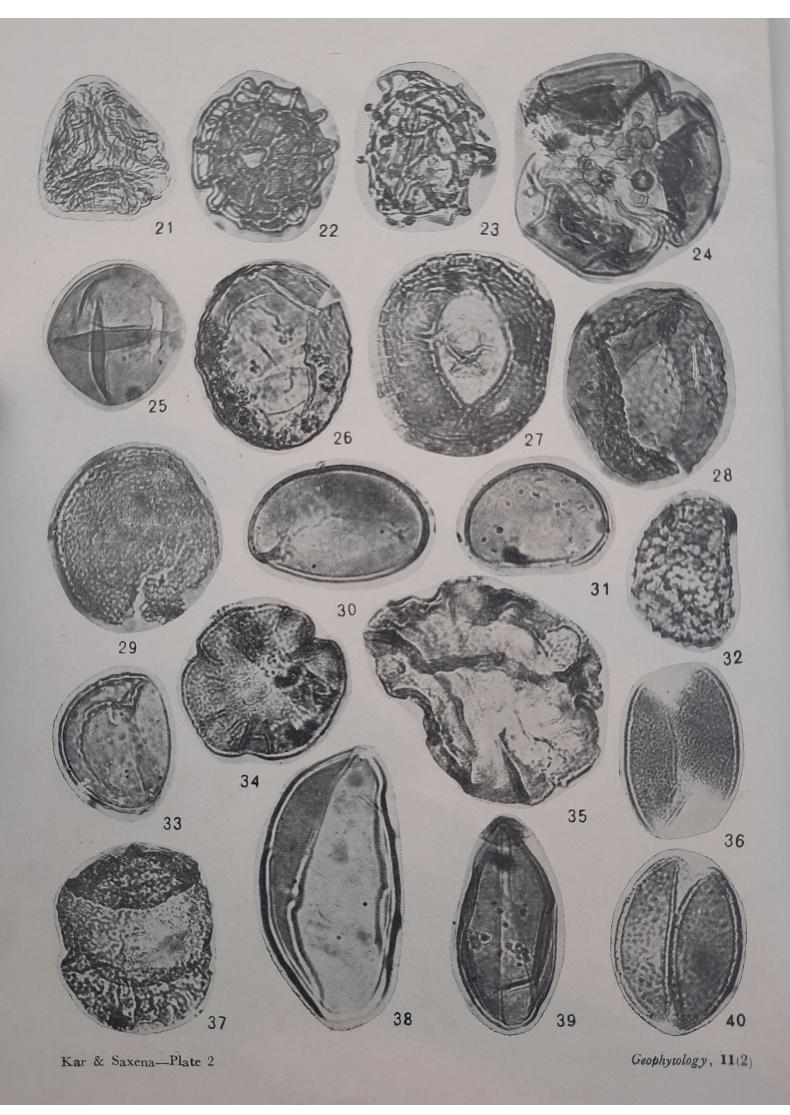
PLATE 4

- 67. Lakiapollis ovatus Venkatachala & Kar, Slide no. 6367/14.
- 68. Lakiapollis matanamadhensis Venkatachala & Kar, Slide no. 6372/5.
- 69. cf. Triorites bellus Sah & Kar, Slide no. 6373/2.
- 70. Myricipites vulgaris Dutta & Sah, Slide no. 6360/5.
- 71, 72. Myricipites globatus sp. nov., Slide nos. 6370/4, 6368/2.
- 73. Myricipites sp., Slide no. 6368/19.
- 74, 75. Semitectotriporites ratariaensis sp. nov., Slide no. 6352/8, 6352/9.
- 76. Malvacearumpollis rudis Kar, Slide no. 6351/10.
- 77. Parmathyrites sp., Slide no. 6359/9.
- 78. Hystrichosphaeridium tubiferum (Ehrenberg) Davey & Williams, Slide no. 6357/2.
- 79. Oligosphaeridium complex (White) Davey & Williams, Slide no. 6369/8.
- 80. cf. Perisselasphaeridium pannosum Davey & Williams, Slide no. 6351/14.
- 81. Tetraporina sp., Slide no. 6359/12.
- 82. Cordosphaeridium gracilis Eisenack emend. Davey & Williams, Slide no. 6366/1.
- 83. Cleistosphaeridium heteracanthum (Deflandre & Cookson) Davey, Downie, Sarjeant & Williams, Slide no. 6370/6.
- 84. Hystrichokolpoma eisenacki Williams & Davey, Slide no. 6356/6.
- 85. Cryptosphaera valvata Sah & Kar, Slide no. 6359/5.



Geophytology, 11(2)

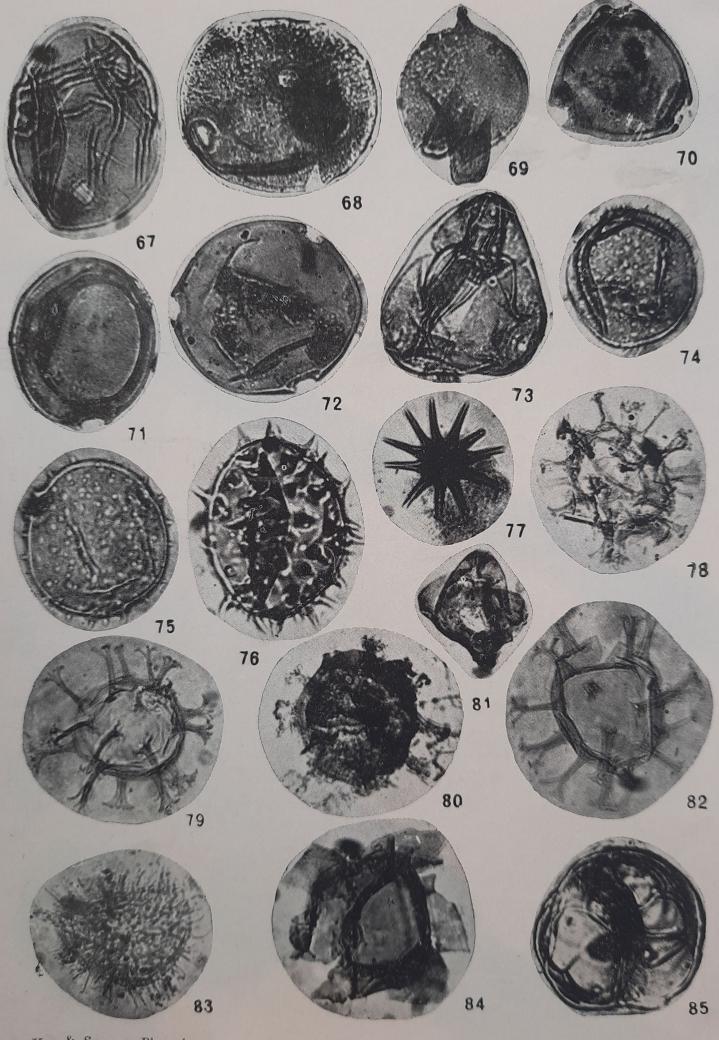
Kar & Saxena—Plate 1





Geophytology, 11(2)

Kar & Saxena—Plate 3



Kar & Saxena—Plate 4

Geophytology 11(2)